

Short Communication

Effect of Lecanemab and Donanemab in Early Alzheimer's Disease: Mechanistic Interpretation in the Amyloid Cascade Hypothesis 2.0 Perspective

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Accepted 14 April 2023

Pre-press 16 May 2023

Abstract. In clinical trials, lecanemab and donanemab showed statistically significant yet marginal slowdown of Alzheimer's disease (AD)-associated cognitive decline. This could be due to their sub-optimal design and/or deployment; alternatively, their limited efficiency could be intrinsic. Distinguishing between the two is of great importance considering the acute need of efficient AD therapy and tremendous resources being invested in its pursuit. The present study analyzes the mode of operation of lecanemab and donanemab within the framework of recently proposed Amyloid Cascade Hypothesis 2.0 and concludes that the second possibility is correct. It suggests that substantial improvement of the efficiency of these drugs in symptomatic AD is unlikely and proposes the alternative therapeutic strategy.

Keywords: Alzheimer's disease, Amyloid Cascade Hypothesis 2.0, A β PP-independent *i*A β generation in AD, intraneuronal A β , lecanemab, donanemab

Lecanemab, the recently approved drug for treatment of early stages of Alzheimer's disease (AD), exhibited statistically significant, yet marginal reduction in the rate of AD-associated cognitive decline [1]. These findings were met with great enthusiasm and the hope that the drug can be improved to arrest the progression of or even to cure the disease [2, 3].

The question is whether the drug or its utilization in the trial have been suboptimal and outcomes could be significantly improved, or if its limited efficiency in early AD is intrinsic. The present study analyzes the mode of operation of lecanemab and donanemab within the framework of the recently proposed Amyloid Cascade Hypothesis 2.0 (ACH2.0) and concludes that the latter is correct. It suggests that, in their trials, both drugs acted preventively, not curatively, on only a small neuronal subpopulation, and that a substantial improvement of their efficiency in symptomatic AD is highly unlikely and proposes the alternative therapeutic strategy.

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The initial Amyloid Cascade Hypothesis (ACH) [4] postulated that AD is caused by amyloid- β ($A\beta$) produced in the $A\beta$ protein precursor ($A\beta$ PP) proteolytic/secretory pathway and deposited extracellularly. Accordingly, two principal categories of ACH-based AD drugs are either those suppressing production and, consequently, secretion of $A\beta$ PP-derived $A\beta$ or agents sequestering or clearing extracellular $A\beta$; lecanemab belongs to the second group. In contrast, the recently proposed ACH2.0 posits that AD is a two-stage disease caused and driven by intraneuronal (rather than extracellular) $A\beta$, $iA\beta$ [5]. The first, asymptomatic, stage is a life-long accumulation of $iA\beta$, which occurs via internalization of extracellular $A\beta$ and through retention of $A\beta$ produced by the gamma-cleavage of the C99 fragment of $A\beta$ PP on intracellular, rather than on plasma, membranes [6–15] (reviewed in [5]). Upon reaching the T1 threshold in affected neurons in a narrow temporal window, $iA\beta$ triggers, presumably following the elicitation of PKR- and HRI-mediated integrated stress response, activation of the $A\beta$ PP-independent $iA\beta$ generation pathway [5]. The bulk, if not the entire output of this pathway are retained intraneuronally and perpetuate the operation of the pathway [5]. $iA\beta$ levels rapidly increase and drive a devastating cascade that includes tau pathology; when they cross the T2 threshold, neurons commit apoptosis and AD symptoms start to manifest (Fig. 1A). When a sufficient fraction of neurons is lost, AD enters the end-stage (Fig. 1B). Thus, in the ACH2.0 paradigm, the primary therapeutic target is intraneuronal, rather than extracellular, $A\beta$.

Importantly, in the framework of the ACH2.0, ACH-based AD drugs (as defined above) could be effective only preventively by reducing the rate of $A\beta$ PP-derived $iA\beta$ accumulation and delaying the T1 crossing or precluding it within the lifespan of an individual [5]. This is because the crossing of the T1 threshold enables the activation of the $A\beta$ PP-independent $iA\beta$ production pathway, which is unaffected by ACH-based drugs [5]. Conceptually, drugs suppressing the production of $A\beta$ PP-derived $A\beta$ would inhibit both components of the influx of $A\beta$ PP-derived $iA\beta$, its retention and internalization: less $A\beta$ is produced, less is retained; less $A\beta$ is secreted, its extracellular pool is smaller and less is taken up. On the other hand, agents sequestering or clearing extracellular $A\beta$ affect only one $iA\beta$ influx component: its cellular uptake; this is precisely what lecanemab does in two ways. First, it lowers internalization of extracellular $A\beta$ simply by reduc-

ing its pool. More importantly, it acts in a specific and targeted manner. Lecanemab is, in essence, a monoclonal antibody (BAN2401), which specifically sequesters “protofibril” $A\beta$, i.e., soluble extracellular $A\beta$ in oligomeric form [1]. Crucially, the latter is the intermediate in the cellular uptake of $A\beta$ [6–11]. Consequently, sequestering oligomeric $A\beta$, lecanemab specifically suppresses internalization of extracellular $A\beta$, one of the two components of the steady-state influx of $A\beta$ PP-derived $iA\beta$ in neuronal cells, and reduces the rate of its accumulation. After the T1 crossing, however, the utilization of ACH-based drugs, including lecanemab, would be futile because the activation of the $A\beta$ PP-independent $iA\beta$ production pathway would render the contribution of $A\beta$ PP-derived $A\beta$ to the $iA\beta$ pool insignificant and the $A\beta$ PP proteolytic pathway irrelevant for the progression of AD [5].

All participants of the lecanemab trial have exhibited early AD symptoms by the commencement of the treatment. By the time AD symptoms manifest, however, the bulk of the affected neurons have already crossed the T1 threshold and would be unresponsive to the drug [5]. Therefore, the only explanation of the observed effect of lecanemab [1] in the ACH2.0 perspective is that at the time of the commencement of the treatment a fraction of affected neurons had not yet reached the T1 threshold and was still responsive to the drug. It is this fraction of the neuronal population that was meaningfully targeted by and responded to the treatment with lecanemab, and positive results were marginal because of the marginal size of the neuronal fraction redeemed, although possibly only temporarily (see below), by the drug.

The presumed mode of the lecanemab’s action, described above, is illustrated in Fig. 1. Figure 1A shows the initial state of $iA\beta$ dynamics in the affected neuronal population at the time of the commencement of the treatment. The bulk of neurons have crossed the T1 threshold. Of those, a fraction have also crossed the T2 threshold and triggered the manifestation of AD symptoms; the majority has $iA\beta$ levels distributed between the T1 and T2 threshold. At this time, a minor subpopulation of affected neurons did not yet cross the T1 threshold (shown in red solely to graphically distinguish them from neurons that crossed the T1; otherwise both groups are identical). Figure 1B–D depict results of the evolution of the initial state in the absence or presence of the drug. No drug is administered in Fig. 1B. The “red” neuronal fraction crosses the T1 threshold, $A\beta$ PP-independent production of $iA\beta$ occurs in all affected

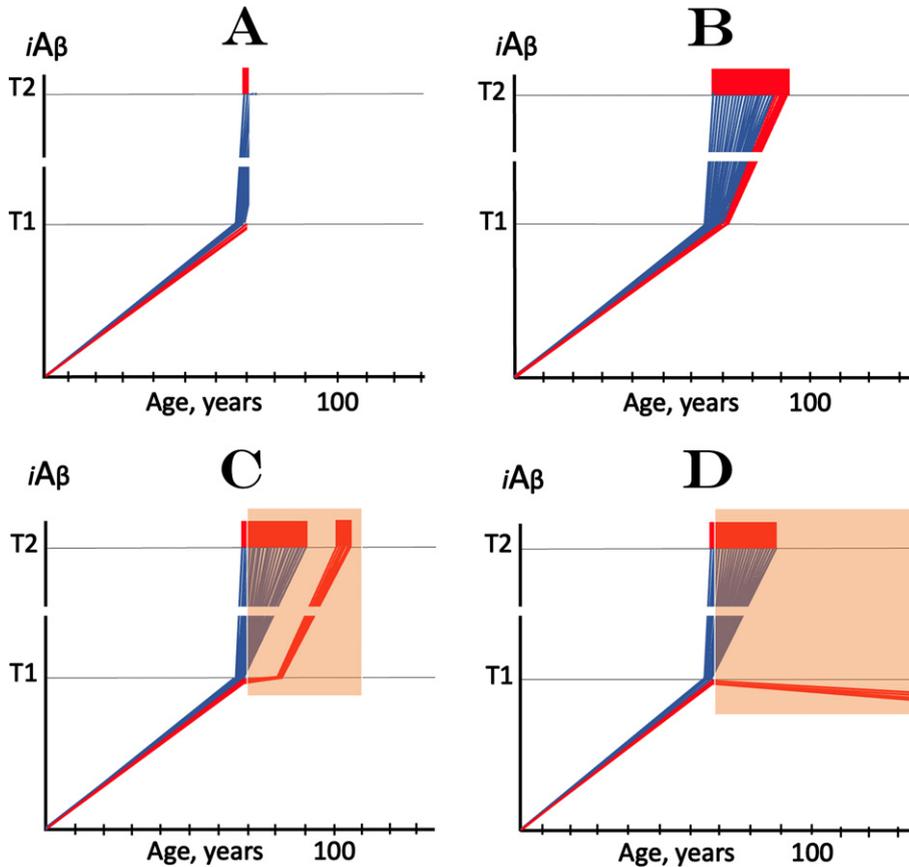


Fig. 1. Effect of lecanemab at early symptomatic AD. $iA\beta$: Level of intraneuronal $A\beta$. $T1$ threshold: Levels of $iA\beta$ triggering, plausibly via activation of PKR and HRI kinases, elicitation of the ISR and initiation of $A\beta$ PP-independent production of $iA\beta$. $T2$ threshold: Levels of $iA\beta$ triggering neurons' entrance into the apoptotic pathway. *Blue and red lines*: Individual affected neurons. *Red lines*: A fraction of affected neurons that did not reach the $T1$ threshold at the time of the commencement of lecanemab treatment. *Red blocks*: Apoptotic zone. *Orange fields*: The duration of lecanemab treatment. *A*: The "initial state" – $iA\beta$ dynamics in affected neurons at the commencement of lecanemab treatment. Note that a small neuronal fraction did not yet reach the $T1$ threshold (shown in red solely to distinguish it from the bulk of neurons that already crossed the $T1$ threshold; otherwise both fractions are identical). *B*: Result of the evolution of the initial state in the absence of a treatment. The "red" neuronal fraction reached the $T1$ threshold, $iA\beta$ levels in both neuronal fractions crossed the $T2$ threshold and AD entered the end-stage. *C, D*: Effect of lecanemab treatment in early AD. Note that neurons that crossed the $T1$ threshold by the commencement of the treatment remain unaffected by it and evolve as shown in *B*. *C*: The rate of accumulation of $A\beta$ PP-derived $iA\beta$ is reduced but its levels continue to increase. Eventually, they reach the $T1$ threshold, cross the $T2$ threshold and cells commit apoptosis. The fate of the "red" neuronal population is the same as in *B* but occurs with a delay; these neurons are redeemed by the drug but only temporarily. *D*: The treatment arrests or reverses the accumulation of $A\beta$ PP-derived $iA\beta$. Levels of $A\beta$ PP-derived $iA\beta$ do not reach the $T1$ threshold and the "red" neuronal fraction is redeemed permanently for the duration of the treatment.

neurons, and $iA\beta$ levels ascend stochastically toward the $T2$ threshold. When a sufficient portion of neurons is lost, AD enters the end-stage shown in Fig. 1B.

Figure 1C and 1D show results of the evolution of the initial state in the presence of the drug (orange fields indicate the duration of the treatment). Neurons that crossed the $T1$ threshold ("blue" neuronal fraction) are not affected by the treatment and evolve toward the same outcome as shown in Fig. 1B. For the "red" neuronal fraction, two distinct outcomes are possible. In one, shown in Fig. 1C, the rate of

accumulation of $A\beta$ PP-derived $iA\beta$ is reduced but its levels continue to increase. Eventually, they would reach and cross the $T1$ threshold, and begin ascending toward the $T2$ threshold. In this scenario, the fate of the "red" neuronal population would be the same as in Fig. 1B but will occur with a delay; neurons would be redeemed by a drug but only temporarily. The outcome shown in Fig. 1D is the arrest or reversal (due to degradation and clearance of $iA\beta$) of the accumulation of $A\beta$ PP-derived $iA\beta$ in the "red" neuronal fraction. In this scenario (Fig. 1D), levels of

A β PP-derived *iA* β would not reach the T1 threshold and the “red” neuronal fraction would be redeemed permanently for the duration of the treatment. The published results of clinical trials of lecanemab [1] do not allow to distinguish between the two outcomes described above because its participants were not yet followed for a sufficient duration.

The therapeutic outcomes in both scenarios (Fig. 1C, D) would, nevertheless, be only marginal because the size of the targeted neuronal subpopulation (“red” neuronal fraction) would be only marginal since by the time AD symptoms manifest, the bulk of the affected neurons have already crossed the T1 threshold [5]. The only way to improve the therapeutic outcome of any treatment targeting A β PP-derived A β , including lecanemab, is by advancing the diagnosis and the commencement of a treatment, thus maximizing the “red” neuronal fraction. This approach, however, is limited, hence the intrinsic limitation of lecanemab or any other A β PP-derived A β -targeting drug in the treatment of symptomatic AD. On the other hand, the present interpretation of the results of the lecanemab’s trial asserts that drugs that cause the arrest or reversal of accumulation of A β PP-derived *iA* β , or sufficient reduction of its levels, would be effective in preventing AD if their administration commences before symptomatic manifestation of the disease, more precisely before the crossing of the T1 threshold and activation of the A β PP-independent *iA* β production. It also mandates clinical trials of lecanemab in prevention of AD using asymptomatic cohorts.

Conceptually similar, both quantitatively and qualitatively, results in clinical trials for treatment of very early symptomatic AD were recently obtained with donanemab, a humanized IgG1 antibody directed at an N-terminal pyroglutamate A β epitope that is present only in established plaques [20, 21]. By sequestering extracellular amyloid-beta deposits, donanemab shifts the equilibrium of extracellular A β processing toward formation of plaques and thus reduces the levels of extracellular soluble A β . This, in turn, suppresses the rate of extracellular A β internalization and inhibits the influx of *iA* β . Therefore, the explanation of the observed effect of donanemab in early AD is identical to that of the effect of lecanemab, namely the suppression of the influx of *iA* β and, consequently, the reduction or reversal of the rate of its accumulation. Likewise, in similarity with lecanemab, the observed effect of donanemab in clinical trials [21] was marginal because, in the trial participants, the drug impacted only the marginal neuronal

subpopulations where the *iA* β levels have not yet crossed the T1 threshold and the A β PP-independent *iA* β production pathway was not yet activated.

A recent *Nature* commentary on the subject [2] posed a question: “Alzheimer’s drug slows mental decline in trial — but is it a breakthrough?” The ACH2.0 provides the unequivocally negative answer. However, it also suggests the strategy to actually achieve such a breakthrough. As expounded upon elsewhere [5], the only viable therapeutic option for symptomatic AD in the ACH2.0 framework is the reduction of the *iA* β levels to those below the T1 threshold; this would cease the operation of the AD-driving A β PP-independent *iA* β generation pathway and arrest the progression of the disease. Moreover, a sufficient depletion of *iA* β levels would substantially reset them and force the resumption of its accumulation (only in the A β PP proteolytic pathway) from a low baseline. This opens an attractive possibility of a transient, once-in-a-lifetime-only treatment of symptomatic AD [5]. Indeed, sufficiently depleted A β PP-derived *iA* β levels would not reach the T1 threshold within the lifespan of an AD patient. Such a therapy would be effective in symptomatic AD because it would potentially redeem all neurons that did not yet cross the T2 threshold and commit apoptosis. Even more attractively, the same strategy could be applied preventively prior to the manifestation of AD symptoms. Any agent capable of targeted degradation of *iA* β and its sufficient depletion within a short duration would potentially be appropriate for enacting the above strategy. Two apparently suitable physiologically operating activities are actually built into the two familiar actors in the AD play: BACE1 and BACE2. Both are capable of multiple cleavages within *iA* β (reviewed in [5]), a capacity enhanced in BACE1 by the Icelandic A β PP mutation [16, 17] (explaining its protective action) and suppressed in BACE2 by the Flemish A β PP mutation [18] (thus causing familial AD). Activators of physiologically occurring *iA* β -cleaving capabilities of BACE1 and/or BACE2 could potentially constitute potent AD drugs [5].

Application of the transient *iA* β depletion therapy in sporadic AD is illustrated in Fig. 2. Figure 2A-D show progressive stages of the disease. In each, levels of *iA* β have crossed the T1 threshold in all affected neurons. The transient *iA* β depletion treatment resets its levels to a low baseline, switches off the now unsustainable AD-driving A β PP-independent *iA* β production pathway, stops the progression of the disease and enables the still viable neurons to recover

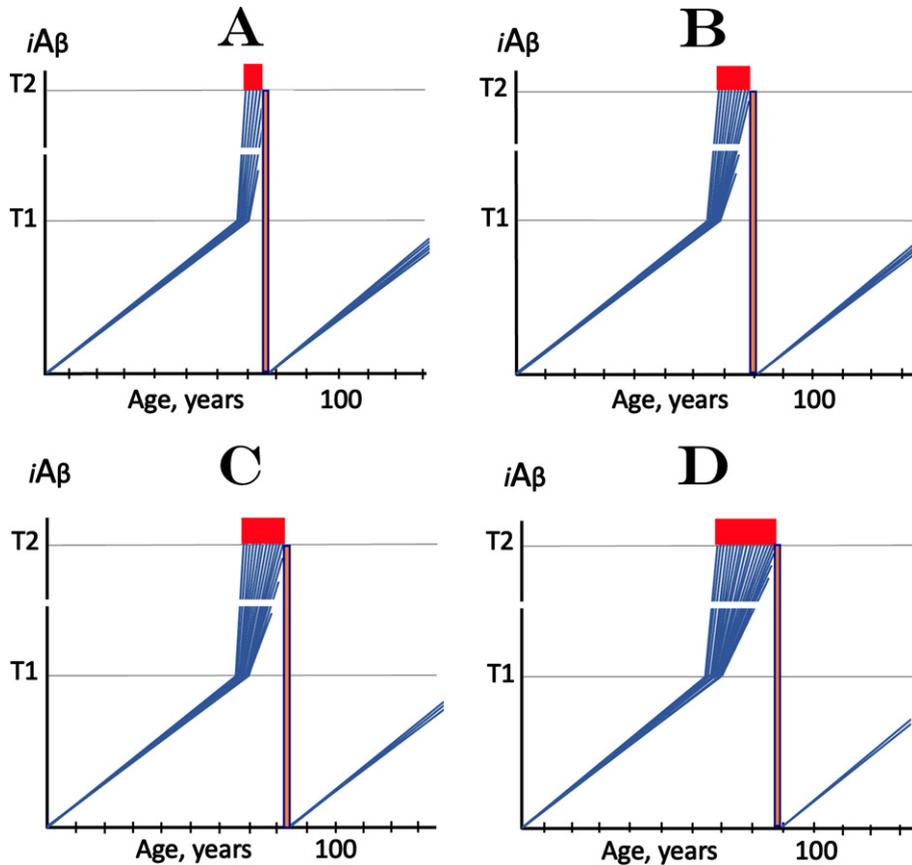


Fig. 2. Effect of transient $iA\beta$ depletion therapy at various stages of symptomatic AD. $iA\beta$: Level of intraneuronal $A\beta$. $T1$ threshold: Levels of $iA\beta$ triggering, plausibly via activation of PKR and HRI kinases, elicitation of the ISR and initiation of $A\beta$ PP-independent production of $iA\beta$. $T2$ threshold: Levels of $iA\beta$ triggering neurons' entrance into the apoptotic pathway. *Blue lines*: Individual affected neurons. *Red blocks*: Apoptotic zone. *Orange boxes*: The duration of the transient $iA\beta$ depletion treatment; levels of $iA\beta$ are reset and the accumulation of $A\beta$ PP-derived $iA\beta$ resumes from a low baseline. *A*: The transient $iA\beta$ depletion via its targeted degradation is implemented, via the enhancement of $iA\beta$ -cleaving activities of BACE1 and/or BACE2 or through employment of any suitable agent capable of $iA\beta$ depletion, at the early symptomatic stage of AD, with the bulk of the affected neurons still viable. Following the reset of $iA\beta$ levels, its build-up starts *de-novo*, supported only by the $A\beta$ PP proteolytic pathway. It is anticipated that $iA\beta$ levels will not reach the $T1$ threshold and AD will not recur within the remaining lifetime of an SAD patient. Note that the reset occurs in neurons that already crossed the $T1$ but not yet the $T2$ thresholds. *B-D*: The transient $iA\beta$ depletion treatment is implemented at progressively advanced stages of AD. The results are analogous to those depicted in *A*. However, at these AD stages increasing number of affected neurons cross the $T2$ threshold and commit apoptosis. This leaves progressively smaller number of affected neurons that retained their viability and can be redeemed. Note that the best therapeutic outcome can be obtained if the transient $iA\beta$ depletion treatment is administered preventively, prior to the activation of the $A\beta$ PP-independent $iA\beta$ production pathway and manifestation of AD symptoms (shown below); in this case all neurons would be redeemed, potentially for the remaining lifetime of an individual.

and reconnect. Since, with the progression of AD, less and less affected neurons with $iA\beta$ levels below the $T2$ threshold are left, progressively smaller neuronal fraction remains viable and can be redeemed. The levels of $iA\beta$, now driven solely by the $A\beta$ PP proteolysis and associated processes (internalization of secreted $A\beta$ and retention of $A\beta$ following the gamma-cleavage of C99 on intracellular membranes), are not expected to reach the $T1$ threshold and the disease is not expected to resume within the remaining lifetime of a patient. The best therapeutic

outcome can be obtained if the transient $iA\beta$ depletion treatment is administered preventively, prior to the activation of the $A\beta$ PP-independent $iA\beta$ production pathway and manifestation of AD symptoms (not shown in Fig. 2); in this case all neurons would be redeemed, potentially for the remaining lifetime of an individual.

Both approaches, (a) targeting the influx of $A\beta$ PP-derived $iA\beta$ and (b) depleting its levels via targeted degradation, can be effectively employed preventively. For the former, two conditions are crucial:

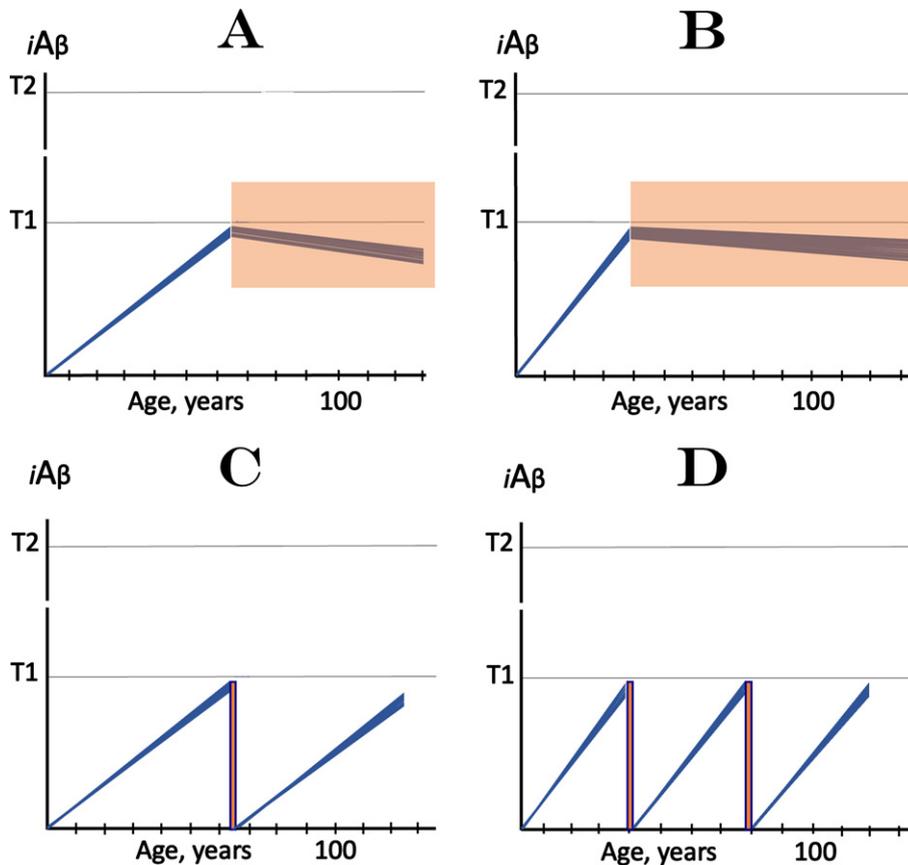


Fig. 3. The prevention of AD: Two approaches. $iA\beta$: Level of intraneuronal $A\beta$. $T1$ threshold: Levels of $iA\beta$ triggering, plausibly via activation of PKR and HRI kinases, elicitation of the ISR and initiation of $A\beta$ PP-independent production of $iA\beta$. $T2$ threshold: Levels of $iA\beta$ triggering neurons' entrance into the apoptotic pathway. *Blue lines*: Individual affected neurons. Note that in all panels the commencement or the implementation of a treatment occurs prior to the crossing of the $T1$ threshold by $A\beta$ PP-derived $iA\beta$. *A, B*: Prevention of AD via suppression of the rate of $A\beta$ PP-derived $iA\beta$ accumulation. It is assumed that a drug employed in this approach, which suppresses the influx of $A\beta$ PP-derived $iA\beta$ is capable of arresting or reversing $A\beta$ PP-derived $iA\beta$ accumulation or of reducing it (not shown) to the extent sufficient to prevent the $T1$ crossing within the lifetime of an individual. Under such a treatment, $iA\beta$ levels would not reach the $T1$ threshold, the $A\beta$ PP-independent $iA\beta$ generation pathway would not be activated, and AD would not occur for the duration of the treatment (*orange fields*); in this scenario, the treatment continues for the remaining part of the lifespan of an individual and constitutes, in effect, the AD "statin". *A*: Prevention of SAD; *B*: prevention of FAD (note the difference in the timing of the commencement of treatment). *C, D*: Prevention of AD by transient $iA\beta$ depletion via its targeted degradation (*orange boxes*; note drastic difference in duration of treatment in comparison with orange fields in *A* and *B*). The duration of the $iA\beta$ depletion treatment is defined by the desired extent of depletion and potentially could be as short as few days, a regiment possibly akin to that of an antibiotic treatment. It is assumed that, following the treatment, the $iA\beta$ pool would collapse, its levels would be reset to a low baseline, and the operation of the $A\beta$ PP-independent $iA\beta$ production pathway would cease. The accumulation of $A\beta$ PP-derived $iA\beta$, would resume at presumably constant rate; in prevention of SAD (*C*) its levels would not reach the $T1$ threshold, $A\beta$ PP-independent production of $iA\beta$ would not be activated, and the disease would not occur within the remaining lifetime of an individual. For prevention of FAD, the treatment is implemented earlier (*D*); following the treatment, accumulation of $A\beta$ PP-derived $iA\beta$ to near- $T1$ levels would require several decades but could occur within the lifetime of an individual and thus necessitate a repeat treatment. This approach could be implemented via the enhancement of $iA\beta$ -cleaving activities of BACE1 and/or BACE2 or through employment of any suitable agent causing selective degradation of $iA\beta$.

1) that the treatment commences prior to the $T1$ threshold crossing; any neurons that crossed would be unredeemable in this approach, and 2) that a drug employed causes the arrest or reversal of accumulation of $A\beta$ PP-derived $iA\beta$, or such reduction in the rate of the latter that would prevent the $T1$ crossing within individual's lifetime; otherwise the relief

would be only temporary. There is more leeway with the $iA\beta$ depletion via its targeted degradation since in this approach neurons are potentially redeemable until they cross the $T2$ threshold. It should be mentioned that the first approach reduces extracellular levels of $A\beta$ and thus may interfere with its protective effect (e.g., antimicrobial function [19]). In contrast,

the proposed transient $iA\beta$ depletion therapy would circumvent or minimize this potential problem due to its limited duration.

Preventive implementation of both strategies is illustrated in Fig. 3. Figures 3A and 3B depict the first approach for prevention of SAD and FAD respectively. The only difference is the timing of the commencement of treatment's administration. In this approach, $iA\beta$ levels would not reach the T1 threshold and AD would not occur for the duration of the treatment (orange fields); the treatment constitutes, in effect, the AD "statin". In Fig. 3C and 3D, the transient $iA\beta$ depletion therapy is deployed (orange boxes). A single transient treatment is potentially sufficient to prevent SAD within the remaining lifetime, and repeated treatments could be required for prevention of FAD.

In conclusion, lecanemab and donanemab are currently the only AD drugs that showed positive effect in clinical trials [1]. Even if they only reduce (rather than arrest or reverse) the rate of $A\beta$ PP-derived $iA\beta$ accumulation, this could be sufficient, if initiated presymptomatically, to prevent the T1 crossing, and, consequently, the occurrence of AD within the individual's lifetime. However, the mode of their administration (frequent infusions of large quantities of the antibody) substantially reduces the feasibility of their utilization as preventive agents. In the ACH2.0 perspective, any compound, possibly a small molecule, interfering with internalization of extracellular $A\beta$, either by preventing its oligomerization or through blocking its cellular receptors, would have effect similar to or exceeding that of lecanemab and donanemab, and could be feasible as an AD preventive agent. As an added benefit, such drug would not deplete extracellular $A\beta$ thus preserving its protective potential. Likewise, the proposed *transient* $iA\beta$ depletion therapy [5] would minimize or circumvent the depletion of extracellular $A\beta$ due to its limited duration. Moreover, whereas both strategies, the suppression of accumulation of $A\beta$ PP-derived $iA\beta$ via the reduction of its influx and the depletion of $iA\beta$ through its targeted degradation can be employed preventively, only the latter is capable of meaningful treatment of AD at its symptomatic stages.

ACKNOWLEDGMENTS

The authors are grateful to Dr. Bjorn R. Olsen (Harvard Medical School) for his support.

FUNDING

This work was supported by grants from the National Institutes of Health (R21 GM056179; R01 AR036819).

CONFLICT OF INTEREST

The authors have no conflict of interest to report.

DATA AVAILABILITY

Data sharing is not applicable to this article as no datasets were generated or analyzed during this study.

REFERENCES

- [1] van Dyck CH, Swanson CJ, Aisen P, Bateman RJ, Chen C, Gee M, Kanekiyo M, Li D, Reyderman L, Cohen S, Froelich L, Katayama S, Sabbagh M, Vellas B, Watson D, Dhadda S, Irizarry M, Kramer LD, Iwatsubo T (2023) Lecanemab in early Alzheimer's disease. *N Engl J Med* **388**, 9-21.
- [2] Prillaman M (2022) Alzheimer's drug slows mental decline in trial — but is it a breakthrough? *Nature* **610**, 15-16.
- [3] Gallagher J (2023) Alzheimer's-slowing drug labelled historic. <https://www.bbc.com/news/health-63060019>
- [4] Hardy JA, Higgins GA (1992) Alzheimer's disease: The amyloid cascade hypothesis. *Science* **256**, 184-185.
- [5] Volloch V, Rits-Volloch S (2022) The Amyloid Cascade Hypothesis 2.0: On the possibility of once-in-a-lifetime-only treatment for prevention of Alzheimer's disease and for its potential cure at symptomatic stages. *J Alzheimers Dis Rep* **6**, 369-399.
- [6] Chafekar S, Baas F, Scheper W (2008) Oligomer-specific amyloid-beta toxicity in cell models is mediated by selective uptake. *Biochem Biophys Acta* **9**, 523-531.
- [7] Wesen E, Jeffries G, Dzebo M, Esbjorner M (2017) Endocytic uptake of monomeric amyloid- β peptides is clathrin- and dynamin-independent and results in selective accumulation of $A\beta(1-42)$ compared to $A\beta(1-40)$. *Sci Rep* **7**, 2021.
- [8] Kumar-Singh S, Theuns J, Van Broeck B, Pirici D, Venekens K, Corsmit E, Cruts M, Dermaut B, Wang R, Van Broeckhoven C (2006) Mean age-of-onset of familial Alzheimer disease caused by presenilin mutations correlates with both increased Abeta42 and decreased Abeta40. *Hum Mutat* **27**, 686-695.
- [9] Hu X, Crick SL, Bu G, Frieden C, Pappu RV, Lee JM (2009) Amyloid seeds formed by cellular uptake, concentration, and aggregation of the amyloid-beta peptide. *Proc Natl Acad Sci U S A* **106**, 20324-20329.
- [10] Yajima R, Tokutake T, Koyama A, Kasuga K, Tezuka T, Nishizawa M, Ikeuchi T (2015) ApoE-isoform-dependent cellular uptake of amyloid- β is mediated by lipoprotein receptor LR11/SorLA. *Biochem Biophys Res Comm* **456**, 482-488.
- [11] Omtri RS, Davidson MW, Arumugam B, Poduslo JF, Kandimalla KK (2012) Differences in the cellular uptake and intracellular itineraries of amyloid beta proteins 40 and 42: Ramifications for the Alzheimer's drug discovery. *Mol Pharmaceutics* **9**, 1887.

- [12] Cook DG, Forman MS, Sung JC, Leight S, Kolson DL, Iwatsubo T, Lee VM, Doms RW (1997) Alzheimer's A beta42 is generated in the endoplasmic reticulum/intermediate compartment of NT2N cells. *Nat Med* **3**, 1021-1023.
- [13] Hartmann T, Bieger SC, Brühl B, Tienari PJ, Ida N, Allsop D, Roberts GW, Masters CL, Dotti CG, Unsicker K, Beyreuther K (1997) Distinct sites of intracellular production for Alzheimer's disease A beta40/42 amyloid peptides. *Nat Med* **3**, 1016-1020.
- [14] Wild-Bode C, Yamazaki T, Capell A, Leimer U, Steiner H, Ihara Y, Haass C (1997) Intracellular generation and accumulation of amyloid beta-peptide terminating at amino acid 42. *J Biol Chem* **272**, 16085-16088.
- [15] Manczak M, Anekonda TS, Henson E, Park BS, Quinn J, Reddy PH (2006) Mitochondria are a direct site of A beta accumulation in Alzheimer's disease neurons: Implications for free radical generation and oxidative damage in disease progression. *Hum Mol Genet* **15**, 1437-1449.
- [16] Jonsson T, Atwal JK, Steinberg S, Snaedal J, Jonsson PV, Bjornsson S, Stefansson H, Sulem P, Gudbjartsson D, Maloney J, Hoyte K, Gustafson A, Liu Y, Lu Y, Bhangale T, Graham RR, Huttenlocher J, Bjornsdottir G, Andreassen OA, Jonsson EG, Palotie A, Behrens TW, Magnusson OT, Kong A, Thorsteinsdottir U, Watts RJ, Stefansson K (2012) A mutation in APP protects against Alzheimer's disease and age-related cognitive decline. *Nature* **488**, 96-99.
- [17] Harper AR, Nayee S, Topol EJ (2015) Protective alleles and modifier variants in human health and disease. *Nat Rev Genet* **16**, 689-701.
- [18] Farzan M, Schnitzler CE, Vasilieva N, Leung D, Choe H (2000) BACE2, a beta-secretase homolog, cleaves at the beta site and within the amyloid-beta region of the amyloid-beta precursor protein. *Proc Natl Acad Sci U S A* **97**, 9712-9717.
- [19] Gosztyla ML, Brothers HM, Robinson SR (2018) Alzheimer's amyloid-beta is an antimicrobial peptide: A review of the evidence. *J Alzheimers Dis* **62**, 1495-1506.
- [20] Mintun M, et al. (2021). Donanemab in Early Alzheimer's Disease. *N Engl J Med* **384**, 1691-1704. DOI: 10.1056/NEJMoa2100708
- [21] Eli Lilly (2023) Donanemab Significantly Slowed Cognitive and Functional Decline in Phase 3 Study of Early Alzheimer's Disease. *Press release*. <https://investor.lilly.com/news-releases/news-release-details/lillys-donanemab-significantly-slowed-cognitive-and-functional>