

# Paraoxonase 1, B Vitamins Supplementation, and Mild Cognitive Impairment

Joanna Perła-Kaján<sup>a</sup>, Olga Włoczkowska<sup>a</sup>, Anetta Ziola-Frankowska<sup>b</sup>, Marcin Frankowski<sup>c</sup>,  
A. David Smith<sup>d</sup>, Celeste A. de Jager<sup>d</sup>, Helga Refsum<sup>e</sup> and Hieronim Jakubowski<sup>a,f,\*</sup>

<sup>a</sup>Department of Biochemistry and Biotechnology, Poznań University of Life Sciences, Poznań, Poland

<sup>b</sup>Department of Analytical Chemistry, Faculty of Chemistry, Adam Mickiewicz University, Poznań, Poland

<sup>c</sup>Department of Analytical and Environmental Chemistry, Faculty of Chemistry, Adam Mickiewicz University, Poznań, Poland

<sup>d</sup>OPTIMA, Department of Pharmacology, University of Oxford, Oxford, United Kingdom

<sup>e</sup>Department of Nutrition, Institute of Basic Medical Sciences, University of Oslo, Oslo, Norway

<sup>f</sup>Department of Microbiology, Biochemistry and Molecular Genetics, Rutgers University, New Jersey Medical School, International Center for Public Health, Newark, NJ, USA

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## Abstract.

**Background:** Identification of modifiable risk factors that affect cognitive decline is important for the development of preventive and treatment strategies. Status of paraoxonase 1 (PON1), a high-density lipoprotein-associated enzyme, may play a role in the development of neurological diseases, including Alzheimer's disease.

**Objective:** We tested a hypothesis that PON1 status predicts cognition in individuals with mild cognitive impairment (MCI).

**Methods:** Individuals with MCI ( $n = 196$ , 76.8-years-old, 60% women) participating in a randomized, double-blind placebo-controlled trial (VITACOG) were assigned to receive a daily dose of folic acid (0.8 mg), vitamin B<sub>12</sub> (0.5 mg) and B<sub>6</sub> (20 mg) ( $n = 95$ ) or placebo ( $n = 101$ ) for 2 years. Cognition was analyzed by neuropsychological tests. Brain atrophy was quantified in a subset of participants ( $n = 168$ ) by MRI. PON1 status, including *PON1 Q192R* genotype, was determined by quantifying enzymatic activity of PON1 using paraoxon and phenyl acetate as substrates.

**Results:** In the placebo group, baseline phenylacetate hydrolase (PhAcase) activity of PON1 (but not paraoxonase activity or *PON1 Q192R* genotype) was significantly associated with global cognition (Mini-Mental State Examination, MMSE; Telephone Inventory for Cognitive Status-modified, TICS-m), verbal episodic memory (Hopkins Verbal Learning Test-revised: Total Recall, HVLT-TR; Delayed Recall, HVLT-DR), and attention/processing speed (Trail Making A and Symbol Digits Modalities Test, SDMT) at the end of study. In addition to PhAcase, baseline iron and triglycerides predicted MMSE, baseline fatty acids predicted SDMT, baseline anti-*N*-Hcy-protein autoantibodies predicted TICS-m, SDMT, Trail Making A, while *BDNF V66M* genotype predicted HVLT-TR and HVLT-DR scores at the end of study. B-vitamins abrogated associations of PON1 and other variables with cognition.

**Conclusion:** PON1 is a new factor associated with impaired cognition that can be ameliorated by B-vitamins in individuals with MCI.

Keywords: *BDNF V66M* genotype, brain atrophy, cognition, iron, mild cognitive impairment, paraoxon, phenyl acetate, PON1 activity, *PON1 Q192R* genotype

\*Correspondence to: Hieronim Jakubowski, PhD, DSc, Department of Microbiology, Biochemistry and Molecular Genetics, Rutgers University, New Jersey Medical School, International

Center for Public Health, 225 Warren Street Newark, NJ 07103, USA. E-mail: jakubows@rutgers.edu.

## INTRODUCTION

Dementia is a major public health issue in aging modern societies that affects 50 million individuals worldwide (as of 2017) and continues to increase [1]. Brain atrophy occurs during normal aging and is a feature of neurodegeneration that becomes more prominent in individuals with mild cognitive impairment (MCI) and is further accelerated in Alzheimer's disease (AD) [2]. As no effective treatment for dementia is available, identification of modifiable risk factors that affect the rate of brain atrophy and cognitive decline is important for the development of preventive and treatment strategies [3, 4]. Reducing the rate of brain atrophy, for example by therapy with B vitamins [5, 6], is likely to slow the conversion of individuals with MCI to AD [2].

The prevalence of MCI, "a cognitive decline greater than that expected for an individual's age and education level but that does not interfere notably with activities of daily life" [7], is ~16% in individuals over 70 years of age [8, 9]. As half the MCI cases will convert to AD within 5 years after diagnosis [7], identification of biomarkers of cognitive decline and elucidation of their mechanisms of action has important public health policy implications.

PON1 is a calcium-dependent hydrolytic enzyme carried on high-density lipoprotein (HDL) in the circulation that contributes to the anti-inflammatory, anti-oxidative, and anti-atherothrombotic activities of HDL [10, 11]. The *PON1* gene has several polymorphisms, including the Q192R, which involves a change from glutamine (Q variant) to arginine (R variant) at position 192 of the amino acid sequence of the PON1 protein and affects its hydrolytic activity with natural [12] and artificial [13] substrates. Historically, the hydrolytic activity of the PON1 enzyme has been assayed with non-natural substrates such as the organophosphate paraoxon (for which the PON1 enzyme has been named) and phenyl acetate [13], both considered as surrogates for an unknown endogenous substrate that promotes atherogenesis [14]. One such endogenous substrate of PON1, homocysteine (Hcy) thiolactone [12, 15], which can adversely impact protein structure/function by post-translational modification [16], has been recently shown to be a predictor of myocardial infarction in coronary artery disease patients [17]. And indeed, the Hcy-thiolactonase activity of PON1 is strongly correlated with the paraoxonase (POase) activity in diverse populations (the United States [12], Poland [18], the United Kingdom [19], and the Netherlands [20])

while Hcy-thiolactone levels are significantly higher in carriers of low POase activity *PON1-192QQ* alleles compared with carriers of high POase activity *PON1-192RR* alleles [21], suggesting that the POase activity is a good surrogate for the physiological Hcy-thiolactonase activity. The phenylacetate hydrolase (PhAcase) activity, much less affected by the *PON1-Q192R* genotype, appears to be a good surrogate for the PON1 enzyme concentration [22].

Accumulating evidence suggests that the status of paraoxonase 1 (PON1) may have a role in neurological disease, including AD [23]. For example, genomic association studies found that a single-nucleotide polymorphism in the *PON1* gene is a risk factor for AD [24]. Other studies show that low PON1 activity is linked to the risk of AD and dementia [25, 26].

Low PhAcase activity of PON1, found in individuals with MCI [27, 28], has been associated with an increased risk of developing vascular dementia [29]. However, how PON1 affects functional and structural aspects of brain function was not known. For this reason, the present work has been undertaken to examine PON1 activity and genotype as predictors of cognition and brain atrophy rate in individuals with MCI subjected to B vitamin therapy or placebo treatment. This aim was accomplished by studying relationships between PON1 activity (PhAcase, POase), *PON1-Q192R* genotype and brain function (quantified by neuropsychological testing) as well as the rate of brain atrophy (quantified by MRI).

## MATERIALS AND METHODS

### *Participants*

We analyzed serum samples from individuals with MCI who fulfilled the Petersen criteria [30] and participated in a randomized controlled trial registered as VITACOG, ISRCTN 94410159 (<http://www.controlled-trials.com>) [5]. MCI patients had Mini-Mental State Examination (MMSE) score of >24/30 and no evidence of dementia. Other patients' characteristics were collected at baseline and have been previously described, as was the study protocol [5]. The present study included 77.6 ± 4.8-year-old participants ( $n=196$ , 60% women) randomly assigned to the treatment ( $n=95$ ) and placebo ( $n=101$ ) groups; some participants ( $n=168$ ) had MRI scans at baseline and at 24.3 ± 0.7 months of follow-up. Each participant received a daily oral TrioB Plus<sup>®</sup> supplement tablet (folic acid, 0.8 mg; vitamin B<sub>12</sub> - cyanocobalamin, 0.5 mg; vitamin B<sub>6</sub> - pyridoxine-HCl,

20 mg) or a placebo tablet for an average of two years. At baseline, 15.3 to 18.1% participants had a history of stroke, transient ischemic attack, or MRI infarct, 4.7 to 12.0% had diabetes, and 7.1 to 7.3% had myocardial infarction (in the treatment and placebo group, respectively); 49.4 to 43.4%, 30.6 to 33.7%, and 16.5 to 20.5% participants used cardiovascular disease drugs, aspirin, and B vitamins, respectively. Blood samples were collected at baseline and at the 24-months follow-up. All participants gave written informed consent. The study was carried out according to the principles of the Declaration of Helsinki and was approved by the Oxfordshire National Health Service research ethics committee (COREC 04/Q1604/100).

#### *MRI scans*

High-resolution T1-weighted images were acquired at baseline and at 2-year follow-up on a 1.5T Sonata MRI system (Siemens Medical Systems) and analyzed as previously described [5, 6].

#### *Cognitive testing*

At baseline and follow-up, neuropsychological tests were carried out by trained research nurses and psychologists blind to patient's clinical dementia rating, as previously described in the trial protocol [5]. The tests are representative of cognitive domains affected in MCI: global cognition (MMSE [31], Telephone Inventory for Cognitive Status modified (TICS-m) [32]); episodic memory (Hopkins Verbal Learning Test-revised (HVLTR) [33]; and attention/processing speed (Map Search [34], Trail Making A [35], Symbol Digits Modalities Test (SDMT) [36]). Executive function (Trail Making B, CLOX), semantic memory (Graded Naming, Category Fluency), the CANTAB Paired Associate Learning (visuospatial episodic memory), and Spatial Recognition Memory (SRM) tests were also used as outcome measures [37, 38].

#### *Inductively coupled plasma-mass spectrometry*

Serum samples (100  $\mu$ L) were mineralized with a mixture of redistilled nitric acid (70%, 300  $\mu$ L), hydrogen peroxide (25–35% for ultra-trace analysis, 100  $\mu$ L), and hydrochloric acid (30% suprapure, 100  $\mu$ L) for 24 h. Serial dilutions of ICP-MS single Al, As, Cu, Fe, and Si standard solutions were used for calibration. Additionally, for ICP-MS Sc, Rh, Be, and Ge in 1%  $\text{HNO}_3 \geq 99.999\%$  trace metals basis were

used as internal standards (automatically added during analysis through T-piece). Reagents and standards for mineralization were purchased from the Sigma Aldrich Merck group, Poznań, Poland. Deionized water was obtained from the Milli-Q Direct 8 Water Purification System (Merck Millipore). A certified reference material, BCR 637 (Institute for Reference Materials and Measurements) as well as reference material ERM-DA120 (European Reference Materials) were analyzed to validate the calibration. Analyses were in agreement with certified values, with recoveries from 93 to 104%. The intra-assay analytical variability, determined with six assays of the same sample in one run, was 1.3% and 6.4% for Cu and Fe, respectively. The inter-assay variability, determined with 20 samples assayed on different days was 20.1%, 13.1%, 1.7%, 13.2%, and 2.1% for Al, As, Cu, Fe, and Si, respectively.

#### *Paraoxonase 1 activity*

Serum PON1 activity was quantified by using POase and arylesterase (PhAcase) assays as described previously [15, 20]. Briefly, reaction mixtures contained 0.05 M K-Hepes buffer (pH 7.4), 1 mM  $\text{CaCl}_2$ , serum (100- and 400-fold diluted for POase and PhAcase assays, respectively), and paraoxon (2 mM) or phenyl acetate (5 mM). The assays were carried out in duplicates at 25°C in a 96-well plate format using Infinite M200Pro Spectrophotometer (Tecan, Männedorf, Switzerland), initiated by the addition of the substrate, monitored by spectroscopy. Each run included blanks in which PON1 activity was inactivated by 10 mM EDTA.

For POase activity assays, the generation of *p*-nitrophenol from paraoxon (MiliporeSigma, St. Louis, MO) was monitored at 412 nm ( $\epsilon = 13,000 \text{ M}^{-1} \text{ cm}^{-1}$ ) for 5 min and reaction rates ( $A_{412}/\text{min}$ ) were calculated. For PhAcase activity assays, the generation of phenol from phenyl acetate (MiliporeSigma, St. Louis, MO) was monitored at 270 nm ( $\epsilon = 1300 \text{ M}^{-1} \text{ cm}^{-1}$ ) for 3 min and the rates ( $A_{270}/\text{min}$ ) calculated. Coefficients of variance for POase and PhAcase activities were 6.1% and 2.1%, respectively. One unit of activity is defined as a change in the absorbance of 0.0001 per min.

#### *PON1 Q192R polymorphism*

The *PON1 Q192R* polymorphism was established by the two substrate activity measurements method [39, 40] (Fig. 1).

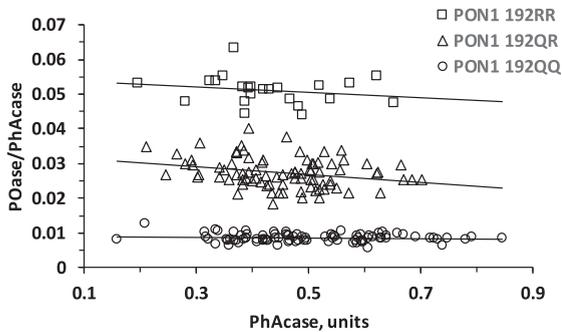


Fig. 1. Population distribution plot for POase versus PhAcase showing resolution of MCI individuals according to *PON1 192QQ*, *PON1 192QR*, and *PON1 192RR* genotypes.

### *BDNF V66M genotyping*

Genomic DNA was isolated from whole human blood using the phenol extraction procedure and stored at  $-80^{\circ}\text{C}$ . The human *BDNF* locus was genotyped by PCR-RFLP as previously described [41]. The *BDNF V66M* alleles were amplified using the primers 5'-AAAGAAGCAAACATCCGAGGACAA G-3' (forward) and 5'-ATTCCTCCAGCAGAAAGA GAAGAGG-3' (reverse) at an annealing temperature  $95^{\circ}\text{C}$ -3 min,  $95^{\circ}\text{C}$ -15 s,  $56^{\circ}\text{C}$ -15 s,  $72^{\circ}\text{C}$ -5 min for 40 cycles. The 274 bp PCR product was digested with *Hin1III* (*NlaIII*) (New England Biolabs) and analyzed on 4% agarose gels. Two bands (216 and 58 bp) indicate the *BDNF-66V* allele while three bands (139, 77, and 258 bp) signify the *BDNF-66M* allele.

### *Anti-N-Hcy-protein autoantibody assays and metabolite analyses*

Values for serum anti-*N*-Hcy-protein autoantibodies [42], plasma total homocysteine (tHcy), and other metabolites [5] were obtained from analyses reported previously.

### *Statistical analysis*

Normality of variables distributions was examined using the Shapiro-Wilk's statistic or Chi-square

test. Non-normally distributed variables were log-transformed. Unpaired two-sided or paired *t*-tests were used for comparisons between two groups of variables. Associations between variables were examined by Pearson's correlations and multiple linear regression using log-transformed data. In Tables 2–8, the independent variables included in a particular model are those that have numerical entries in columns for each model. Other independent variables are indicated in the legend to each table. Statistical software packages PSPP, version 1.0.1 (<http://www.gnu.org>) and Statistica, version 13 (TIBCO Software Inc., Palo Alto, CA, USA, <http://statistica.io>) were used. Probability values were 2-sided and *p* value  $< 0.05$  was considered statistically significant.

## RESULTS

### *Baseline PON1 characteristics in the MCI cohort*

We quantified serum PON1 activities in a cohort of MCI patients ( $n = 196$ ) using paraoxon (for POase) or phenylacetate (PhAcase) as substrates. These measurements allow identification of *PON1 Q192R* genotype for each participant in the study [39, 40]. A plot of POase/PhAcase versus PhAcase shows three distinct groups of data points that identify *192QQ*, *192QR*, and *192RR* individuals in our cohort (Fig. 1).

The frequencies of the *192QQ*, *192QR*, and *192RR* genotypes in the MCI cohort were 44.4%, 43.8%, and 11.7%, respectively (Table 1), similar to other European populations [18, 19, 21, 40, 43]. The *192Q* and *192R* alleles were in the Hardy-Weinberg equilibrium (established by the Chi-square Test for Normality), as observed in other populations [12, 21, 39, 43].

The *192QQ* and *192RR* genotypes were associated with low and high POase activities, respectively (Table 1). In contrast, the *Q192R* polymorphism had an inverse effect on the PhAcase activity: *192QQ* and *192RR* genotypes were associated with high and low

Table 1  
POase and PhAcase activities of PON1 stratified by *PON1 Q192R* genotype in participants with MCI\*

Genotype	<i>n</i> (%)	POase		PhAcase	
		Units $\times 100$	<i>p</i> versus <i>QQ</i>	Units	<i>p</i> versus <i>QQ</i>
<i>192QQ</i>	87 (44.4)	$0.43 \pm 0.12$		$0.51 \pm 0.13$	
<i>192QR</i>	86 (43.8)	$1.23 \pm 0.30$	1.3E-54	$0.45 \pm 0.10$	7.9E-3
<i>192RR</i>	23 (11.7)	$2.16 \pm 0.54$	1.8E-50	$0.43 \pm 0.11$	2.2E-29
All	196 (100)	$0.98 \pm 0.64$		$0.47 \pm 0.12$	

\*PhAcase, phenylacetate hydrolase; POase, paraoxonase.

PhAcase activities, respectively (Table 1), consistent with findings in other populations [21, 44, 45].

*Neuropsychological measures of cognition are associated with PhAcase activity of PONI at baseline*

Associations between baseline PhAcase, POase, *PONI Q192R* genotype, and neuropsychological measures of cognition and other baseline variables are shown in Table 2.

Pearson correlation analysis showed that baseline PhAcase activity was significantly associated with neuropsychological measures in three domains of cognition at baseline: verbal episodic memory (HVL-TR, HVL-DR) (Fig. 2A, B), semantic memory (Category Fluency) (Fig. 2C), and attention/processing speed (Trail Making A) (Fig. 2D). PhAcase was also significantly associated with other variables at baseline: tHcy (Fig. 2E), Fe (Fig. 2F), Al, Si (but not with Cu and As), creatinine, taurine, cysteine, age, *BDNF V66M* genotype, and, as expected [21, 44, 45], with *PONI Q192R* genotype (Table 2). In contrast, baseline POase activity was not associated with neuropsychological measures of cognition

but was associated with Si, As, anti-*N*-Hcy autoantibodies, and *PONI Q192R* genotype (Table 2).

In multiple regression analysis, baseline PhAcase activity was significantly associated with iron, creatinine, and *PONI Q192R* genotype. In contrast, baseline POase activity was associated only with *PONI Q192R* genotype (Table 2).

*PONI measures at baseline determine cognition at the end of study: The placebo group*

Multiple regression analysis for the placebo group in models including neuropsychological test score at the end of study as a dependent variable and *PONI* measures and other variables at baseline as independent variables revealed that baseline PhAcase, but not POase nor *PONI Q192R* genotype, was a predictor of cognition in four domains at the end of study: global cognition (MMSE, TICS-m), verbal episodic memory (HVL-TR, HVL-DR), and attention/processing speed (SDMT, Trail Making A). There was no association between baseline PhAcase and scores in other neuropsychological tests at the end of study: PAL Total Errors and CATNAB SRM (visuospatial episodic memory), Trail Making

Table 2  
Baseline determinants of *PONI* activities\*

Variable (n = 186–196)	LnPhAcase				LnPOase			
	Pearson correlation		Multiple regression		Pearson correlation		Multiple regression	
	β	p	β	p	β	p	β	p
<i>PONI Q192R</i>	-0.17	<b>0.016</b>	-0.15	<b>0.026</b>	0.89	<b>0.000</b>	0.90	<b>0.000</b>
<i>BDNF V66M</i>	-0.17	<b>0.014</b>			0.00	0.972		
LnFe	-0.32	<b>0.000</b>	-0.21	<b>0.000</b>	-0.01	0.877	-0.06	0.077
LnCu	0.07	0.340			-0.06	0.404		
LnAl	-0.25	<b>0.001</b>			-0.01	0.942		
LnAs	0.02	0.775			0.18	<b>0.015</b>		
LnSi	-0.20	<b>0.006</b>			-0.16	<b>0.034</b>		
LnCreatinine	-0.26	<b>0.000</b>	-0.23	<b>0.001</b>	-0.05	0.495	-0.05	0.160
LnTaurine	0.14	<b>0.044</b>			0.03	0.632		
LnHcy	-0.21	<b>0.003</b>			-0.04	0.607		
LnCys	-0.17	<b>0.018</b>			0.02	0.757		
LnHVL-DR	0.18	<b>0.017</b>			0.01	0.845		
LnHVL-TR	0.15	<b>0.035</b>			0.05	0.526		
LnTrail Making A	-0.14	<b>0.043</b>			0.01	0.918		
LnCategory_Fluency	0.16	<b>0.024</b>			0.03	0.396		
LnAnti- <i>N</i> -Hcy	-0.03	0.692			0.15	<b>0.046</b>		
Age	-0.20	<b>0.005</b>			0.00	0.962		
Sex	0.11	0.133			0.011	0.122		
				F = 12.7, p = 0.000, Adjusted R <sup>2</sup> = 0.16				F = 242.7, p = 0.000, Adjusted R <sup>2</sup> = 0.80

\*Ln, natural logarithm; PhAcase, phenylacetate hydrolase; POase, paraoxonase.

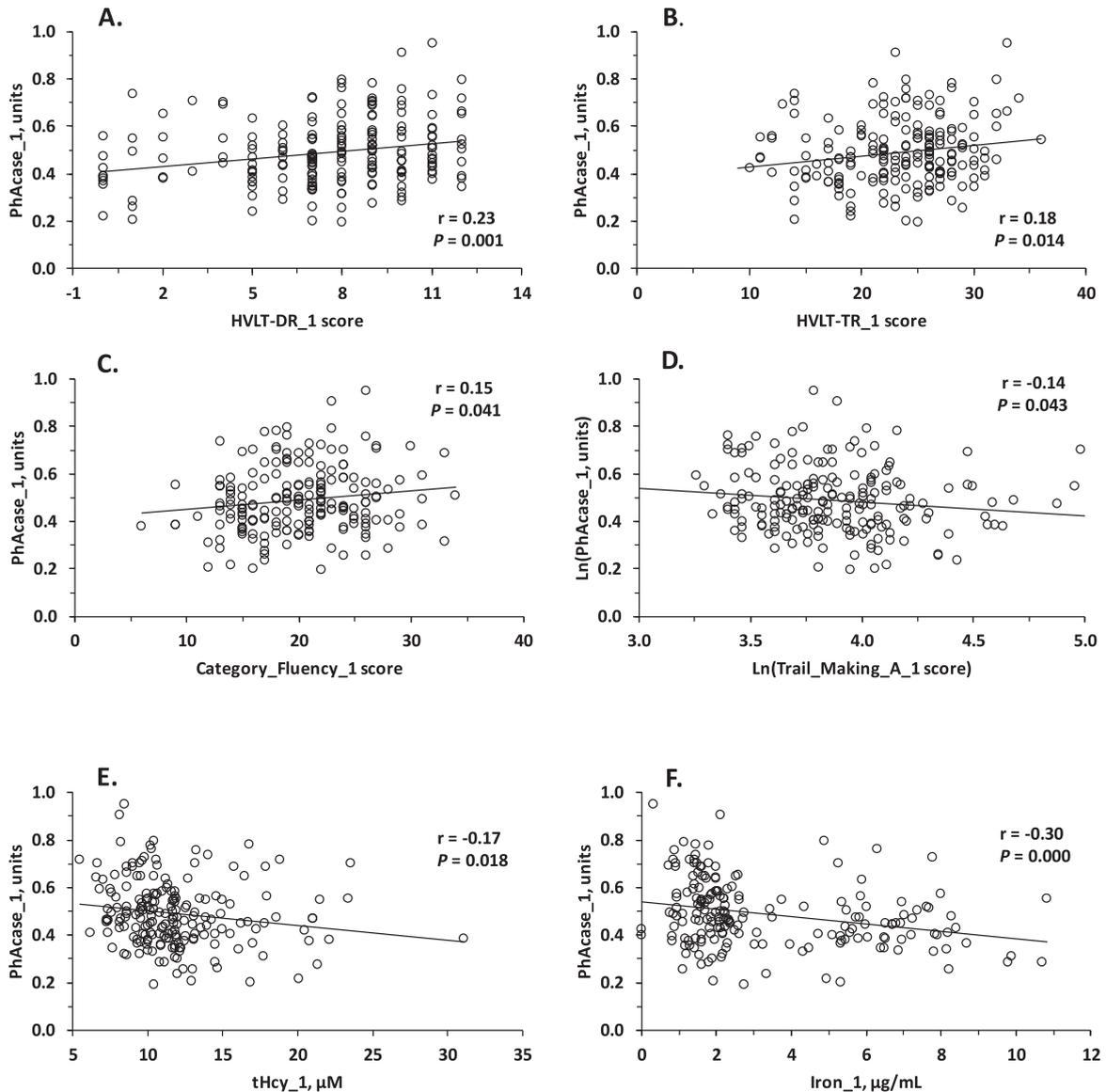


Fig. 2. Relationships between PhAcase activity of serum PON1 and verbal episodic memory (A) HVLT-DR, (B) HVLT-TR test scores, tHcy (C), and iron (D) at baseline.

B and CLOX (executive function), Map Search (complex attention/processing speed), Graded Naming and Category Fluency (semantic memory).

#### Global cognition: MMSE\_2

Baseline variables that determined global cognition score in the MMSE\_2 test at the end of study were PhAcase ( $\beta = -0.24$ ,  $p = 0.034$ ), Fe ( $\beta = 0.24$ ,  $p = 0.031$ ), tHcy ( $\beta = -0.32$ ,  $p = 0.017$ ), triglycerides ( $\beta = -0.22$ ,  $p = 0.044$ ), MMSE\_1 score ( $\beta = 0.26$ ,  $p = 0.017$ ), rate of brain atrophy ( $\beta = -0.27$ ,

$p = 0.029$ ), and age ( $\beta = -0.24$ ,  $p = 0.056$ ); adjusted  $R^2$  was 0.43 (Table 3, Model 1)s.

*PON1* Q192R genotype did not affect these associations and MMSE\_2 was not associated with *PON1* Q192R genotype in models with (Model 1:  $\beta = -0.02$ ,  $p = 0.854$ ) or without PhAcase (Model 3:  $\beta = 0.06$ ,  $p = 0.624$ ). MMSE\_2 was also not associated with POase in models with (Model 4:  $\beta = -0.28$ ,  $p = 0.246$ ) or without *PON1* Q192R genotype (Model 5:  $\beta = -0.04$ ,  $p = 0.725$ ).

We also found that MMSE\_2 score was significantly associated with baseline iron independently of

Table 3  
Determinants of global cognition at the end of study: LnMMSE.2, placebo group\*

Variable (n = 82–112)	Pearson correlation	Multiple regression#												
		Model 1		Model 2		Model 3		Model 4		Model 5		Model 6		
		β	p	β	p	β	p	β	p	β	p	β	p	
LnHcy.1	-0.28	0.003	-0.32	<b>0.017</b>	-0.29	<b>0.030</b>	-0.32	<b>0.023</b>	-0.31	<b>0.024</b>	-0.27	<b>0.050</b>	-0.29	<b>0.037</b>
LnPOase.1	-0.06	0.526												
LnPhAcase.1	-0.11	0.286	-0.24	<b>0.034</b>	-0.23	<b>0.033</b>								
PONI-Q192R	-0.01	0.964	-0.02	0.854										
LnTG.1	-0.08	0.387	-0.22	<b>0.044</b>	-0.20	0.058	0.06	0.624	0.30	0.211	-0.20	0.071	-0.19	0.084
LnFe.1	0.05	0.650	0.24	<b>0.031</b>	0.23	<b>0.030</b>	0.28	<b>0.014</b>	0.26	<b>0.023</b>	0.29	<b>0.010</b>	0.27	<b>0.013</b>
LnAtrophy.rate	-0.31	0.006	-0.27	<b>0.029</b>	-0.25	<b>0.038</b>	-0.24	0.057	-0.25	<b>0.045</b>	-0.24	0.062	0.33	<b>0.002</b>
LnMMSE.1	0.34	0.000	0.26	<b>0.017</b>	0.29	<b>0.005</b>	0.32	<b>0.004</b>	0.30	<b>0.006</b>	0.43	<b>0.000</b>	0.33	<b>0.002</b>
Age	-0.28	0.003	-0.24	<b>0.056</b>	-0.26	<b>0.034</b>	-0.24	0.063	-0.23	<b>0.071</b>	-0.27	<b>0.039</b>	-0.26	<b>-0.041</b>
				F = 5.63, p = 0.000, Adjusted R <sup>2</sup> = 0.43		F = 5.63, p = 0.000, Adjusted R <sup>2</sup> = 0.43		F = 4.86, p = 0.000, Adjusted R <sup>2</sup> = 0.39		F = 4.57, p = 0.000, Adjusted R <sup>2</sup> = 0.39		F = 4.73, p = 0.000, Adjusted R <sup>2</sup> = 0.38;		F = 5.21, p = 0.000, Adjusted R <sup>2</sup> = 0.38

\*Ln, natural logarithm; PhAcase, phenylacetate hydrolase; POase, paraoxonase; TG, triglyceride; .1, baseline; .2, end of study. # Adjusted for sex, LnAnti-N-Hcy.1, and *BDNF V66M* genotype.

the PON1 status in models with or without PhAcase, POase, and/or *PONI Q192R* genotype (Models 1–6).

The R<sup>2</sup> values in Models 1–5 versus Model 6 (Table 3) suggested that PhAcase explained a greater fraction of the MMSE.2 score variation (at least 5%; Model 2 versus Model 6) than did POase (0%; Model 5 versus Model 6) or *PONI Q192R* genotype (1%; Model 3 versus Model 6). Because higher score in the MMSE test indicates better cognitive outcome, these findings suggest that higher PhAcase activity has a detrimental effect on general cognition, similar to the detrimental effect of elevated tHcy.

*Global cognition/memory: TICS-m.2*

Baseline variables that significantly determined global cognition/memory score in the TICS-m.2 test at the end of study were PhAcase (β = -0.27, p = 0.005), tHcy (β = -0.23, p = 0.039), TICS-m.1 score (β = 0.23, p = 0.017), rate of brain atrophy (β = -0.28, p = 0.007), and age (β = -0.34, p = 0.036); adjusted R<sup>2</sup> was 0.52 (Table 4, Model 2).

*PONI Q192R* genotype did not affect these associations and TICS-m.2 was not significantly associated with *PONI Q192R* genotype in models with (Model 1: β = 0.09, p = 0.372) or without PhAcase (Model 3: β = 0.18, p = 0.065). TICS-m.2 was also not associated with POase in models with (Model 4: β = -0.25, p = 0.285) or without *PONI Q192R* genotype (Model 5: β = 0.12, p = 0.241).

We also found that TICS-m.2 score was significantly associated with baseline anti-N-Hcy autoantibodies in models with (Model 1: β = -0.20, p = 0.049) and without PhAcase (Model 3: β = -0.24, p = 0.021; Model 4; β = -0.22, p = 0.042; Model 5: β = -0.24, p = 0.028).

The R<sup>2</sup> values in Models 1–5 versus Model 6 (Table 4) suggested that PhAcase explained a greater fraction of variation (at least 6%; Model 2 versus Model 6) in the TICS-m.2 score than did POase (0%; Model 5 versus Model 6) or *PONI Q192R* (2%; Model 3 versus Model 6) genotype. Because higher score in the TICS-m test indicates better cognition, these findings suggest that higher PhAcase activity has a detrimental effect on global cognition/memory, similar to the detrimental effect of elevated tHcy or anti-N-Hcy autoantibodies (Table 4).

*Episodic memory: HVLT-TR.2*

Baseline variables that significantly determined episodic memory score in the HVLT-TR.2 test at

Table 4  
Determinants of global memory at the end of study: LnTICSm\_2, placebo

Variable (n = 82–112)	Pearson correlation	Multiple regression <sup>#</sup>											
		Model 1		Model 2		Model 3		Model 4		Model 5		Model 6	
		$\beta$	p	$\beta$	p	$\beta$	p	$\beta$	p	$\beta$	p	$\beta$	p
<b>LnHcy_1</b>	-0.39	0.000	-0.23	<b>0.040</b>	-0.21	0.071	-0.23	0.056	-0.20	0.087	-0.20	0.086	
<b>LnPhAcase_1</b>	-0.16	0.113	-0.27	<b>0.005</b>									
LnPOase_1	-0.04	0.721											
<i>PON1-Q192R</i>	0.03	0.744			0.18	0.065	-0.25	0.285	0.12	0.241			
<b>LnAnti-N-Hcy_1</b>	0.03	0.763	-0.18	0.068	-0.24	<b>0.021</b>	0.39	0.076	-0.24	<b>0.028</b>	-0.20	0.052	
<b>LnAtrophy_rate</b>	-0.41	0.000	-0.28	<b>0.007</b>	-0.23	<b>0.035</b>	-0.24	<b>0.042</b>	-0.22	<b>0.040</b>	-0.23	<b>0.035</b>	
LnTICSm_1	0.43	0.000	0.23	<b>0.017</b>	0.35	<b>0.001</b>	0.31	<b>0.004</b>	0.36	<b>0.001</b>	0.33	<b>0.001</b>	
Age	-0.36	0.000	-0.34	<b>0.036</b>	-0.39	<b>0.001</b>	-0.38	<b>0.001</b>	-0.40	<b>0.001</b>	-0.38	<b>0.001</b>	
			F = 9.39, p = 0.000, Adjusted R <sup>2</sup> = 0.51	F = 10.93, p = 0.000, Adjusted R <sup>2</sup> = 0.52	F = 9.31, p = 0.000, Adjusted R <sup>2</sup> = 0.48	F = 8.31, p = 0.000, Adjusted R <sup>2</sup> = 0.48	F = 8.90, p = 0.000, Adjusted R <sup>2</sup> = 0.46;	F = 10.35, p = 0.000, Adjusted R <sup>2</sup> = 0.46					

\*Ln, natural logarithm; PhAcase, phenylacetate hydrolase; POase, paraoxonase; \_1, baseline; \_2, end of study. <sup>#</sup>Adjusted for sex.

the end of study were PhAcase ( $\beta = -0.20$ ,  $p = 0.028$ ), HVLT-TR\_1 score ( $\beta = 0.44$ ,  $p = 0.000$ ), age ( $\beta = -0.26$ ,  $p = 0.008$ ), and *BDNF V66M* genotype ( $\beta = 0.22$ ,  $p = 0.021$ ); adjusted R<sup>2</sup> was 0.54 (Table 5, Model 2).

*PON1 Q192R* genotype did not affect these associations and HVLT-TR\_2 was not associated with *PON1 Q192R* genotype in models with (Model 1:  $\beta = 0.03$ ,  $p = 0.714$ ) or without PhAcase (Model 3:  $\beta = 0.07$ ,  $p = 0.423$ ). HVLT-TR\_2 was also not significantly associated with POase in models with (Model 4:  $\beta = -0.40$ ,  $p = 0.053$ ) or without *PON1 Q192R* genotype (Model 5:  $\beta = -0.02$ ,  $p = 0.840$ ).

The association of HVLT-TR\_2 score with *BDNF V66M* genotype was independent of the *PON1* status and was observed in models with or without PhAcase, POase, and/or *PON1 Q192R* genotype (Models 1–6). The carriers of *BDNF 66V* allele had a higher HVLT-TR\_2 score (better cognition) compared with carriers of *BDNF 66M* allele.

The R<sup>2</sup> values in Models 1–5 versus Model 6 (Table 5) suggested that PhAcase explained a greater fraction of variation (at least 3%; Model 2 versus Model 6) in the HVLT-TR\_2 score than did POase (-2%; Model 5 versus Model 6) or *PON1 Q192R* (-1%; Model 3 versus Model 6) genotype. Because higher score in the HVLT-TR test indicates better cognition, these findings suggest that higher PhAcase activity has a detrimental effect on episodic memory.

#### Episodic memory: HVLT-DR\_2

Baseline variables that significantly determined episodic memory score in the HVLT-DR\_2 test at the end of study were PhAcase ( $\beta = -0.29$ ,  $p = 0.015$ ), HVLT-DR\_1 score ( $\beta = 0.44$ ,  $p = 0.001$ ), and *BDNF V66M* genotype ( $\beta = 0.29$ ,  $p = 0.015$ ); adjusted R<sup>2</sup> was 0.37 (Table 6, Model 2).

*PON1 Q192R* genotype did not affect these associations: HVLT-DR\_2 was not associated with *PON1 Q192R* genotype in models with (Model 1:  $\beta = -0.14$ ,  $p = 0.231$ ) or without PhAcase (Model 3:  $\beta = -0.05$ ,  $p = 0.655$ ). HVLT-DR\_2 was also not significantly associated with POase in models with (Model 4:  $\beta = -0.54$ ,  $p = 0.053$ ) or without *PON1 Q192R* genotype (Model 5:  $\beta = -0.16$ ,  $p = 0.192$ ).

The association of HVLT-DR\_2 score with *BDNF V66M* genotype was independent of the *PON1* status in models with or without PhAcase, POase, and/or *PON1 Q192R* genotype (Models 1–6). The carriers of *BDNF 66V* allele had a higher HVLT-DR\_2 score

Table 5  
Determinants of verbal episodic memory at the end of study: LnHVLTR-TR\_2, placebo group\*

Variable (n = 82–112)	Pearson correlation		Multiple regression <sup>#</sup>											
	β	p	Model 1		Model 2		Model 3		Model 4		Model 5		Model 6	
			β	p	β	p	β	p	β	p	β	p	β	p
LntHcy-1	-0.36	0.000												
LnPOase_1	0.02	0.840							-0.40	0.053	-0.02	0.840		
<b>LnPhAcase_1</b>	-0.01	0.939	-0.19	<b>0.046</b>	-0.20	<b>0.028</b>								
<i>PONI-Q192R</i>	0.07	0.517	0.03	0.714			0.07	0.423	0.44	<b>0.037</b>				
LnCreatinine_1	-0.33	0.000												
LnAtrophy_rate	-0.35	0.001	-0.19	0.177	-0.19	0.076	-0.18	0.105	-0.20	0.058	-0.17	0.116	-0.15	0.212
LnHVLTR-TR_1	0.69	0.000	0.45	<b>0.000</b>	0.44	<b>0.000</b>	0.43	<b>0.000</b>	0.41	0.000	0.42	<b>0.000</b>	0.43	<b>0.000</b>
<i>BDNF V66M</i>	0.09	0.345	0.21	<b>0.034</b>	0.22	<b>0.021</b>	0.024	<b>0.017</b>	0.25	<b>0.011</b>	0.27	<b>0.008</b>	0.21	<b>0.019</b>
Age	-0.29	0.002	-0.25	<b>0.012</b>	-0.26	<b>0.008</b>	-0.25	<b>0.014</b>	-0.25	<b>0.023</b>	-0.27	<b>0.008</b>	-0.30	<b>0.001</b>
Sex	0.21	0.030												
			F = 8.89, p = 0.000, Adjusted R <sup>2</sup> = 0.53		F = 10.37, p = 0.000, Adjusted R <sup>2</sup> = 0.54		F = 8.97, p = 0.000, Adjusted R <sup>2</sup> = 0.50		F = 8.82, p = 0.000, Adjusted R <sup>2</sup> = 0.52		F = 8.95, p = 0.000, Adjusted R <sup>2</sup> = 0.49;		F = 12.35, p = 0.000, Adjusted R <sup>2</sup> = 0.51	

\*Ln, natural logarithm; PhAcase, phenylacetate hydrolase; POase, paraoxonase; -1, baseline; -2, end of study. <sup>#</sup>Adjusted for sex, creatinine\_1, LntHcy\_1, and *TCN 776CG* genotype. LntHcy\_1 was not significant in any of the models.

(better cognition) compared with carriers of *BDNF* 66M allele.

The  $R^2$  values in Models 1–5 versus Model 6 (Table 6) suggested that PhAcase explained a greater fraction of variation in the HVLT-TR\_2 score (at least 4%; Model 2 versus Model 6) than did POase (0%; Model 5 versus Model 6) or *PON1 Q192R* genotype (-3%; Model 3 versus Model 6). Because higher score in the HVLT-DR test indicates better cognition, these findings suggest that higher PhAcase activity has a detrimental effect on episodic memory.

#### *Attention/processing speed: Trail Making A\_2*

Baseline variables that significantly determined attention/speed score in the Trail Making A\_2 test at the end of study were PhAcase ( $\beta=0.24$ ,  $p=0.015$ ), tHcy ( $\beta=0.32$ ,  $p=0.008$ ), anti-*N*-Hcy autoantibodies ( $\beta=0.24$ ,  $p=0.025$ ), Trail Making A\_1 score ( $\beta=0.32$ ,  $p=0.000$ ), *PON1 Q192R* genotype ( $\beta=0.19$ ,  $p=0.049$ ), and age ( $\beta=0.31$ ,  $p=0.005$ ); adjusted  $R^2$  was 0.57 (Table 7, Model 1). The carriers of *PON1 192R* allele had a higher Trail Making A\_2 score compared with carriers of *PON1 192Q* allele. Trail Making A\_2 tended to be associated with PhAcase in a model without *PON1 Q192R* genotype (Model 2;  $\beta=0.19$ ,  $p=0.055$ ) but was not associated with *PON1 Q192R* genotype in models without PhAcase (Models 3 and 4) nor with POase in models with (Model 4:  $\beta=0.16$ ,  $p=0.453$ ) or without *PON1 Q192R* genotype (Model 5:  $\beta=0.14$ ,  $p=0.148$ ).

The association of HVLT-DR\_2 score with anti-*N*-Hcy autoantibodies was independent of the *PON1* status in models with or without PhAcase, POase, and/or *PON1 Q192R* genotype (Models 1–6).

The  $R^2$  values in Models 1–5 versus Model 6 (Table 7) suggested that PhAcase explained a greater fraction of variation in the Trail Making A\_2 score (2–4%; Models 2 and 1 versus Model 6) than did POase (0%; Model 5 versus Model 6) or *PON1 Q192R* genotype (0%; Model 3 versus Model 6). Because higher score in Trail Making A test indicates worse cognition, these findings suggest that higher PhAcase activity has a detrimental effect on attention/processing speed.

#### *Attention/processing speed: SDMT\_2*

Baseline variables that significantly determined global cognition score in the SDMT\_2 test at the end of study were PhAcase ( $\beta=-0.16$ ,  $p=0.011$ ), tHcy ( $\beta=-0.19$ ,  $p=0.013$ ), anti-*N*-Hcy autoantibodies

( $\beta=-0.15$ ,  $p=0.027$ ), fatty acids (FA:  $\beta=-0.20$ ,  $p=0.017$ ), brain atrophy ( $\beta=-0.24$ ,  $p=0.001$ ), and SDMT\_1 score ( $\beta=0.67$ ,  $p=0.000$ ); adjusted  $R^2$  was 0.79 (Table 8, Model 2). SDMT\_2 was associated with POase only in a model with *PON1 Q192R* genotype (Model 4:  $\beta=-0.33$ ,  $p=0.028$ ) but not without (Model 5:  $\beta=-0.06$ ,  $p=0.336$ ). Similarly, SDMT\_2 was associated with *PON1 Q192R* genotype in a model with POase (Model 4:  $\beta=0.29$ ,  $p=0.047$ ) but not without (Model 3:  $\beta=0.00$ ,  $p=0.963$ ). In contrast, SDMT\_2 was not associated with *PON1 Q192R* genotype in models with (Model 1:  $\beta=-0.18$ ,  $p=0.008$ ) or without PhAcase (Model 3:  $\beta=0.00$ ,  $p=0.963$ ). Because a higher score in SDMT test indicates better cognition, these findings suggest that higher PhAcase activity has a detrimental effect on attention/processing speed.

Associations of the SDMT\_2 score with anti-*N*-Hcy autoantibodies and fatty acids (FA\_1) were independent of the *PON1* status in models with or without PhAcase, POase, and/or *PON1 Q192R* genotype (Models 1–6). Carriers of high levels of anti-*N*-Hcy autoantibodies or fatty acids had a lower SDMT\_2 score indicating worse cognition.

#### *B vitamin treatment abrogates effects of PON1 measures on cognition*

Effects of B vitamin treatment on PhAcase, POase, and neuropsychological measures of cognition at the end of study are shown in Table 9. In the B vitamin group, there was a small but significant decrease in PhAcase activity at the end-of-study in the B vitamin group from  $0.497 \pm 0.143$  units at baseline to  $0.468 \pm 0.117$  units at the end-of-study after a 2-year-long treatment ( $p=0.003$ ). A similar decrease in PhAcase activity was also observed in the placebo group, from  $0.481 \pm 0.132$  units at baseline to  $0.448 \pm 0.121$  units at the end of study ( $p=2.E-5$ ). In contrast, there was no change in POase activity both in the B vitamin and placebo groups at the end of study. These findings indicate that PhAcase and POase activities were not affected by the B vitamin treatment.

The efficacy of the B vitamin treatment was confirmed by tHcy measurements, which showed a significant reduction in tHcy from  $11.8 \pm 3.4 \mu\text{M}$  at baseline to  $8.9 \pm 2.2 \mu\text{M}$  at the end-of-study in the B vitamin group ( $p=7.E-13$ ), as previously described [5]. In contrast, in the placebo group, tHcy tended to increase from  $12.1 \pm 4.1 \mu\text{M}$  at baseline to  $13.1 \pm 4.7 \mu\text{M}$  ( $p=0.112$ ).

Table 6  
Determinants of verbal episodic memory at the end of study: LnHVL-DR\_2, placebo group\*

Variable (n = 82–112)	Pearson correlation		Multiple regression <sup>#</sup>											
	β	p	Model 1		Model 2		Model 3		Model 4		Model 5		Model 6	
			β	p	β	p	β	p	β	p	β	p	β	p
LntHcy_1	-0.37	0.000												
LnPOase_1	-0.04	0.710							-0.54	0.053	-0.16	0.192		
<b>LnPhAcase_1</b>	0.01	0.942	-0.32	<b>0.012</b>	-0.29	<b>0.015</b>								
<i>PONI-Q192R</i>	0.00	0.998	-0.14	0.231			-0.05	0.655	0.40	0.125				
LnFe_1	-0.19	0.090												
LnAtrophy_rate	-0.30	0.008	-0.24	0.053	-0.21	0.054	-0.22	0.089	-0.26	<b>0.047</b>	-0.24	0.068	-0.22	0.087
LnHVL-DR_1	0.55	0.000	0.46	<b>0.000</b>	0.44	<b>0.001</b>	0.38	<b>0.004</b>	0.41	<b>0.002</b>	0.40	<b>0.002</b>	0.38	<b>0.003</b>
<i>APOE</i>	-0.21	0.038												
<i>BDNF V66M</i>	0.05	0.641	0.31	<b>0.010</b>	0.29	<b>0.015</b>	0.31	<b>0.015</b>	0.38	0.004	0.36	0.005	0.31	<b>0.010</b>
Age	-0.33	0.000												
Sex	0.14	0.167												
			F = 4.68, p = 0.000, Adjusted R <sup>2</sup> = 0.37		F = 5.23, p = 0.000, Adjusted R <sup>2</sup> = 0.37		F = 3.93, p = 0.000, Adjusted R <sup>2</sup> = 0.29		F = 4.14, p = 0.000, Adjusted R <sup>2</sup> = 0.34		F = 4.41, p = 0.000, Adjusted R <sup>2</sup> = 0.32;		F = 4.93, p = 0.000, Adjusted R <sup>2</sup> = 0.32	

\*Ln, natural logarithm; PhAcase, phenylacetate hydrolase; POase, paraoxonase; \_1, baseline; \_2, end of study. <sup>#</sup>Adjusted for sex, LnAnti-N-Hcy\_1, LntHcy\_1, and *APOE* genotype. LntHcy\_1 was not significant in any of the models.

Table 7  
Determinants of attention/processing speed at the end of study: LnTrail\_making\_A.2, placebo group\*

Variable (n = 78–110)	Pearson correlation		Multiple regression <sup>#</sup>											
			Model 1		Model 2		Model 3		Model 4		Model 5		Model 6	
	β	p	β	p	β	p	β	p	β	p	β	p	β	p
<b>LnHcy-.1</b>	0.38	0.000	0.32	<b>0.008</b>	0.32	<b>0.008</b>	0.31	<b>0.014</b>	0.31	<b>0.014</b>			0.26	<b>0.028</b>
LnPOase.1	0.16	0.120							0.16	0.453	0.14	0.148		
<b>LnPhAcase.1</b>	0.14	0.163	0.24	<b>0.015</b>	0.19	0.055								
<i>PONI-Q192R</i>	0.08	0.434	0.19	<b>0.049</b>			0.13	0.193	-0.02	0.295				
<b>LnAnti-N-Hcy.1</b>	-0.02	0.877	0.24	<b>0.025</b>	0.27	<b>0.011</b>	0.26	<b>0.020</b>	0.24	<b>0.028</b>	0.25	<b>0.025</b>	0.28	<b>0.011</b>
LnAtrophy_rate	0.34	0.002	0.30	<b>0.007</b>	0.30	<b>0.007</b>	0.28	<b>0.013</b>	0.29	<b>0.013</b>	0.29	<b>0.010</b>	0.29	<b>0.011</b>
LnTrail_making_A_.1	0.52	0.000	0.32	<b>0.001</b>	0.32	<b>0.001</b>	0.29	<b>0.003</b>	0.29	<b>0.003</b>	0.28	<b>0.004</b>	0.30	<b>0.003</b>
Age	0.37	0.000	0.31	<b>0.005</b>	0.31	<b>0.005</b>	0.32	<b>0.005</b>	0.32	<b>0.006</b>	0.32	<b>0.006</b>	0.32	<b>0.005</b>
			F = 6.70, p = 0.000, Adjusted R <sup>2</sup> = 0.57		F = 6.53, p = 0.000, Adjusted R <sup>2</sup> = 0.54		F = 6.05, p = 0.000, Adjusted R <sup>2</sup> = 0.52		F = 5.61, p = 0.000, Adjusted R <sup>2</sup> = 0.51		F = 6.20, p = 0.000, Adjusted R <sup>2</sup> = 0.52;		F = 6.59, p = 0.000, Adjusted R <sup>2</sup> = 0.52	

\*Ln, natural logarithm; PhAcase, phenylacetate hydrolase; POase, paraoxonase; .1, baseline; .2, end of study. <sup>#</sup> Adjusted for sex, LnFA.1, LnTG.1, *COMT V158M* and *DHFR 19bpins* genotypes.

Table 8  
Determinants of attention/processing speed at the end of study: LnSDMT.2, placebo group

Variable (n = 78–109)	Pearson correlation		Multiple regression <sup>#</sup>											
			Model 1		Model 2		Model 3		Model 4		Model 5		Model 6	
	β	p	β	p	β	p	β	p	β	p	β	p	β	p
<b>LnHcy-.1</b>	-0.31	0.001	-0.20	<b>0.013</b>	-0.19	<b>0.013</b>	-0.19	<b>0.025</b>	-0.19	<b>0.019</b>	-0.19	<b>0.022</b>	-0.18	<b>0.026</b>
LnPOase.1	-0.17	0.107							-0.33	<b>0.028</b>	-0.06	0.336		
<b>LnPhAcase.1</b>	-0.14	0.162	-0.18	<b>0.008</b>	-0.16	<b>0.011</b>								
<i>PONI-Q192R</i>	-0.04	0.702	-0.06	0.417			0.00	0.963	0.29	<b>0.047</b>				
<b>LnFA.1</b>	-0.14	0.157	-0.20	<b>0.020</b>	-0.20	<b>0.017</b>	-0.21	<b>0.020</b>	-0.19	<b>0.032</b>	-0.24	<b>0.005</b>	-0.24	<b>0.006</b>
<b>LnAnti-N-Hcy.1</b>	-0.01	0.912	-0.15	<b>0.041</b>	-0.15	<b>0.027</b>	-0.17	<b>0.025</b>	-0.14	<b>0.047</b>	-0.15	<b>0.036</b>	-0.17	<b>0.017</b>
LnAtrophy_rate	-0.35	0.002	-0.24	<b>0.001</b>	-0.24	<b>0.001</b>	-0.22	<b>0.005</b>	-0.23	<b>0.002</b>	-0.21	<b>0.005</b>	-0.22	<b>0.004</b>
LnSDMT_.1	0.80	0.000	0.65	<b>0.000</b>	0.67	<b>0.000</b>	0.67	<b>0.000</b>	0.65	<b>0.000</b>	0.67	<b>0.000</b>	0.68	<b>0.000</b>
Age	-0.35	0.000												
			F = 23.20, p = 0.000, Adjusted R <sup>2</sup> = 0.78		F = 27.39, p = 0.000, Adjusted R <sup>2</sup> = 0.79		F = 22.18, p = 0.000, Adjusted R <sup>2</sup> = 0.76		F = 22.04, p = 0.000, Adjusted R <sup>2</sup> = 0.78		F = 24.03, p = 0.000, Adjusted R <sup>2</sup> = 0.77;		F = 27.23, p = 0.000, Adjusted R <sup>2</sup> = 0.77	

\*Ln, natural logarithm; PhAcase, phenylacetate hydrolase; POase, paraoxonase; .1, baseline; .2, end of study. <sup>#</sup> Adjusted for sex, and LnTG.1.

The B vitamin treatment improved neuropsychological test scores in several domains of cognition. For example, a significant decrease in global cognition observed in the placebo group (MMSE score:  $27.7 \pm 2.3$  at the-end-of-study versus  $28.2 \pm 1.5$  at baseline,  $p=0.029$ ) was abrogated by the B vitamin treatment ( $27.8 \pm 2.2$  versus  $28.1 \pm 1.8$ ,  $p=0.224$ ). Similarly, there was a significant decrease in verbal episodic memory (HVLT-DR score) in the placebo group from  $7.4 \pm 3.2$  at baseline to  $6.9 \pm 3.6$  at the end of study ( $p=0.046$ ) that was prevented by the B vitamin treatment ( $7.7 \pm 2.9$  versus  $7.4 \pm 3.5$ ,  $p=0.199$ ). A significant decrease in attention/processing speed observed in the placebo group (SDMT score:  $35.4 \pm 11.0$  at the-end-of-study versus  $37.1 \pm 9.9$  at baseline,  $p=0.002$ ) was abrogated by the B vitamin treatment ( $37.4 \pm 11.0$  versus  $37.9 \pm 10.0$ ,  $p=0.214$ ) and the SDMT score tended to be improved in the B-vitamin group at the end of study ( $37.4 \pm 11.0$  versus  $35.4 \pm 11.0$ ,  $p=0.064$ ) (Table 9).

TICS-m score, a measure of global cognition/memory, was significantly increased both in the B vitamin ( $26.9 \pm 5.0$  versus  $24.8 \pm 2.9$ ,  $p=0.003$ ) and placebo groups ( $26.5 \pm 4.4$  versus  $25.0 \pm 2.7$ ,  $p=0.002$ ) at the end of study (Table 9). However, analysis of male and female subgroups showed that the effect of B vitamin treatment on cognition (TICS-m score) was sex-dependent. Specifically, in the male subgroup at the end-of-study, the TICS-m score was increased in the B vitamin-treated ( $26.4 \pm 3.9$  versus  $24.5 \pm 2.8$ ,  $p=0.013$ ) but not in the placebo-treated ( $25.9 \pm 5.3$  versus  $25.2 \pm 2.9$ ,  $p=0.445$ ) group. In contrast, in the female subgroup, the TICS-m score was increased both in the B vitamin group ( $26.6 \pm 4.6$  versus  $24.8 \pm 2.9$ ,  $p=0.005$ ) and in the placebo group ( $26.7 \pm 3.5$  versus  $24.8 \pm 2.6$ ,  $p=0.0002$ ).

Other neuropsychological test scores such as HVLT-TR (verbal episodic memory), Trail Making A and Map Search (attention/processing speed), and Category Fluency (semantic memory), did not differ between the end-of-study and baseline both in the B vitamin and placebo groups.

Multiple regression analysis for the B vitamin treatment group revealed that the treatment abrogated associations between baseline PhAcase and neuropsychological measures in several domains of cognition: global cognition (MMSE, TICS-m), verbal episodic memory (HVLT-TR, HVLT-DR), and attention/processing speed (Trail Making A, SDMT) at the end of study (Table 10). Associations between baseline POase, *PON1 Q192R* genotype, and SDMT\_2

score observed in the placebo group (Table 8) were abrogated in the B vitamin treatment group (Table 10). Similarly, the association between *PON1 Q192R* genotype and Trail Making A\_2 score observed in the placebo group (Table 7) was abrogated in the B vitamin treatment group (Table 10).

Associations between tHcy and cognition (MMSE, TICS-m, and Trail Making A test scores) as well as associations between anti-N-Hcy autoantibodies and cognition (TICS-m and SDMT test scores), observed in the placebo group (Tables 4 and 8), were absent in the B vitamin treatment group (Table 10). Similarly, an association between iron and global cognition (MMSE score), observed in the placebo group (Table 3, was abrogated by the B vitamin treatment; Table 10). The associations of brain atrophy rate with these cognitive measures, observed in the placebo group, were also abrogated by the B vitamin treatment. For each neuropsychological test at the end of study, adjusted  $R^2$  values were reduced by 23 to 55% in the B vitamin group compared to the placebo group. The only variable that remained significantly associated with a neuropsychological test score in the B vitamin group at the end of study was a corresponding neuropsychological test score at baseline (Table 10).

#### *Baseline PON1 measures not associated with brain atrophy rate at the end of study*

To determine whether *PON1* measures could affect brain atrophy, we carried out multiple regression analysis in models including brain atrophy rate as a dependent variable and baseline PhAcase, POase, or *PON1 Q192R* genotype, iron, tHcy, age, sex, and/or brain volume as independent variables.

In the placebo group at the end of study, we found that PhAcase ( $\beta=-0.13$ ,  $p=0.263$ ;  $R^2=0.21$ ), POase ( $\beta=-0.04$ ,  $p=0.712$ ;  $R^2=0.24$ ), and *PON1 Q192R* genotype ( $\beta=0.01$ ,  $p=0.961$ ;  $R^2=0.19$ ) were not associated with the brain atrophy rate, regardless of absence or presence of iron in the examined models. These models became non-significant in the absence of tHcy. As previously reported [5], tHcy was associated with the brain atrophy rate ( $\beta=0.45$ ,  $p=0.000$ ); overall  $p$  values were  $<0.005$ .

In the B vitamin group at the end of study, baseline PhAcase, POase, or *PON1 Q192R* genotype were also not associated with the brain atrophy rate, while the dependence of the brain atrophy rate on baseline tHcy was abrogated by the B vitamin treatment, as previously reported [5].

Table 9  
PhAcase, POase, and neuropsychological measures of cognition at baseline and the end of study

Variable (n = 97–133)	Mean±SD at baseline		Mean±SD at the end-of-study (EOS)		p B-vit. versus placebo group <sup>#</sup>		p EOS versus baseline <sup>#</sup>	
	Placebo group	B-vitamin group	Placebo group	B-vitamin group	Baseline	EOS	Placebo group	B-vit. group
PhAcase <sup>§</sup> , units	0.481 ± 0.132	0.497 ± 0.143	0.448 ± 0.121	0.468 ± 0.117	0.400	0.246	<b>2.E–5*</b>	<b>0.003*</b>
POase, unitsX100	10.4 ± 6.3	9.3 ± 7.0	10.2 ± 5.8	9.5 ± 6.6	0.256	0.425	0.235*	0.327*
tHcy, µM	12.1 ± 4.1	11.8 ± 3.4	13.1 ± 4.7	8.9 ± 2.2	0.503	<b>1.E–14</b>	0.112	<b>7.E–13</b>
MMSE	28.2 ± 1.5	28.1 ± 1.8	27.7 ± 2.3	27.8 ± 2.2	0.750	0.626	<b>0.029</b>	0.224
TICS-m <sup>§</sup>	25.0 ± 2.7	24.8 ± 2.9	26.5 ± 4.4	26.9 ± 5.0	0.540	0.722	<b>0.009</b>	<b>0.004</b>
HVLT-TR <sup>§</sup>	23.0 ± 5.1	23.5 ± 5.4	22.9 ± 5.8	23.3 ± 6.7	0.592	0.918	<b>0.002*</b>	<b>0.003*</b>
HVLT-DR	7.4 ± 3.2	7.7 ± 2.9	6.9 ± 3.6	7.4 ± 3.5	0.241	0.303	0.586*	0.052*
Trail Making A	53.1 ± 24.5	51.0 ± 19.0	56.7 ± 41.6	52.4 ± 39.8	0.603	0.255	<b>0.046*</b>	0.199*
SDMT <sup>§</sup>	37.1 ± 9.9	37.9 ± 10.0	35.4 ± 11.0	37.4 ± 11.0	0.312	0.064	0.697*	0.694*
Category Fluency	19.8 ± 5.1	20.3 ± 4.9	19.3 ± 5.8	20.1 ± 4.8	0.451	0.161	<b>0.002*</b>	0.214*
Map Search	53.4 ± 14.9	53.0 ± 13.6	53.4 ± 17.7	54.6 ± 16.5	0.799	0.506	0.252*	0.233*
							0.909*	0.577*

<sup>§</sup>Normally distributed data. <sup>#</sup>p-values were derived from Log-transformed values: two-sided T-test except where noted; \*paired T-test.

Table 10  
Determinants of cognition at the end of study – B vitamin group\*

Variable (n = 82–112)	Global cognition				Episodic memory				Attention/processing speed					
	LnMMSE_2 <sup>1</sup>		LnTICSm_2 <sup>2</sup>		LnHVLT-TR_2 <sup>3</sup>		LnHVLT-DR_2 <sup>4</sup>		LnTrail_Making_A_2 <sup>5</sup>		LnSDMT_2 <sup>6</sup>		LnSDMT_2 <sup>7</sup>	
	β	p	β	p	β	p	β	p	β	p	β	p	β	p
LntHcy_1	-0.03	0.798	-0.02	0.859					-0.02	0.873	-0.21	<b>0.024</b>	-0.18	0.062
LnPOase_1													0.38	0.094
LnPhAcase_1	0.09	0.569	0.24	0.056	-0.02	0.843	0.14	0.306	0.16	0.288	0.13	0.184		
<i>PONI-Q192R</i>									0.05	0.686			-0.31	0.160
LnTG_1	0.03	0.792									0.23	<b>0.046</b>	0.20	0.079
LnFA_1											0.06	0.632	0.01	0.952
LnFe_1	0.17	0.201												
LnAnti-N-Hcy_1			0.00	0.981					0.06	0.632	-0.18	0.051	-0.14	0.143
<i>BDNF V66M</i>					-0.01	0.916	-0.03	0.826						
LnAtrophy_rate	-0.16	0.182	-0.13	0.259	-0.08	0.479	0.03	0.826	0.12	0.318	-0.09	0.308	-0.07	0.442
LnMMSE_1	0.52	<b>0.000</b>												
LnTICS-m_1			0.38	<b>0.002</b>										
LnHVLT-TR_1					0.63	<b>0.000</b>								
LnHVLT-DR_1							0.47	<b>0.001</b>						
LnTrail Making_1									0.62	<b>0.000</b>				
LnSDMT_1											0.65	<b>0.000</b>	0.66	<b>0.000</b>
	F = 2.44, p = 0.000, Adjusted R <sup>2</sup> = 0.20		F = 4.44, p = 0.000, Adjusted R <sup>2</sup> = 0.28		F = 6.37, p = 0.000, Adjusted R <sup>2</sup> = 0.38		F = 2.64, p = 0.000, Adjusted R <sup>2</sup> = 0.20		F = 2.81, p = 0.000, Adjusted R <sup>2</sup> = 0.30;		F = 11.03, p = 0.000, Adjusted R <sup>2</sup> = 0.61		F = 9.26, Adjusted R <sup>2</sup> = 0.60	

\*Ln, natural logarithm; PhAcase, phenylacetate hydrolase; POase, paraoxonase; TG, triglycerides; FA, fatty acids; \_1, baseline; \_2, end of study. <sup>1–7</sup>Adjusted for sex, age. Additional adjustment for: <sup>1,4</sup> Anti-N-Hcy, tHcy\_1; <sup>1</sup> *BDNF V66M* genotype; <sup>3</sup> Creatinine, *TCN 776CG* genotype; <sup>4</sup> *APOE* genotype; <sup>5</sup> Fe\_1, FA\_1, TG\_1, *COMT V158M* and *DHFR 19bpins* genotypes.

## DISCUSSION

The present prospective study shows that PON1 is an important predictor of cognition in MCI patients and that B vitamins ameliorate detrimental effects of PON1 on cognition. Specifically, we found that baseline PhAcase activity was a stronger predictor of cognition than POase activity or *PON1 Q192R* genotype and that baseline PhAcase activity affected cognition in domains that were not affected by POase activity or *PON1 Q192R* genotype at the end of study two years later. For example, PhAcase activity predicted global cognition (MMSE and TICS-m scores), verbal episodic memory (HVL-TR and HVL-DR scores), and attention/processing speed (SDMT and Trail Making A scores) at the end of study.

In contrast, baseline POase activity predicted cognition only in the attention/processing speed domain (SDMT score) as did *PON1 Q192R* genotype (SDMT score, Trail Making A score).

These findings suggest that some aspects of PON1 status, such as PhAcase activity, can affect several domains of cognition while other aspects, such as POase activity or *PON1 Q192R* genotype, can affect cognition only in domains that were affected by PhAcase activity. Because PhAcase activity reflects PON1 protein levels [22] and *PON1 Q192R* genotype has no effect on the PON1 protein concentration [21, 46], the predominant associations of cognition with PhAcase identified in the present study suggest that the level of PON1 protein expression is the relevant predictor of cognition (or determines these effects on cognition).

Although baseline PON1 variables predicted global cognition (MMSE, TICS-m scores), verbal episodic memory (HVL-TR and HVL-DR scores), attention/processing speed (Trail Making A score), and attention/processing speed (SDMT score) at the end of study, the PON1 variables did not predict the rate of brain atrophy. The lack of PON1 effect on brain atrophy suggests that PON1 affects functional (cognitive performance), but not structural aspects of cognition (brain atrophy). The mechanism by which PON1 affects cognition remains to be examined in future studies.

Previous findings showed that B vitamin treatment was beneficial by reducing the rate of brain atrophy [5]. In the present study we found that B vitamin treatment can also be beneficial by abrogating the detrimental effects of PON1 on cognition. At the same time, B vitamin treatment eliminated (or reduced) the detrimental effect of brain atrophy on

cognition observed in the placebo group at the end of study.

In many cases genes have multiple functions that differ depending on a tissue. PON1 possesses several enzymatic activities and has been implicated in cardiovascular and neurodegenerative diseases, suggesting that its function is tissue-dependent. Our present study provides support for this notion. Specifically, we found that serum PON1 PhAcase activity was a *negative* predictor of cognition (i.e., high PON1 activity was associated with worse cognition) while *PON1 Q192R* genotype and POase activity had a marginal, if any, effect on cognition. Further, we found that the status of PON1 was not associated with brain atrophy. In contrast, previous prospective study found that the cardiovascular risk was significantly reduced in individuals with the highest PON1 activity (PhAcase and POase) and *PON1 192RR* genotype versus those with the lowest PON1 activity and *PON1 192QQ* genotype (i.e., high PON1 activity was associated with better outcome) [47]. Taken together, these findings suggest that PON1 has different functions in the central nervous system (CNS) and in the cardiovascular system.

How higher PON1 PhAcase activity could lead to impairments in cognition is not clear. In the cardiovascular system PON1 activity was negatively associated with indices of systemic oxidative stress such as multiple oxidized fatty acids [47]. Because PON1 does not possess any intrinsic redox activity (PhAcase and POase are hydrolytic, not redox, activities) and indeed does not protect low-density lipoprotein from oxidation [48–50], such associations could be due to the interaction between *PON1* genotype/activity and the expression of redox-related proteins [51]. Thus, detrimental effects of PON1 on cognition observed in the present study could be mediated by effects of PON1 on the expression of proteins maintaining brain homeostasis, which remains to be examined in future studies, for example in a *Pon1*-KO mouse model.

In addition to PON1, we identified iron, triglycerides, and fatty acids, and confirmed anti-*N*-Hcy autoantibodies as predictors of cognition in MCI. Specifically, iron and triglycerides predicted global cognition (MMSE score); anti-*N*-Hcy autoantibodies and fatty acids predicted attention/processing speed (SDMT score); and anti-*N*-Hcy autoantibodies predicted global cognition (TICS-m score) and attention/processing speed (SDMT and Trail Making A scores). Iron and other metals [52], triglycerides and fatty acids [53] have been postulated to affect

cognition in AD. The association of anti-*N*-Hcy autoantibodies with cognition in MCI has been previously reported [42]. Value of these variables as predictors of cognitive decline in MCI patients need to be confirmed in larger studies. Mechanisms underlying these associations remain to be investigated in future studies.

We also identified *BDNF V66M* genotype as a predictor of cognition in MCI. Specifically, *BDNF V66M* genotype predicted verbal episodic memory (HVL-TR and HVL-DR scores) with carriers of *BDNF 66V* allele showing a better cognition than carriers of *BDNF 66M* allele. This finding is consistent with findings in patients with schizophrenia as well as in a control population showing impaired episodic memory in Wechsler Memory Scale test (lower score) in individuals with *BDNF 66M* allele [54].

In conclusion, our findings in participants with MCI provide the first experimental evidence suggesting that *PON1* could play a role in the CNS by affecting cognition in the general domain (global cognition) as well as in more specific domains such as verbal episodic memory and attention/processing speed. These findings also support a concept that *PON1* is a risk factor for cognitive decline in MCI that can be abrogated by B vitamins, thereby highlighting a novel positive aspect of B vitamin treatment on the CNS.

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