Review

Alzheimer's Disease, Oligomers, and Inflammation

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Abstract. The production of soluble amyloid- β oligomers (A β Os) and the activation of inflammation are two important early steps in the pathogenesis of Alzheimer's disease (AD). The central role of oligomers as responsible for the neuronal dysfunction associated with the clinical features has been extended to the other protein misfolding disorders definable, on this basis, as oligomeropathies. In AD, recent evidence indicates that the mechanism of inflammation as a consequence of neurodegeneration must be assessed in favor of a more direct role of glial activation in the alteration of synaptic function. Our own experimental models demonstrate the efficacy of anti-inflammatory treatments in preventing the cognitive deficits induced acutely by A β Os applied directly in the brain. Moreover, some promising clinical tools are based on immunological activation reducing the presence of cerebral A β deposits. However, the strategies based on the control of inflammatory factors as well as the amyloid aggregation show poor or non-therapeutic efficacy. Numerous studies have examined inflammatory factors in biological fluids as possible markers of the neuroinflammation in AD. In some cases, altered levels of cytokines or other inflammatory markers in cerebrospinal fluid correlate with the severity of the disease. Here we propose, according to the precision medicine principles, innovative therapeutic approaches to AD based on the patient's inflammatory profile/state. The earlier intervention and a multifactor approach are two other elements considered essential to improve the chances of effective therapy in AD.

Keywords: Alzheimer's disease therapy, amyloid, anti-inflammatory drugs, glial cells, immune system, oligomeropathy, precision medicine, toll-like receptors

INTRODUCTION

In Alzheimer's disease (AD), together with the specific neuropathological features, cerebral amyloid deposits and neurofibrillary tangles, the glial reactivity testify to the involvement of inflammation in the pathological process [1]. However, the loss of synapses is the structural change that correlates best with the cognitive decline in AD patients [2]. The neuronal dysfunction induced by soluble small aggregates of amyloid- β oligomers (A β Os)—considered the real toxic species in AD [3, 4]—might explain

the positive correlation of the total cerebral $A\beta$ burden with clinical features [5]; the clinical correlation no longer holds when limited to the amyloid plaque distribution, and is only partial when associated with tangle distribution [6].

According to the latest version of the amyloid cascade hypothesis [7], the pathological scenario of AD identifies the soluble small aggregates of Aβ, AβOs, as the main species responsible for the neuronal dysfunction [8, 9], and this concept has been extended to all the other protein misfolding disorders. We have coined the term *oligomeropathy* to illustrate the close relationship between neurodegeneration and these heterogeneous formations of soluble aggregates [10, 11]. The mechanism of disease spreading described in AD models, but also proposed in Parkinson's

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disease or frontotemporal dementia, might employ the oligomers as the species responsible for cell-to-cell transmission [12, 13]. Although several aspects of this mechanism are still elusive, the intercellular passage might employ small aggregates to spread the pathology [14, 15]. Penetration of the oligomers into the cells could be associated with a seeding mechanism and prion-like replication, followed by their release by themselves or combined with vesicle structures, such as exosomes [16]. Thus, the oligomeric structures might play a central role for both neuronal dysfunction and disease propagation.

Gliosis is generally considered a consequence of amyloid deposition and neurodegeneration, activating the production of toxic factors in a vicious circle that amplifies the neuronal dysfunction. This concept has been recently flanked by a more direct influence of inflammatory factors in the early phase of the pathological process [17]. Physiological role of surveillance exerted by microglia and astrocytes for the presence of pathogens and cellular debris is combined with the synaptic modeling, formation, and elimination during development, but also in adulthood [18–20]. This later activity is an important element in favor of the early participation of glial cells in AD pathogenesis.

The intimate contact between microglia and astrocytes with synapses has been investigated from the functional point of view, inspiring several hypotheses on the positive or detrimental roles of immune factors in physiological and pathological aging. It is very possible that the alteration of the glial cells dynamic relationship with synaptic structures and function contributes with the aberrant pruning to the neuronal dysfunction mechanisms [21–22].

The genetic evidence also helps support an essential, rather than secondary role, of the inflammation and several genes involved in the immune pathway have been associated with AD. In particular, TREM2 (triggering receptor expressed on myeloid cells 2) and CD33 polymorphisms have been recently linked to the risk of AD [23–26]. The expression of these factors in the brain is restricted to microglial cells, indicating the direct involvement of these cells in AD pathogenesis.

AD AS AN OLIGOMEROPATHY

In vitro models proposed a direct neurotoxic effect of $A\beta$ at the beginning of the 1990s [27]; the effect was initially related to the fibrillogenic capacity of

A β peptides [28, 29]. The critical role of oligomeric forms in the pathogenesis of AD emerged from observations that, in contrast with the common belief at that time, the fibrillogenic capacity of the A β peptides did not always correlate with their toxicity [30, 31]. The soluble species of A β aggregates were purified from AD brain and CSF and induced neuronal dysfunction in experimental conditions [3].

The origin and size of the ABO species involved in the neurodegenerative mechanisms and the cellular pathways mediating their effects have been widely debated [32, 33]. Although numerous biological alterations are associated with their presence, their biological activities can be triggered essentially by three mechanisms: the interaction with specific entities on cell membranes [34]; the unspecific membrane perturbation [35] and the capacity to form a pore channel inside the membrane [32]. These basic mechanisms involving oxidative stress, mitochondrial alterations, glial activation, and glutamate receptors are common to virtually all oligomers, regardless of the initial misfolded protein sequence [36-38]; other more specific aspects are associated with sequence, size and conformation [13].

In an AβO preparation, species of the same size can be found that are neither toxic nor recognized by oligomer-specific antibodies, indicating that ABOs may also show size-independent differences in toxicity [39], and differences between two ABOs of similar size and dissimilar toxicity may be seen [40]. These studies suggest that the toxicity of small amyloidogenic oligomers is governed primarily by the degree of solvent exposure of hydrophobic residues and is weakly influenced by their secondary structures [41]. Chaperones play an important part not only in the oligomer conformation and assembling but also in their biological interaction. Chaperons bind to protein oligomers, interfering with amyloid fibril formation, but can also directly inhibit the toxicity of these species [42]. Considering the size of oligomers and their hydrophobic exposure as the main determinants of their neurotoxicity [43, 44], molecular chaperons increase the size of the oligomers and mask hydrophobic patches exposed on their surface [45].

The main components of senile plaques and oligomers are the peptides $A\beta_{1-40}$ and $A\beta_{1-42}$. Both have self-aggregation capacity but with different kinetics; the longer sequence can spontaneously aggregate within minutes, while in similar conditions $A\beta_{1-40}$ takes hours and days to assemble in fibrils. Once the $A\beta$ Os are formed, the biological effects are

relatively independent from the initial sequences [4]. Other $A\beta$ peptide species in the brain are $A\beta_{1-43}$, and $A\beta_{1-37}$ or $A\beta_{1-38}$, and although most part of the experimental studies have been used using $A\beta_{1-42}$ solutions, the natural substrate of oligomers is a mixture of peptides.

The N-terminally truncated pyroglutamylated form of $A\beta$ ($A\beta pE$), also identified in AD brain with high self-aggregation capacity, has been indicated as the seed of nucleation of the $A\beta_{1-42}$ solution [46–48]. The synergistic effect of $A\beta/A\beta pE$ hetero-oligomers results in a species with a high level of toxicity, stabilizing the oligomeric structures and retarding fibril formation [49]. Active and passive immunization based on $A\beta pE$ have given beneficial effects in experimental models of AD [50, 51].

Amyloid deposition is considered an early pathological event in AD [52]; as recently indicated by a meta-analysis, high levels of Aβ were associated with a small cognitive impairment and decline in cognitively normal older adults, suggesting a possible dementia prodromal condition in these subjects [53]. However, the lack of correlation between amyloid plaque distribution and the cognitive decline argues against a direct pathogenic role of AB deposits in AD. Furthermore, the amyloid plaques do not correlate with the clinical progression of the disease: after the clinical onset, the AB plaque burden was substantially stable [54]. Positron emission tomography (PET) analysis in longitudinal studies showed that in mild cognitive impairment (MCI), an AD prodromal condition, and in AD subjects the cerebral fibrillar AB levels reached a plateau early and remained unchanged during the disease progression. Structural changes detected by MRI and cognitive impairment, measured by neuropsychological tests, proceed independently of amyloid accumulation [55]. The presence and the toxic role of ABOs might explain these findings: the early neuronal dysfunction in the preclinical phase of AD is due to the soluble aggregates released by the AB plaques.

The burden of $A\beta Os$, which unfortunately are not visualized by PET probes, is generally associated with high levels of fibrillar $A\beta$; the senile plaques are considered to be in dynamic equilibrium with the oligomers, and sequester soluble aggregates which are then released from the deposits [30]. However, in a small percentage of subjects the development of AD is not strictly related to the high level of fibrillar $A\beta$ in the preclinical phase; in this case the production of $A\beta Os$ is independent of the accumulation of

A β plaques [56]. This is possible because the passage from the monomeric form of A β to the insoluble fibrils is not an invariable, linear process that reaches the fibrillar structures through oligomeric and protofibrillar species. In some cases, oligomers did not generate fibrils (off pathway), and the toxicity of these stable products should be analyzed case by case [57, 58]; in other conditions, it has been proposed that oligomers form fibrils through metastable intermediates [59].

In transgenic models, cognitive decline but also synaptic alterations were evident in plaque-free conditions [60]. Single-transgenic mice (APP23) in a pre-plaque stage displayed memory deficits and long-term potentiation impairment, with synaptic hippocampal damage. APP23 tg mice have hippocampal alterations in the trafficking of synaptic NMDAR subunits NR2A and NR2B and ultrastructural analysis shows A β O localized intracellularly in the synaptic compartments. Importantly, the behavioral and biochemical alterations of NMDAR signaling are associated with the inhibition of long-term synaptic potentiation and inversion of metaplasticity at CA1 synapses in hippocampal slices from another transgenic mouse (Tg2576) [61].

Gandy et al. [62] showed that single-transgenic mice (APP E693Q) present a significant oligomerdependent delay in acquisition of the Morris water maze task at 12 months of age, which it is not dependent on the development of AD-like plaque pathology or macrohemorrhage. The presence of ABOs in the pre-plaque period of APP tg mice and in other transgenic models that never develop AB plaques has been proved and might explain the pathological alterations in the absence of the fibrillar AB formations. Although other hypotheses have been put forward to explain the neuronal dysfunction independent of the plaques, i.e., imbalance of amyloid-β protein precursor (ABPP) metabolism [63], or intraneuronal accumulation of ABPP [64], a specific approach depleting ABOs affected the neuronal dysfunction observed in plaque-free animals [65].

In humans, the diffuse presence of A β Os may possibly precede or follow the formation of plaques; the variable production of toxic species and the neuronal vulnerability can differentiate the dynamic of the pathological process and the functional consequences. The recent distinction between the toxicity of oligomers according to their aggregation size and the bidirectional connections between the large and small oligomers confirmed the complexity and the non-linear trend of A β Os formation [66].

We recently developed a simple method to analyze the effect of ABO in vivo by direct intracerebroventricular (ICV) injection in wild type mice of a lowconcentration solution of AβOs (1 μM) followed by a behavioral test, novel object recognition test (NORT), histological and biochemical analysis [4, 67]. We realize that this approach can cover only a few aspects of AD complexity, but our purpose was to dissect the biological effects of ABOs without any confounding factors. We took particular care about two aspects: the chemicophysical characteristics of the ABOs and the behavioral test. As our own experience and numerous studies indicate, the preparation of ABO as a tricky step that needs to be efficiently controlled to ensure the quality of the product and keep their chemicophysical features constant over time.

We used the synthesis of the water-soluble depsi- $A\beta_{1-42}$ peptide, from which the native sequence is easily obtained in aggregate-free conditions. In basic conditions, the switching of the depsi-peptide means its aggregation can be triggered only when required [67]. AFM analysis of the $A\beta O$ solutions is routinely done to confirm the structural characteristics of the species prepared. NORT is a behavioral test with some undoubted advantages: the procedure is relatively easy, the results can be recorded, and it is based on spontaneous behavior without any training. We have developed the test within the home cage to reduce any interference from anxiety induced by placement in a new space. The animals' motor behavior was constantly monitored.

The memory impairment induced by the A β Os was specific, and A β fibers and monomers were not active, were reversible, and involved glial activation in a complex interaction with neurons. We were able to clarify the potential role of prion protein (PrP) as a ligand of A β Os with no direct functional consequences. Although we confirmed the previous reports of the high-affinity binding of A β Os to PrP, the application of A β Os in PrP/KO mice showed the same memory impairment as in the wild type, arguing against a functional consequence of PrP-A β O interaction [4]. The role of this interaction is still debated [68, 69]. However, our paradigm is adequate to determine the biological consequences of A β O exposure *in vivo*.

INFLAMMATION IN AD

The concept of inflammation in AD and other neurodegenerative disorders describes a particular biological process different from the classical inflammation outside the CNS; although the factors involved are often the same, the timing and the activities are quite distinct [70, 71].

Inflammation within the brain is a typical doubleedged sword: it involves the positive activation of surveillance with phagocytic activity of both microglia and astrocytes to eliminate the debris and the pathogenic elements including protein aggregates, while on the other hand, the glial activation induces the production of factors with harmful consequences for the neuronal system. The neuroinflammation in AD is persistent with no resolution, a sort of chronic reaction of the innate immune system that causes neuronal alterations. The immune response is terminated with removal of the stimulating pathogen, and the resolution is an active process regulated by specialized pro-resolving mediators [72, 73]. This mechanism seems to be altered in AD and there are fewer specialized pro-resolving mediators in autoptic samples [74, 75]. This defect may also possibly be associated with the continued presence of AB aggregates, which aliment the chronic inflammation [76].

In the last few years, the role of inflammation in AD pathogenesis has gained importance with the accumulation of experimental evidence that immune factors can interfere in the early phase of the disease [77, 78]. As mentioned in the introduction, these results are now backed by genetic findings that indicate the immune system, together with lipid metabolism and vesicle traffic, are the main biological pathways where genetic alterations are associated with AD [79]. Not only the polymorphisms TREM2, CD33, or CR1 but several other missense mutations on genes encoding for immune factors have been identified as influencing the risk of developing AD.

The initial condition that indicated the glial reaction as merely a consequence of the neurodegeneration and Aβ deposits has progressively evolved. The activation of glial cells with the production of cytokines, radicals, and other immune factors, not necessarily detrimental, shifts from a secondary phenomenon to become an essential aspect of the pathological condition. Microglia as well as astroglia can model synaptic pruning and function, and in several experimental conditions it has been proven that the loss of glial cells or their alteration clearly influence neuronal function not only during development, but also in adults [18, 80–82]. This does not mean that AD and the other neurodegenerative diseases can be thought of as inflammatory disorders, but we have

to consider that the non-neuronal components are potentially able to induce synaptic loss or damage in the absence of amyloid plaques or neurodegeneration [83], confirming the multifactor characteristics of AD.

In this complex scenario, peripheral conditions like diabetes, obesity, or metabolic syndrome can substantially contribute to creating an inflammatory milieu that facilitates the detrimental effects on neuronal function [84, 85]. Clinical studies have suggested that patients with AD who experienced an infection have accelerated cognitive decline, positively related with peripheral levels of TNFα [86]. Patients suffering from chronic periodontitis are also at higher risk of AD. Plasma concentrations of TNFα and antibodies to periodontal bacteria are higher in patients with AD than in non-demented control subjects and are independently associated with AD [87-89]. This sustained release of proinflammatory cytokines may affect the brain in much the same way as systemic inflammation. The gut microbiota has also been indicated as a possible source of elements that can influence—once again from the inflammatory point of view—AD pathogenesis [90].

TREM2 AND AD

Since genetic evidence become available on the rare mutations of genes encoding TREM2 associated with AD [22], numerous investigations have focused on the biological mechanism responsible for this relationship [91]. TREM2 is expressed in several myeloid lineages but in the CNS, the expression of the receptors is limited to microglial cells [92, 93]. This feature has been considered particularly attractive from the biological point of view because, for the first time, a non-neuronal component is directly involved in AD pathogenesis.

In general, TREMs are a signal relevant to the clearance of cellular debris and pathogens, with no excessive inflammation and tissue destruction. TREM2 is a glycoprotein of 230 amino acids, belonging to the immunoglobulin (Ig) superfamily. On the functional level, TREM2 inhibits cytokine and chemokine expression, acts as a negative regulator of toll-like receptor (TLRs)-mediated response, and promotes microglial phagocytosis [94]. Apparently TREM2 can drive the microglial state from the inflammatory (M1) to anti-inflammatory condition (M2); TREM2 overexpression induces the expression of all the immune markers that

characterize the M2 phenotype like ARG1, IL-10, and IL4 with reparative meaning [95, 96]. Microglial knock-down for TREM2 exposed to apoptotic neurons increased the expression of pro-inflammatory cytokines, but reduced phagocytic activity [99]. Since apoptotic neurons express TREM2-specific ligands, the absence of TREM2 production by microglia would result in the reduction of its phagocytic activity. Mazaheri et al. [97] used a trascriptomic approach to show that the differently expressed messenger RNAs in wild type and TREM2 –/– cells had impairment of the genes involved in chemotaxis, migration, and motility. Their deficit in chemotaxis was confirmed with functional analysis and the defect was rescued with the re-expression of TREM2.

In AD, transgenic mice the expression of TREM2 in the brain was higher than in wild type mice and the action on its expression can influence the AD phenotype [98]. The knock-down of TREM2 or the reduction of its expression increased cerebral $A\beta$ deposition, synaptic loss, and cognitive decline in various AD models [99, 100]. While over-expression of TREM2 resulted in a reduction of AD neuropathology and improved behavior. Only in one specific model (APP/PS1-21) were there positive consequences induced by the TREM2 knock-down [101].

After the initial missense mutation (R47H) identified as a risk factor in AD, a second polymorphism (R62H) was associated with the disease [102–103]; TREM2 variants confer a similar risk for AD as one copy of ApoE ε4 [104]. Apparently the R47H mutation is associated with a reduction of the function of TREM2. The partial deletion of TREM2 in APP23 mice did not affect the number of plaques but reduced the number of microglial cells surrounding the amyloid deposits. Thus the mutation might alter the chemotactic activity of microglial cells toward senile plaques [105]. In addition, these variants might abolish the capacity of microglia to recognize and/or bind specific ligands. New functional data support this view, demonstrating that TREM2 variants may have different expression and either in boost or reduce its ligand binding ability on microglia [106]. Investigation in autoptic material revealed a low level of expression of TREM2 in healthy control brain and a clear increase in the microglia surrounding the senile plaques in AD brains [107]. However, the neuropathological results at the late stages of disease gave limited information on the role of TREM2, which at a certain point may lose its function, depriving microglia of its regulatory surveillance.

In AD-related TREM2, the surface membrane expression of receptors was the same as in control or other pathologies, but the signal has less efficacy [108]. The mutations can alter the folding kinetics, which might imply slower replenishment at the cell surface or the inability to bind properly to their signaling sustainers. Specific investigations have confirmed a small conformational change in the R47H TREM2 mutation but no changes in the R62H one. Investigations on TREM2 binding capacity to its ligands [104] confirmed that the presence of R62H and R47H mutations affected ligand engagement, probably because of a defect in ligand recognition that consequently impairs microglial activity.

In a recent paper, Ulland et al. [109] reported that microglia in AD patients carrying TREM2 risk variants and in TREM2-deficient mice crossed with 5xFAD mice had abundant autophagic vesicles, metabolic alteration, and energy failure, reflecting defective activation of mTOR signaling. Activation of the alternative pathway and signaling with cyclocreatine restored the energy metabolism, reduced the autophagy and improved the viability of TREM2-/macrophages. The treatment of TREM2-/- 5XFAD in vivo by supplementation with cyclocreatine in drinking water restored mTOR signaling and the energy metabolism and reduced the autophagic markers in microglia. Cyclocreatine, while not affecting the total plaque number, significantly reduced plaqueassociated neurite dystrophy, compared to untreated TREM2 -/-5XFAD mice, to the levels observed in 5XFAD mice. The authors suggest that these data indicate that cyclocreatine improves microglial metabolism and boosts the protective response to Aß plagues in TREM2-deficient 5XFAD mice. The functional consequences of the beneficial effects observed on histological examination were not analyzed, so we do not know how cyclocreatine affected the neuronal dysfunction. Furthermore, the complete absence of TREM2 expression and the very aggressive AD model (5XFAD), are extreme experimental conditions, hardly to be compared to AD conditions. However, the data do indicate a precise pathway to improve the neuroprotective activity of microglia.

Another specific target in neurodegenerative conditions was proposed by Keren-Shaul et al. [110], who identified a new type of microglial cell closely involved in neurodegeneration (DAM) with a specific panel of receptor expression. The activity of this subpopulation was beneficial and its activation was partially TREM2-dependent. Apparently DAM is needed to mitigate the disease by supporting

phagocytosis but it occurs at relatively late stages of the disease [111].

CR1 AND AD

Together with TREM2, recent large genome-wide association studies have identified single nucleotide polymorphisms in the C3b/C4b receptor (CR1) [105, 112]. CR1 is a transmembrane protein expressed by astrocytes involved in phagocytosis enhancement by C3b-, C4b-, C1q-, mannose-binding lectin-, and ficolins-opsonized particles. The complement cascades driven by C1 and C3 are both fundamental in brain development and synaptic pruning [113-115], but their overactivation might be detrimental. Recently it has been shown how excessive microglial activation can culminate in significant loss of synapses. Clq protein steeply accumulates at the synapses in normal mouse and human aging brains [116] and is highly upregulated in most or all neurodegenerative diseases, including AD [117] and dementia [118], where complement cascade components have emerged as significant enhancers of synapse vulnerability. Synapse loss was evident in these mice at the ages investigated but prevented, together with a decrease in the phagocytic activity of microglia, by inhibiting C1q, C3, and the complement receptor CR3.

An essential contribution has also been proposed of astrocyte phagocytosis in this mechanism dependent on the ApoE genotype [119]. According to these findings, ApoE isoforms might control the rate at which C1q protein accumulates in the aging brain. ApoE ε2, a protective allele against AD, enhances the phagocytic capacity of astrocytes. The decrease in C1q accumulation in aged knock-in expressing ApoE ε 2 animals supports the hypothesis that ApoE ε 2 helps keep the synaptic environment clean from senescent synapses and aberrant immune reactions. In contrast, ApoE ε4 reduces the overall phagocytic capacity of astrocytes, leading to faster accumulation of senescent synapses and their debris. Similarly, ApoE isoforms may strongly control the rate at which astrocytes and microglia help clear amyloid plaque accumulation [120]. The same authors noted that C1q accumulation at synapses is not enough to trigger their degeneration because another signal is required to activate the complement cascade, culminating in removal of the complement-coated synapses by microglia [121]. The presence of AβOs is sufficient to trigger this cascade. Accumulation of senescent synapses and their debris in the aged brain may induce reactive gliosis and neuroinflammation, with eventual synapse loss—an early pathophysiological aspect of AD. This was confirmed in A β O-treated mice, showing reductions in synaptic markers, though this was no longer detectable when A β Os were injected in C1qa knockout mice, suggesting cooperation between C1q and A β Os to trigger synapse loss via CR3 [121]

INFLAMMATION AND MEMORY

Using our acute experimental model, we showed the early involvement of inflammation in the memory impairment induced by ICV application of AβOs from depsi-peptide. A single ICV injection of ABOs (1 μM) in wild type mice before the initial exposure to the objects in the NORT, nullified the animals' ability to distinguish the familial from the novel object. Similar results were obtained with Aβ₁₋₄₀ or Aβ₁₋₄₂ oligomers [4] and, with a distinct mechanism, oligomers from α-synuclein, the essential component of Lewy bodies in Parkinson's disease and dementia with Lewy bodies, impaired the memory [24]. A toll-like receptor 4 (TLR-4) antagonist completely rescued the ABOs effect and in TLR-4-KO mice AβOs lost their ability to induce memory impairment [122].

In a similar model, the depressive effect associated with the inflammatory reaction induced by ABOs was also abolished in TLR-4-KO mice [123]. The involvement of TLR-4 indicated the possible involvement of inflammation in this acute model confirmed by the protective effect of indomethacin, a classical NSAID: injected intraperitoneally the drug antagonized the amnesic effect of AβOs. Within 24 hours, after the ABO injection transient gliosis was observed with a peak effect at 4-8 hours, quantified by western blot analysis and histopathology of GFAP and Iba-1, markers of astroglia and microglia, respectively, in hippocampus and cortex. With a parallel time course, ABOs also induced the expression of a panel of cytokines, including IL1β, TNFα, IL-6, and IL-10. The essential role of neuroinflammation in this experimental model was confirmed by the antagonism of the memory impairment and gliosis by IL-1RA [122].

The information provided by this relatively simple approach is the intimate relationship between neuronal dysfunction and neuroinflammation, both activated by A β Os through TLR-4, indicating the need for treatments that interfere with different aspects of

the pathological mechanism (Fig. 1). These results are in line with various other findings [123, 124]. An interesting example is the study by Xu et al. [125] where the ABOs from human ABPP-expressing cultured cells or purified from AD brain tissue, applied ICV in wild type mice, induced microglial activation in the hippocampus determined by morphological analysis and by measuring mRNA changes in multiple inflammatory genes. The environmental enrichment, a well-documented approach to induce cognitive and neuronal activation, silenced the innate immune response activated by ABOs. Similar results were obtained by Beauquis et al. [126]. In this case, environmental enrichment prevented astroglial pathological changes in the hippocampus of plaquefree ABPP transgenic mice. Here again it was hard to distinguish the glial activation from the neuronal dysfunction when the brain was exposed to AβOs. PET analysis that combined the identification of Aβ deposits with the microglial activation using 11C-(R)-PK11195 binding in MCI subjects confirmed a positive relation between amyloid load and levels of microglial activation. ¹¹C-(R)-PK11195 binding by PET revealed increased inflammation in a majority (85%) of amyloid-positive MCI cases, its cortical distribution overlapping that of amyloid deposition [79].

THERAPEUTIC OPPORTUNITIES

Despite the accumulation of results in favor of the toxic activity of oligomers and the robust scientific support for the involvement of inflammation in AD pathogenesis, we are still waiting for a disease-modifying drug based on interference with AB aggregation and/or inflammation. According to the Aβ cascade, most of the therapeutic approaches in AD has been aimed at amyloid aggregates or ABPP metabolism, and the develop of antibodies against AB in particular has produced various anti-dementia candidates. Active anti-Aß immunization was proposed at the end of 1990s, on the basis of clear evidence in AD experimental models [127] and the vaccination was clinically tested a few years later. Although the first trial was stopped due to intolerable side effects [128], passive or active immunization is still considered one of the most interesting approaches for AD [129, 130]. This concept is supported by the biological efficacy in reducing the AB burden, proven in experimental models and confirmed in human neuropathological samples [131, 132]. A recent phase II investigation with aducanumab, an antibody targeting

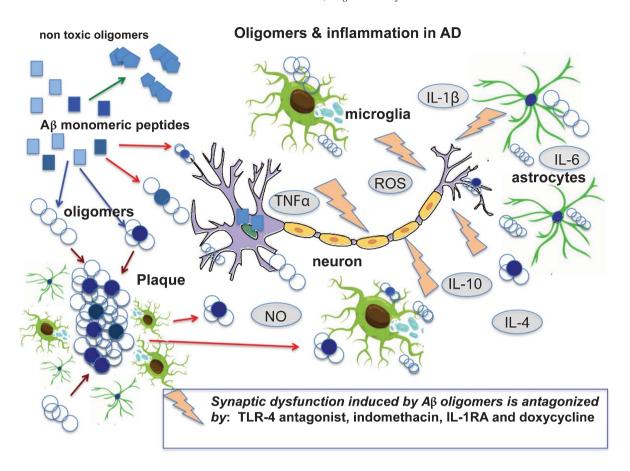


Fig. 1. The cartoon illustrates several aspects of AD pathogenesis starting from the formation of $A\beta$ oligomers, a variety of entities with different sizes, peptide combinations and conformations (in light blue and blue); the seeding passage with conformation change (from square to circle) may also occur intracellularly. The oligomers in dynamic equilibrium with senile plaques can act directly at the neuronal level or induce neuronal dysfunction through glial activation by the production of cytokines radicals and other factors. In our experimental studies [133], we found that anti-inflammatory drugs, doxycyline and a TLR-4 antagonist antagonized the cognitive decline induced by direct intracerebral injection of $A\beta$ oligomers.

aggregated oligomers and fibers, but not monomer AB, gave promising results [133]. This antibody, the last of a long list that have failed to show efficacy in AD, was derived from a lymphocyte library collected from healthy elderly subjects with no signs of cognitive impairment and cognitively impaired elderly people with unusually slow cognitive decline. This human origin might boost the effectiveness and make the difference in favor of this treatment. Currently, aducanumab is in phase III clinical trials that are expected to be completed in 2020. The therapeutic possibilities deriving from the control of inflammation in AD are supported by several epidemiological and observational studies reporting a reduction of the risk of AD in subjects exposed to NSAID in a preclinical period [134]. However, when the investigation moved on from observational studies to a curative approach, virtually all the randomized

controlled trials failed to show any benefit effect with NSAIDs [135, 136]. The availability of numerous anti-inflammatory drugs, mostly various chemical species of COX inhibitors, has favored the design of many studies. When the efficacy was formally investigated in a randomized clinical trial, a few studies founds the anti-inflammatory treatment induced a modest improvement. A positive tendency in favor of the treatment has been obtained with indomethacin, but the small size of the populations studied did not permit any firm conclusion [137, 138]. The side effects of indomethacin do not recommend its prolonged or widespread use, and after a quarter of a century since the initial pilot study reported encouraging results in AD subjects treated with this drug, we still have no final opinion on its efficacy.

More specific reviews focused on therapeutic studies in AD [139, 140] have noted that beside the

complexity of the disease, there are several potential explanations for these unsatisfactory results: identification of the wrong target; selection of the wrong drug; inappropriate design of the trial or treatment schedules; and, finally, insufficient selection of the subjects. Other aspects referring to the natural history of the disease might suggest starting the treatment earlier, in the preclinical phase. In this case, biological markers are needed, to follow the drug's effect and the progress of the disease. In addition, the complex AD pathogenesis will presumably need multitarget therapy [141].

These concepts acquire particular significance in the light of the itemized considerations in this review, since both the accumulation of ABOs and the neuroinflammation are early events in AD pathological process. It is reasonable to assume that at the very outset, the primus movens is the presence of oligomers, released from senile plaques or formed independently. The appearance of ABOs has different biological implications, with different lag periods for each individual the inflammatory cascade is also activated (Fig. 1). The constant presence of ABOs is associated with uncontrolled inflammation, so immune factors and ABOs may well combine deleterious effects on the neuronal system with an external contribution from the periphery. At this point, a single treatment to eliminate the ABOs or an antiinflammatory intervention would be unlikely to give curative results. Thus, to change the trajectory of the disease early intervention on both flanks is necessary. The contribution of inflammation should be monitored by measuring biological markers in the blood or-though less convenient-in the CSF, in combination with the profiles of genes encoding inflammatory factors [142, 143] and analysis of the microbioma. Furthermore, as reviewed by Knezevic & Mizrahi [144], PET analysis offers the possibility of monitoring microglial activation in the brain of AD and MCI subjects, with a series of ligands.

Several inflammatory markers in plasma and CSF are elevated in AD and MCI subjects compared to age-matched controls [145–147] and in some cases the level of changes correlated with the severity of the disease, including sTREM2 in AD liquor [148]. An interesting approach was to monitor inflammatory parameters in CSF following the treatment in humans given CHF5074, an ibuprofen derivative with microglial modulation activity [149]; there was a constant reduction, possibly indicating drug efficacy. However, the correlation between the inflammatory factors in plasma and the disease progression,

determined by clinical markers, is only weak, when it occurs, because of the biological variability of the parameters considered. This variability more than an inevitable subjective oscillation, may describe real differences reflecting the heterogeneity of the disease, especially in its very early or prodromal phase [150]: under the broad umbrella of AD a variety of pathogenic pathways coexist.

The elements of the pathological scenario may have different weight, and inflammation must certainly be considered in relation to the time and the characteristics of the subjects. According to the precision medicine principle, the aim is to identify a common profile in a selected group of subjects, considering all the available parameters and the possibility of distinguishing different conditions. The application of precision medicine principles using the inflammatory profile to orient a personalized approach to AD was proposed by Wilcock's group [151,156]. They indicate M1 and M2 as broad inflammatory profiles, based on the measure of plasmatic proteins like ILRa, CAM1, haptoglobin, and fibrinogen, that can distinguish the AD population in the early phase from the immune point of view [153]. This polarization could be useful to identify subjects requiring a treatment to promote the process of restoration and repair associated with the M2 phenotype [153].

However, correct manipulation of the immune system will not be sufficient to interfere with AD progression, and a multifactorial approach will be necessary, focused on several targets including inflammation, amyloid aggregation and probably neuroprotection, to exert an effective anti-dementia action. The same molecules could share different activities. With intracerebral injection of ABOs and AβPP transgenic animals, we demonstrated that the antibiotic doxycycline had not only a well-defined anti-amyloidogenic profile [154, 155] but also antiinflammatory activity [156, 157]. Thus, doxycycline might be an interesting candidate for testing in the early phase of AD [158]. The drug has a clear safety profile and is currently under investigation in a preventive trial in subjects at genetic risk of fatal insomnia [159], a rare prion disease, included in the category of oligomeropathies [160].

CONCLUSIONS

The biological complexity of AD emerges from the scientific information accumulated in the last three decades, progress in imaging analysis, and general improvements in the diagnostic tools, which have extended our knowledge about the clinical progression of the disease, though less clearly in the preclinical phase. The aspects covered in this review point to the presence of ABOs and activation of the inflammatory cascade as early steps in the pathogenic scenario of AD, which involves several other important features, like tau phosphorylation, mitochondrial damage, and peripheral components. The control of inflammation and of amyloid aggregation is not a new target for AD therapies; large numbers of studies have been designed to test the effects of anti-amyloid or anti-inflammatory drugs but, until now, with scant results. To improve this approach, we have to move essentially in three directions: 1) improve the schedule and timing of treatment, possibly in a preclinical phase; 2) identify appropriate inflammatory markers that together with the genotype information can individualize treatment according to precision medicine principles; and 3) employ specific tools but also contemplate a multifactorial approach coherent with the distinct alterations seen in the disease.

Some of these aims would in principle be relatively simple to reach, while others call for careful investigation to identify appropriate tools. What is absolutely necessary is close collaboration between all the expertise at various levels—academic, pharmaceutical, and regulatory—to develop innovative trials where the quality of the patients selected is given priority, even at the expense of the numbers.

DISCLOSURE STATEMENT

Authors' disclosures available online (https://www.j-alz.com/manuscript-disclosures/17-0819r2).

REFERENCES

- [1] Heneka MT, Carson MJ, El Khoury J, Landreth GE, Brosseron F, Feinstein DL, Jacobs AH, Wyss-Coray T, Vitorica J, Ransohoff RM, Herrup K, Frautschy SA, Finsen B, Brown Selkoe GC, Verkhratsky A, Yamanaka K, Koistinaho J, Latz E, Halle A, Petzold GC, Town T, Morgan D, Shinohara ML, Perry VH, Holmes C, Bazan NG, Brooks DJ, Hunot S, Joseph B, Deigendesch N, Garaschuk O, Boddeke E, Dinarello CA, Breitner JC, Cole GM, Golenbock DT, Kummer MP (2015) Neuroinflammation in Alzheimer's disease. Lancet Neurol 14, 388-405.
- [2] Terry RD, Masliah E, Salmon DP, Butters N, DeTeresa R, Hill R, Hansen LA, Katzman R. (1991) Physical basis of cognitive alterations in Alzheimer's disease: Synapse loss is the major correlate of cognitive impairment. *Ann Neurol* 30, 572-578.

- [3] Walsh DM, Klyubin I, Fadeeva JV, Cullen WK, Anwyl R, Wolfe MS, Rowan MJ, Selkoe DJ (2001) Naturally secreted oligomers of amyloid beta protein potently inhibit hippocampal long-term potentiation in vivo. *Nature* 416, 535-539.
- [4] Balducci C, Beeg M, Stravalaci M, Bastone A, Sclip A, Biasini E, Tapelll, Colombo L Canzoni C, Borsello T, Chiesa R, Gobbi M, Salmona M, Forloni G (2010) Aβ oligomers impair memory independently of cellular prion protein. *Proc Natl Acad Sci U S A* 107, 2295-2300.
- [5] Näslund J, Haroutunian V, Mohs R, Davis KL, Davies P, Greengard P, Buxbaum JD (2000) Correlation between elevated levels of amyloid beta-peptide in the brain and cognitive decline. *JAMA* 283, 1571-1577.
- [6] Savva GM, Wharton SB, Ince PG, Forster G, Matthews FE, Brayne C (2009) Age, neuropathology, and dementia. N Engl J Med 360, 2302-2309.
- [7] Selkoe DJ, Hardy J (2016) The amyloid hypothesis of Alzheimer's disease at 25 years. EMBO Mol Med 8, 595-608.
- [8] Haass C, Selkoe DJ (2007) Soluble protein oligomers in neurodegeneration: Lessons from the Alzheimer's amyloid beta-peptide. Nat Rev Mol Cell Biol 8, 101-112.
- [9] Wilcox KC, Lacor PN, Pitt J, Klein WL (2011) Aβ oligomer-induced synapse degeneration in Alzheimer's disease. *Cell Mol Neurobiol* 31, 939-948.
- [10] Forloni G, Artuso V, La Vitola P, Balducci C (2016) Oligomeropathies and pathogenesis of Alzheimer and Parkinson's diseases. *Mov Disord* 31, 771-781.
- [11] Ono K (2017) Alzheimer's disease as oligomeropathy. Neurochem Int. doi: 10.1016/j.neuint.2017.08.010 [Epub ahead of print]
- [12] Jucker M, Walker LC (2013) Self-propagation of pathogenic protein aggregates in neurodegenerative diseases. *Nature* 501, 45-51.
- [13] Sengupta U, Nilson AN, Kayed R (2016) The role of amyloid-β oligomers in toxicity, propagation, and immunotherapy. *EBioMedicine* **6**, 42-49.
- [14] Lv ZY, Tan CC, Yu JT, Tan L (2017) Spreading of pathology in Alzheimer's disease. *Neurotox Res* 32, 707-722.
- [15] Dean DN, Das PK, Rana P, Burg F, Levites Y, Morgan SE, Ghosh P, Rangachari V (2017) Strain-specific fibril propagation by an Aβ dodecamer. Sci Rep 7, 40787.
- [16] Tasaki M, Ueda M, Ochiai S, Tanabe Y, Murata S, Misumi Y, Su Y, Sun X, Shinriki S, Jono H, Shono M, Obayashi K, Ando Y (2010) Transmission of circulating cell-free AA amyloid oligomers in exosomes vectors via a prionlike mechanism. *Biochem Biophys Res Commun* 400, 559-562.
- [17] Chung WS, Welsh CA, Barres BA, Stevens B (2015) Do glia drive synaptic and cognitive impairment in disease? *Nat Neurosci* 18, 1539-1545.
- [18] Jiang S, Bhaskar K (2017) Dynamics of the complement, cytokine, and chemokine systems in the regulation of synaptic function and dysfunction Relevant to Alzheimer's disease. J Alzheimers Dis 57, 1123-1135.
- [19] Wolf SA, Boddeke HW, Kettenmann H (2017) Microglia in physiology and disease. Annu Rev Physiol 79, 619-643.
- [20] Plaza-Zabala A, Sierra-Torre V, Sierra A (2017) Autophagy and microglia: Novel partners in neurodegeneration and aging. Int J Mol Sci 18, 598.
- [21] Liddelow SA, Guttenplan KA, Clarke LE, Bennett FC, Bohlen CJ, Schirmer L, Bennett ML, Münch AE, Chung WS, Peterson TC, Wilton DK, Frouin A, Napier BA, Panicker N, Kumar M, Buckwalter MS, Rowitch DH, Dawson

- VL, Dawson TM, Stevens B, Barres BA (2017) Neurotoxic reactive astrocytes are induced by activated microglia. *Nature* **541**, 481-487.
- [22] Kim SK, Nabekura J, Koizumi S (2017) Astrocyte-mediated synapse remodeling in the pathological brain. Glia 65, 1719-1727.
- [23] Guerreiro R, Wojtas A, Bras J, Carrasquillo M, Rogaeva E, Majounie E, Cruchaga C, Sassi C, Kauwe JS, Younkin S, Hazrati L, Collinge J, Pocock J, Lashley T, Williams J, Lambert JC, Amouyel P, Goate A, Rademakers R, Morgan K, Powell J, St George-Hyslop P, Singleton A, Hardy J, Alzheimer Genetic Analysis Group (2013) TREM2 variants in Alzheimer's disease. N Engl J Med 368, 117-127.
- [24] Jonsson T, Stefansson H, Steinberg S, Jonsdottir I, Jonsson PV, Snaedal J, Bjornsson S, Huttenlocher J, Levey AI, Lah JJ, Rujescu D, Hampel H, Giegling I, Andreassen OA, Engedal K, Ulstein I, Djurovic S, Ibrahim-Verbaas C, Hofman A, Ikram MA, van Duijn CM, Thorsteinsdottir U, Kong A, Stefansson K (2013) Variant of TREM2 associated with the risk of Alzheimer's disease. N Engl J Med 368, 107-116.
- [25] Rosenberg RN, Lambracht-Washington D, Yu G, Xia W (2016) Genomics of Alzheimer disease: A review. JAMA Neurol 73, 867-874.
- [26] Efthymiou AG, Goate AM (2017) Late onset Alzheimer's disease genetics implicates microglial pathways in disease risk. Mol Neurodegener 12, 43.
- [27] Yankner BA, Duffy LK, Kirschner DA (1990) Neurotrophic and neurotoxic effects of amyloid beta protein: Reversal by tachykinin neuropeptides. *Science* 250, 279-282.
- [28] Pike CJ, Burdick D, Walencewicz AJ, Glabe CG, Cotman CW (1993) Neurodegeneration induced by beta-amyloid peptides in vitro: The role of peptide assembly state. *J Neurosci* **13**, 1676-1687.
- [29] Forloni G (1993) Beta-Amyloid neurotoxicity. Funct Neurol 8, 211-225.
- [30] Forloni G, Lucca E, Angeretti N, Della Torre P, Salmona M (1997) Amidation of beta-amyloid peptide strongly reduced the amyloidogenic activity without alteration of the neurotoxicity. *J Neurochem* 69, 2048-2054.
- [31] Lambert MP, Barlow AK, Chromy BA, Edwards C, Freed R, Liosatos M, Morgan TE, Rozovsky I, Trommer B, Viola KL, Wals P, Zhang C, Finch CE, Krafft GA, Klein WL (1998) Diffusible, nonfibrillar ligands derived from Abeta1-42 are potent central nervous system neurotoxins. Proc Natl Acad Sci U S A 95, 6448-6453.
- [32] Benilova I, Karran E, De Strooper B (2012) The toxic Aβ oligomer and Alzheimer's disease: An emperor in need of clothes. *Nat Neurosci* 15, 349-357.
- [33] Bobo C, Chaignepain S, Henry S, Vignaud H, Améadan A, Marchal C, Prado E, Doutch J, Schmitter JM, Nardin C, Lecomte S, Cullin C (2017) Synthetic toxic Aβ1-42 oligomers can assemble in different morphologies. *Biochim Biophys Acta* 1861(5 Pt A), 1168-1176.
- [34] Jarosz-Griffiths HH, Noble E, Rushworth JV, Hooper NM (2016) amyloid-β receptors: The good, the bad, and the prion protein. *J Biol Chem* **291**, 3174-3183.
- [35] Evangelisti E, Cascella R, Becatti M, Marrazza G, Dobson CM, Chiti F, Stefani M, Cecchi C (2016) Binding affinity of amyloid oligomers to cellular membranes is a generic indicator of cellular dysfunction in protein misfolding diseases. Sci Rep 6, 32721.

- [36] Kayed R, Head E, Thompson JL, McIntire TM, Milton SC, Cotman CW, Glabe CG (2003) Common structure of soluble amyloid oligomers implies common mechanism of pathogenesis. *Science* 300, 486-489.
- [37] Roberts HL, Brown DR (2015) Seeking a mechanism for the toxicity of oligomeric α-synuclein. *Biomolecules* 5, 282-305.
- [38] Ugalde CL, Finkelstein DI, Lawson VA, Hill AF (2016) Pathogenic mechanisms of prion protein, amyloid-β and α-synuclein misfolding: The prion concept and neurotoxicity of protein oligomers. J Neurochem 139, 162-180.
- [39] Ladiwala AR, Litt J, Kane RS, Aucoin DS, Smith SO, Ranjan S, Davis J, Van Nostrand WE, Tessier PM (2012) Conformational differences between two amyloid β oligomers of similar size and dissimilar toxicity. *J Biol Chem* 287, 24765-24773.
- [40] Ojha J, Masilamoni G, Dunlap D, Udoff RA, Cashikar AG (2011) Sequestration of toxic oligomers by HspB1 as a cytoprotective mechanism. *Mol Cell Biol* 31, 3146-3157.
- [41] Bemporad F, Chiti F (2012) Protein misfolded oligomers: Experimental approaches, mechanism of formation, and structure-toxicity relationships. *Chem Biol* 19, 315-327.
- [42] Narayan P, Meehan S, Carver JA, Wilson MR, Dobson CM, Klenerman D (2012) Amyloid-β oligomers are sequestered by both intracellular and extracellular chaperones. *Biochemistry* 51, 9270-9276.
- [43] Mannini B, Mulvihill E, Sgromo C, Cascella R, Kho-darahmi R, Ramazzotti M, Dobson CM, Cecchi C, Chiti F (2014) Toxicity of protein oligomers is rationalized by a function combining size and surface hydrophobicity. ACS Chem Biol 9, 2309-2317.
- [44] Arosio P, Michaels TC, Linse S, Månsson C, Emanuelsson C, Presto J, Johansson J, Vendruscolo M, Dobson CM, Knowles TP (2016) Kinetic analysis reveals the diversity of microscopic mechanisms through which molecular chaperones suppress amyloid formation. *Nat Commun* 7, 10948.
- [45] Mannini B, Chiti F (2017) Chaperones as suppressors of protein misfolded oligomer toxicity. Front Mol Neurosci 10 98
- [46] Pivtoraiko VN, Abrahamson EE, Leurgans SE, DeKosky ST, Mufson EJ, Ikonomovic MD (2015) Cortical pyroglutamate amyloid-β levels and cognitive decline in Alzheimer's disease. Neurobiol Aging 36, 12-19.
- [47] Nussbaum JM, Schilling S, Cynis H, Silva A, Swanson E, Wangsanut T, Tayler K, Wiltgen B, Hatami A, Rönicke R, Reymann K, Hutter-Paier B, Alexandru A, Jagla W, Graubner S, Glabe CG, Demuth HU, Bloom GS (2012) Prion-like behaviour and tau-dependent cytotoxicity of pyroglutamylated amyloid-β. Nature 485, 651-455.
- [48] Sofola-Adesakin O, Khericha M, Snoeren I, Tsuda L, Partridge L (2016) pGluAβ increases accumulation of Aβ in vivo and exacerbates its toxicity. Acta Neuropathol Commun 4, 109.
- [49] Goldblatt G, Cilenti L, Matos JO, Lee B, Ciaffone N, Wang QX, Tetard L, Teter K, Tatulian SA (2017) Unmodified and pyroglutamylated amyloid β peptides form hypertoxic hetero-oligomers of unique secondary structure. FEBS J 284, 1355-1369.
- [50] Li G, Hu ZW, Chen PG, Sun ZY, Chen YX, Zhao YF, Li YM (2017) Prophylactic vaccine based on pyroglutamate-3 amyloid β generates strong antibody response and rescues cognitive decline in Alzheimer's disease model mice. ACS Chem Neurosci 8, 454-459.

- [51] Antonios G, Borgers H, Richard BC, Brauß A, Meißner J, Weggen S, Pena V, Pillot T, Davies SL, Bakrania P, Matthews D, Brownlees J, Bouter Y, Bayer TA (2015) Alzheimer therapy with an antibody against N-terminal Abeta 4-X and pyroglutamate Abeta 3-X. Sci Rep 5, 17338
- [52] Jack CR Jr, Barrio JR, Kepe V (2013) Cerebral amyloid PET imaging in Alzheimer's disease. *Acta Neuropathol* 126, 643-657.
- [53] Baker JE, Lim YY, Pietrzak RH, Hassenstab J, Snyder PJ, Masters CL, Maruff P (2016) Cognitive impairment and decline in cognitively normal older adults with high amyloid-β: A meta-analysis. *Alzheimers Dement* **6**, 108-121.
- [54] Nordberg A, Rinne JO, Kadir A, Långström B (2010) The use of PET in Alzheimer disease. *Nat Rev Neurol* 6, 78-87.
- [55] Kadir A, Marutle A, Gonzalez D, Schöll M, Almkvist O, Mousavi M, Mustafiz T, Darreh-Shori T, Nennesmo I, Nordberg A (2011) Positron emission tomography imaging and clinical progression in relation to molecular pathology in the first Pittsburgh Compound B positron emission tomography patient with Alzheimer's disease. *Brain* 134, 301-317.
- [56] Coutinho AM, Porto FH, Duran FL, Prando S, Ono CR, Feitosa EA, Spindola L, de Oliveira MO, do Vale PH, Gomes HR, Nitrini R, Brucki SM, Buchpiguel CA (2015) Brain metabolism and cerebrospinal fluid biomarkers profile of non-amnestic mild cognitive impairment in comparison to amnestic mild cognitive impairment and normal older subjects. Alzheimers Res Ther 7, 58.
- [57] Thapa A, Jett SD, Chi EY (2016) Curcumin attenuates amyloid-β aggregate toxicity and modulates amyloid-β aggregation pathway. ACS Chem Neurosci 7, 56-68.
- [58] Henry S, Vignaud H, Bobo C, Decossas M, Lambert O, Harte E, Alves ID, Cullin C, Lecomte S (2015) Interaction of Aβ(1-42) amyloids with lipids promotes "off-pathway" oligomerization and membrane damage. Biomacromolecules 16, 944-950.
- [59] Kodali R, Wetzel R (2007) Polymorphism in the intermediates and products of amyloid assembly. *Curr Opin Struct Biol* 17, 48-55.
- [60] Balducci C, Tonini R, Zianni E, Nazzaro C, Fiordaliso F, Salio M, Vismara L, Gardoni F, Di Luca M, Carli M, Forloni G (2010) Cognitive deficits associated with alteration of synaptic metaplasticity precede plaque deposition in AβPP23 transgenic mice. *J Alzheimers Dis* 21, 1367-1381.
- [61] Balducci C, Mehdawy B, Mare L, Giuliani A, Lorenzini L, Sivilia S, Giardino L, Calzá L, Lanzillotta A, Sarnico I, Pizzi M, Usiello A, Viscomi AR, Ottonello S, Villetti G, Imbimbo BP, Nisticò G, Forloni G, Nisticò R (2011) The γ-secretase modulator CHF5074 restores memory and hippocampal synaptic plasticity in plaque-free Tg2576 mice. J Alzheimers Dis 24, 799-816.
- [62] Gandy S, Simon AJ, Steele JW, Lublin AL, Lah JJ, Walker LC, Levey AI, Krafft GA, Levy E, Checler F, Glabe C, Bilker WB, Abel T, Schmeidler J, Ehrlich ME (2010) Days to criterion as an indicator of toxicity associated with human Alzheimer amyloid-beta oligomers. *Ann Neurol* 68, 220-230.
- [63] Hamm V, Héraud C, Bott JB, Herbeaux K, Strittmatter C, Mathis C, Goutagny R (2017) Differential contribution of APP metabolites to early cognitive deficits in a TgCRND8 mouse model of Alzheimer's disease. Sci Adv 3, e1601068.

- [64] Winton MJ, Lee EB, Sun E, Wong MM, Leight S, Zhang B, Trojanowski JQ, Lee VM (2011) Intraneuronal APP, not free Aβ peptides in 3xTg-AD mice: Implications for tau versus Aβ-mediated Alzheimer neurodegeneration. J Neurosci 31, 7691-7699.
- [65] Knight EM, Kim SH, Kottwitz JC, Hatami A, Albay R, Suzuki A, Lublin A, Alberini CM, Klein WL, Szabo P, Relkin NR, Ehrlich M, Glabe CG, Gandy S, Steele JW (2016) Effective anti-Alzheimer Aβ therapy involves depletion of specific Aβ oligomer subtypes. Neurol Neuroimmunol Neuroinflamm 3, e237.
- [66] Yang T, Li S, Xu H, Walsh DM, Selkoe DJ (2017) Large soluble oligomers of amyloid β-protein from Alzheimer brain are far less neuroactive than the smaller oligomers to which they dissociate. *J Neurosci* 37, 152-163.
- [67] Balducci C, Forloni G (2014) In vivo application of beta amyloid oligomers: A simple tool to evaluate mechanisms of action and new therapeutic approaches. *Curr Pharm Des* 20, 2491-2505.
- [68] Beeg M, Stravalaci M, Bastone A, Salmona M, Gobbi M (2011) A modified protocol to prepare seed-free starting solutions of amyloid-β (Aβ)1-40 and Aβ1-42 from the corresponding depsipeptides. *Anal Biochem* 411, 297-299.
- [69] Forloni G, Sclip A, Borsello T, Balducci C (2013) The neurodegeneration in Alzheimer disease and the prion protein. *Prion* 7, 60-65.
- [70] Salazar SV, Strittmatter SM (2017) Cellular prion protein as a receptor for amyloid-β oligomers in Alzheimer's disease. Biochem Biophys Res Commun 483, 1143-1147.
- [71] De Simoni MG, Del Bo R, De Luigi A, Simard S, Forloni G (1995) Central endotoxin induces different patterns of interleukin (IL)-1 beta and IL-6 messenger ribonucleic acid expression and IL-6 secretion in the brain and periphery. *Endocrinology* 136, 897-902.
- [72] Schwartz M, Deczkowska A (2016) Neurological disease as a failure of brain-immune crosstalk: The multiple faces of neuroinflammation. *Trends Immunol* 37, 668-679.
- [73] Serhan CN, Brain SD, Buckley CD, Gilroy DW, Haslett C, O'Neill LA, Perretti M, Rossi AG, Wallace JL (2007) Resolution of inflammation: State of the art, definitions and terms, FASEB J 21, 325-332.
- [74] Serhan CN (2014) Pro-resolving lipid mediators are leads for resolution physiology. *Nature* 510, 92-101.
- [75] Wang X, Zhu M, Hjorth E, Cortés-Toro V, Eyjolfsdottir H, Graff C, Nennesmo I, Palmblad J, Eriksdotter M, Sambamurti K, Fitzgerald JM, Serhan CN, Granholm AC, Schultzberg M (2015) Resolution of inflammation is altered in Alzheimer's disease. *Alzheimers Dement* 11, 40-50.
- [76] Zhu M, Wang X, Schultzberg M, Hjorth E (2015) Differential regulation of resolution in inflammation induced by amyloid-β42 and lipopolysaccharides in human microglia. J Alzheimers Dis 43, 1237-1250.
- [77] Heneka TM, Golenbock DT, Latz E (2015) Innate immune in Alzheimer's disease. *Nat Immunol* 16, 229-236.
- [78] Parbo P, Ismail R, Hansen KV, Amidi A, Mårup FH, Gottrup H, Brændgaard H, Eriksson BO, Eskildsen SF, Lund TE, Tietze A, Edison P, Pavese N, Stokholm MG, Borghammer P, Hinz R, Aanerud J, Brooks DJ (2017) Brain inflammation accompanies amyloid in the majority of mild cognitive impairment cases due to Alzheimer's disease. Brain 40, 2002-2011.

- [79] Carter SF, Schöll M, Almkvist O, Wall A, Engler H, Långström B, Nordberg A (2012) Evidence for astrocytosis in prodromal Alzheimer disease provided by 11C-deuterium-L-deprenyl: A multitracer PET paradigm combining 11C-Pittsburgh compound B and 18F-FDG. J Nucl Med 53, 37-46.
- [80] Guerriero R, Hardy J (2014) Genetics of Alzheimer's disease. Neurotherapeutics 11, 732-737.
- [81] Morris GP, Clark IA, Zinn R, Vissel B (2013) Microglia: A new frontier for synaptic plasticity, learning and memory, and neurodegenerative disease research, *Neurobiol Learn Mem* 105, 40-53.
- [82] Zorec R, Horvat A, Vardjan N, Verkhratsky A (2015) Memory formation shaped by astroglia. Front Integr Neurosci 9, 56-60
- [83] Maezawa I, Zimin PI, Wulff, H, Jin L-W (2011). Amyloidβ protein oligomer at low nanomolar concentrations activates microglia and induces microglial neurotoxicity. *J Biol Chem* 286, 3693-3706.
- [84] Li J, Cesari M, Liu F, Dong B, Vellas B (2017) Effects of diabetes mellitus on cognitive decline in patients with Alzheimer disease: A systematic review. Can J Diabetes 41, 114-119.
- [85] Rojas-Gutierrez E, Muñoz-Arenas G, Treviño S, Espinosa B, Chavez R, Rojas K, Flores G, Díaz A, Guevara J (2017) Alzheimer's disease and metabolic syndrome: A link from oxidative stress and inflammation to neurodegeneration. Synapse. doi: 10.1002/syn.21990 [Epub ahead of print]
- [86] Panza F, Frisardi V, Seripa D, Imbimbo BP, Sancarlo D, D'Onofrio G, Addante F, Paris F, Pilotto A, Solfrizzi V (2011) Metabolic syndrome, mild cognitive impairment, and dementia. *Curr Alzheimer Res* 8, 492-509.
- [87] Holmes, Cunningham C, Zotova E, Woolford J, Dean C, Kerr S, Culliford D, Perry VH (2009) Systemic inflammation and disease progression in Alzheimer disease. *Neurology* 73, 768-774.
- [88] Kamer AR, Craig RG, Pirraglia E, Dasanayake AP, Norman RG, Boylan RJ, Nehorayoff A, Glodzik L, Brys M, de Leon MJ (2009) TNF-α and antibodies to periodontal bacteria discriminate between Alzheimer's disease patients and normal subjects. *J Neuroimmunol* **216**, 92-97.
- [89] Kamer AR, Dasanayake AP, Craig RG, Glodzik-Sobanska L, Bry M, de Leon MJ (2008) Inflammation and Alzheimer's disease: Possible role of periodontal diseases. Alzheimers Dement 4, 242-250.
- [90] Cattaneo A, Cattaneo N, Galluzzi S, Provasi S, Lopizzo N, Festari C, Ferrari C, Guerra UP, Paghera B, Muscio C, Bianchetti A, Volta GD, Turla M, Cotelli MS, Gennuso M, Prelle A, Zanetti O, Lussignoli G, Mirabile D, Bellandi D, Gentile S, Belotti G, Villani D, Harach T, Bolmont T, Padovani A, Boccardi M, Frisoni GB, INDIA-FBP Group (2017) Association of brain amyloidosis with pro-inflammatory gut bacterial taxa and peripheral inflammation markers in cognitively impaired elderly. Neurobiol Aging 49, 60-68.
- [91] Walter J (2016) The triggering receptor expressed on myeloid cells 2: A molecular link of neuroinflammation and neurodegenerative diseases. *J Biol Chem* 291, 4334-4341.
- [92] Takahashi K, Rochford CD, Neumann H (2005) Clearance of apoptotic neurons without inflammation by microglial triggering receptor expressed on myeloid cells-2. *J Exp Med* 20, 647-657.
- [93] Hsieh CL, Koike M, Spusta SC, Niemi EC, Yenari M, Nakamura MC, Seaman WE (2009) A role for TREM2

- ligands in the phagocytosis of apoptotic neuronal cells by microglia. *J Neurochem* **109**, 1144-1156.
- [94] Lue LF, Schmitz C, Walker DG (2015) What happens to microglial TREM2 in Alzheimer's disease: Immunoregulatory turned into immunopathogenic? *Neuroscience* 302, 138-150.
- [95] Turnbull IR, Gilfillan S, Cella M, Aoshi T, Miller M, Piccio L, Hernandez M, Colonna M (2006) Cutting edge: TREM-2 attenuates macrophage activation. *J Immunol* 177, 3520-3524.
- [96] Tang Y, Le W (2006) Differential roles of M1 and M2 microglia in neurodegenerative diseases. *Mol Neurobiol* 53, 1181-1194.
- [97] Mazaheri F, Snaidero N, Kleinberger G, Madore C, Daria A, Werner G, Krasemann S, Capell A, Trümbach D, Wurst W, Brunner B, Bultmann S, Tahirovic S, Kerschensteiner M, Misgeld T, Butovsky O, Haass C (2017) TREM2 deficiency impairs chemotaxis and microglial responses to neuronal injury. EMBO Rep 18, 1186-1198.
- [98] Frank S, Burbach GJ, Bonin M, Walter M, Streit W, Bechmann I, Deller T (2008) TREM2 is upregulated in amyloid plaque-associated microglia in aged APP23 transgenic mice. Glia 56, 1438-1447.
- [99] Jay TR, Miller CM, Cheng PJ, Graham LC, Bemiller S, Broihier ML, Xu G, Margevicius D, Karlo JC, Sousa GL, Cotleur AC, Butovsky O, Bekris L, Staugaitis SM, Leverenz JB, Pimplikar SW, Landreth GE, Howell GR, Ransohoff RM, Lamb BT (2008) TREM2 deficiency eliminates TREM2+ inflammatory macrophages and ameliorates pathology in Alzheimer's disease mouse models, *J Exp Med* 212, 287-295.
- [100] Wang Y, Cella M, Mallinson K, Ulrich JD, Young KL, Robinette ML, Gilfillan S, Krishnan GM, Sudhakar S, Zinselmeyer BH, Holtzman DM, Cirrito JR, Colonna M (2015) TREM2 lipid sensing sustains the microglial response in an Alzheimer's disease model, Cell 160, 1061-1071.
- [101] Ulrich JD, Finn MB, Wang Y, Shen A, Mahan TE, Jiang H, Stewart FR, Piccio L, Colonna M, Holtzman DM (2014) Altered microglial response to Aβ plaques in APPPS1-21 mice heterozygous for TREM2, Mol Neurodegener 9, 20.
- [102] Jin SC, Benitez BA, Karch CM, Cooper B, Skorupa T, Carrell D, Norton JB, Hsu S, Harari O, Cai Y, Bertelsen S, Goate AM, Cruchaga C (2014) Coding variants in TREM2 increase risk for Alzheimer's disease. *Hum Mol Genet* 23, 5838-5846.
- [103] Ridge PG, Hoyt KB, Boehme K, Mukherjee S, Crane PK, Haines JL, Mayeux R, Farrer LA, Pericak-Vance MA, Schellenberg GD, Kauwe JSK, Alzheimer's Disease Genetics Consortium (ADGC) (2016) Assessment of the genetic variance of late-onset Alzheimer's disease. Neurobiol Aging 41, 200.e13-200.e20.
- [104] Jay TR, von Saucken VE, Landreth GE (2017) TREM2 in neurodegenerative diseases. Mol Neurodegener 12, 56.
- [105] Song W, Hooli B, Mullin K, Jin SC, Cella M, Ulland TK, Wang Y, Tanzi RE, Colonna M (2017) Alzheimer's disease-associated TREM2 variants exhibit either decreased or increased ligand-dependent activation, Alzheimers Dement 13, 381-387.
- [106] Sirkis DW, Bonham LW, Aparicio RE, Geier EG, Ramos EM, Wang Q, Karydas A, Miller ZA, Miller BL, Coppola G, Yokoyama JS (2016) Rare TREM2 variants associated with Alzheimer's disease display reduced cell surface expression. Acta Neuropathol Commun 4, 98.

- [107] Lue LF, Schmitz CT, Serrano G, Sue LI, Beach TG, Walker DG (2015) TREM2 protein expression changes correlate with Alzheimer's disease neurodegenerative pathologies in post-mortem temporal cortices. *Brain Pathol* 25, 469-480
- [108] Kober DL, Alexander-Brett JM, Karch CM, Cruchaga C, Colonna M, Holtzman MJ, Brett TJ (2016) Neurodegenerative disease mutations in TREM2 reveal a functional surface and distinct loss-of-function mechanisms, *ELife* 5, pii: e20391. doi: 10.7554/eLife.20391
- [109] Ulland TK, Song WM, Huang SC, Ulrich JD, Sergushichev A, Beatty WL, Loboda AA, Zhou Y, Cairns NJ, Kambal A, Loginicheva E, Gilfillan S, Cella M, Virgin HW, Unanue ER, Wang Y, Artyomov MN, Holtzman DM, Colonna M (2017) TREM2 maintains microglial metabolic fitness in Alzheimer's disease. Cell 170, 649-663
- [110] Keren-Shaul H, Spinrad A, Weiner A, Matcovitch-Natan O, Dvir-Szternfeld R, Ulland TK, David E, Baruch K, Lara-Astaiso D, Toth B, Itzkovitz S, Colonna M, Schwartz M, Amit I (2017) A unique microglia type associated with restricting development of Alzheimer's disease. *Cell* 169, 1276-1290.
- [111] Jun G, Naj AC, Beecham GW, Wang LS, Buros J, Gallins PJ, Buxbaum JD, Ertekin-Taner N, Fallin MD, Friedland R, Inzelberg R, Kramer P, Rogaeva E, St George-Hyslop P; Alzheimer's Disease Genetics Consortium, Cantwell LB, Dombroski BA, Saykin AJ, Reiman EM, Bennett DA, Morris JC, Lunetta KL, Martin ER, Montine TJ, Goate AM, Blacker D, Tsuang DW, Beekly D, Cupples LA, Hakonarson H, Kukull W, Foroud TM, Haines J, Mayeux R, Farrer LA, Pericak-Vance MA, Schellenberg GD (2010) Meta-analysis confirms CR1, CLU, and PICALM as Alzheimer disease risk loci and reveals interactions with APOE genotypes. Arch Neurol 67, 1473-1484.
- [112] Stevens B, Allen NJ, Vazquez LE, Howell GR, Christopherson KS, Nouri N, Micheva KD, Mehalow AK, Huberman AD, Stafford B, Sher A, Litke AM, Lambris JD, Smith SJ, John SW, Barres BA (2007) The classical complement cascade mediates CNS synapse elimination Cell 131, 1164-1178.
- [113] Bialas AR, Stevens B (2013) TGF-β signaling regulates neuronal C1q expression and developmental synaptic refinement, *Nat Neurosci* 16, 1773-1782.
- [114] Schafer DP, Lehrman EK, Kautzman AG, Koyama R, Mardinly AR, Yamasaki R, Ransohoff RM, Greenberg ME, Barres BA, Stevens B (2012) Microglia sculpt postnatal neural circuits in an activity and complementdependent manner. *Neuron* 74, 691-705.
- [115] Stephan AH, Madison DV, Mateos JM, Fraser DA, Lovelett EA, Coutellier L, Kim L, Tsai HH, Huang EJ, Rowitch DH, Berns DS, Tenner AJ, Shamloo M, Barres BA (2013) A dramatic increase of C1q protein in the CNS during normal aging. *J Neurosci* 33, 13460-13474.
- [116] Tenner AJ, Fonseca MI (2006) The double-edged flower: Roles of complement protein C1q in neurodegenerative diseases. Adv Exp Med Biol 586, 153-176.
- [117] Kim J, Yoon H, Basak J, Kim J (2014) Apolipoprotein E in synaptic plasticity and Alzheimer's disease: Potential cellular and molecular mechanisms. *Mol Cells* 37, 767-776
- [118] Lui H, Zhang J, Makinson SR, Cahill MK, Kelley KW, Huang HY, Shang Y, Oldham MC, Martens LH, Gao F, Coppola G, Sloan SA, Hsieh CL, Kim CC, Bigio EH, Weintraub S, Mesulam MM, Rademakers R, Mackenzie

- IR, Seeley WW, Karydas A, Miller BL, Borroni B, Ghidoni R, Farese RV Jr, Paz JT, Barres BA, Huang EJ (2016) Progranulin deficiency promotes circuit-specific synaptic pruning by microglia via complement activation. *Cell* **165**, 921-935.
- [119] Chung WS, Verghese PB, Chakraborty C, Joung J, Hyman BT, Ulrich JD, Holtzman DM, Barres BA (2016) Novel allele-dependent role for APOE in controlling the rate of synapse pruning by astrocytes. *Proc Natl Acad Sci U S A* 113, 10186-10191.
- [120] Hong S, Beja-Glasser VF, Nfonoyim BM, Frouin A, Li S, Ramakrishnan S, Merry KM, Shi Q, Rosenthal A, Barres BA, Lemere CA, Selkoe DJ, Stevens B (2016) Complement and microglia mediate early synapse loss in Alzheimer mouse models. *Science* 352, 712-716.
- [121] Schafer DP, Lehrman EK, Kautzman AG, Koyama R, Mardinly AR, Yamasaki R, Ransohoff RM, Greenberg ME, Barres BA, Stevens B (2012) Microglia sculpt postnatal neural circuits in an activity and complementdependent manner. *Neuron* 74, 691-705.
- [122] Balducci C, Frasca A, Zotti M, La Vitola P, Mhillaj E, Grigoli E, Iacobellis M, Grandi F, Messa M, Colombo L, Molteni M, Trabace L, Rossetti C, Salmona M, Forloni G (2017) Toll-like receptor 4-dependent glial cell activation mediates the impairment in memory establishment induced by β-amyloid oligomers in an acute mouse model of Alzheimer's disease. Brain Behav Immunol 60, 188-197.
- [123] Ledo JH, Azevedo EP, Beckman D, Ribeiro FC, Santos LE, Razolli DS, Kincheski GC, Melo HM, Bellio M, Teixeira AL, Velloso LA, Foguel D, De Felice FG, Ferreira ST (2016) Cross talk between brain innate immunity and serotonin signaling underlies depressive-like behavior induced by Alzheimer's amyloid-β oligomers in mice. *J Neurosci* 36, 12106-12116.
- [124] Woodling NS, Colas D, Wang Q, Minhas P, Panchal M, Liang X, Mhatre SD, Brown H, Ko N, Zagol-Ikapitte I, van der Hart M, Khroyan TV, Chuluun B, Priyam PG, Milne GL, Rassoulpour A, Boutaud O, Manning-Boğ AB, Heller HC, Andreasson KI (2016) Cyclooxygenase inhibition targets neurons to prevent early behavioural decline in Alzheimer's disease model mice. *Brain* 139, 2063-2081.
- [125] Xu H, Gelyana E, Rajsombath M, Yang T, Li S, Selkoe D (2016) Environmental enrichment potently prevents microglia-mediated neuroinflammation by human amyloid β-protein oligomers. J Neurosci 36, 9041-9056.
- [126] Beauquis J, Pav.a P, Pomilio C, Vinuesa A, Podlutskaya N, Galvan V, Saravia F (2013). Environmental enrichment prevents astroglial pathological changes in the hippocampus of APP transgenic mice, model of Alzheimer's disease. *Exp Neurol* 239, 28-37.
- [127] Schenk D, Barbour R, Dunn W, Gordon G, Grajeda H, Guido T, Hu K, Huang J, Johnson-Wood K, Khan K, Kholodenko D, Lee M, Liao Z, Lieberburg I, Motter R, Mutter L, Soriano F, Shopp G, Vasquez N, Vandevert C, Walker S, Wogulis M, Yednock T, Games D, Seubert P (1999) Immunization with amyloid-beta attenuates Alzheimer-disease-like pathology in the PDAPP mouse. Nature 400, 173-177.
- [128] Orgogozo JM, Gilman S, Dartigues JF, Laurent B, Puel M, Kirby LC, Jouanny P, Dubois B, Eisner L, Flitman S, Michel BF, Boada M, Frank A, Hock C (2003) Subacute meningoencephalitis in a subset of patients with AD after Abeta42 immunization. *Neurology* 61, 46-54.

- [129] Liu J, Yang B, Ke J, Li W, Suen WC (2016) Antibodybased drugs and approaches against amyloid-β species for Alzheimer's disease immunotherapy. *Drugs Aging* 33, 685-697.
- [130] Wang Y, Yan T, Lu H, Yin W, Lin B, Fan W, Zhang X, Fernandez-Funez P (2017) Lessons from anti-amyloidβ immunotherapies in Alzheimer disease: Aiming at a moving target. Neurodegener Dis 17, 242-250.
- [131] Nicoll JA, Wilkinson D, Holmes C, Steart P, Markham H, Weller RO (2003) Neuropathology of human Alzheimer disease after immunization with amyloid-beta peptide: A case report. Nat Med 9, 448-452.
- [132] Patton RL, Kalback WM, Esh CL, Kokjohn TA, Van Vickle GD, Luehrs DC, Kuo YM, Lopez J, Brune D, Ferrer I, Masliah E, Newel AJ, Beach TG, Castaño EM, Roher AE (2006) Amyloid-beta peptide remnants in AN-1792immunized Alzheimer's disease patients: A biochemical analysis. Am J Pathol 169, 1048-1063.
- [133] Sevigny J, Chiao P, Bussiere T, Weinreb PH, Williams L, Maier M, Dunstan R, Salloway S, Chen T, Ling Y, O'Gorman J, Qian F, Arastu M, Li M, Chollate S, Brennan MS, Quintero-Monzon O, Scannevin RH, Arnold HM, Engber T, Rhodes K, Ferrero J, Hang Y, Mikulskis A, Grimm J, Hock C, Nitsch RM, Sandrock A (2016) The antibody aducanumab reduces Abeta plaques in Alzheimer's disease. *Nature* 537, 50-56.
- [134] in t' Veld BA, Ruitenberg A, Hofman A, Launer LJ, van Duijn CM, Stijnen T, Breteler MM, Stricker BH (2001) Nonsteroidal antiinflammatory drugs and the risk of Alzheimer's disease. N Engl J Med 345, 1515-1521.
- [135] Wang J, Tan L, Wang HF, Tan CC, Meng XF, Wang C, Tang SW, Yu JT (2015) Anti-inflammatory drugs and risk of Alzheimer's disease: An updated systematic review and meta-analysis. J Alzheimers Dis 44, 385-396.
- [136] Miguel-Álvarez M, Santos-Lozano A, Sanchis-Gomar F, Fiuza-Luces C, Pareja-Galeano H, Garatachea N, Lucia A (2015) Non-steroidal anti-inflammatory drugs as a treatment for Alzheimer's disease: A systematic review and meta-analysis of treatment effect. *Drugs Aging* 32, 139-147
- [137] Rogers J, Kirby LC, Hempelman SR, Berry DL, McGeer PL, Kaszniak AW, Zalinski J, Cofield M, Mansukhani L, Willson P, Kogan F (1993) Clinical trial of indomethacin in Alzheimer's disease. *Neurology* 43, 1609-1611.
- [138] de Jong D, Jansen R, Hoefnagels W, Jellesma-Eggenkamp M, Verbeek M, Borm G, Kremer B (2008) No effect of oneyear treatment with indomethacin on Alzheimer's disease progression: A randomized controlled trial. *PLoS One* 3, e1475.
- [139] Anand R, Gill KD, Mahdi AA (2014) Therapeutics of Alzheimer's disease: Past, present and future. *Neurophar-macology* 76(Pt A), 27-50.
- [140] Banik A, Brown RE, Bamburg J, Lahiri DK, Khurana D, Friedland RP, Chen W, Ding Y, Mudher A, Padjen AL, Mukaetova-Ladinska E, Ihara M, Srivastava S, Padma Srivastava MV, Masters CL, Kalaria RN, Anand A (2016) Translation of pre-clinical studies into successful clinical trials for Alzheimer's disease: What are the roadblocks and how can they be overcome? *J Alzheimers Dis* 47, 815-843.
- [141] Calzá L, Baldassarro VA, Giuliani A, Lorenzini L, Fernandez M, Mangano C, Sivilia S, Alessandri M, Gusciglio M, Torricella R, Giardino L (2013) From the multifactorial nature of Alzheimer's disease to multitarget therapy: The

- contribution of the translational approach. *Curr Top Med Chem* **13**, 1843-1852.
- [142] Albani D, Tettamanti M, Batelli S, Polito L, Dusi S, Ateri E, Forloni G, Lucca U (2012) Interleukin-1α, interleukin-1β and tumor necrosis factor-α genetic variants and risk of dementia in the very old: Evidence from the "Monzino 80-plus" prospective study. Age (Dordr) 34, 519-526.
- [143] Su F, Bai F, Zhang Z (2016) Inflammatory cytokines and Alzheimer's disease: A review from the perspective of genetic polymorphisms. *Neurosci Bull* 32, 469-480.
- [144] Knezevic D, Mizrahi R (2018) Molecular imaging of neuroinflammation in Alzheimer's disease and mild cognitive impairment. *Prog Neuropsychopharmacol Biol Psychiatry* 80(Pt B), 123-131.
- [145] Leung R, Proitsi P, Simmons A, Lunnon K, Güntert A, Kronenberg D, Pritchard M, Tsolaki M, Mecocci P, Kloszewska I, Vellas B, Soininen H, Wahlund LO, Lovestone S (2013) Inflammatory proteins in plasma are associated with severity of Alzheimer's disease. *PLoS One* 8, e64971.
- [146] Demirci S, Aynali A, Demirci K, Demirci S, Aridoğan BC (2017) The serum levels of resistin and its relationship with other proinflammatory cytokines in patients with Alzheimer's disease. Clin Psychopharmacol Neurosci 15, 59-63.
- [147] Lai KSP, Liu CS, Rau A, Lanctôt KL, Köhler CA, Pakosh M, Carvalho AF, Herrmann N (2017) Peripheral inflammatory markers in Alzheimer's disease: A systematic review and meta-analysis of 175 studies. J Neurol Neurosurg Psychiatry 88, 876-882.
- [148] Suérez-Calvet M, Kleinberger G, Araque Caballero MÁ, Brendel M, Rominger A, Alcolea D, Fortea J, Lleó A, Blesa R, Gispert JD, Sénchez-Valle R, Antonell A, Rami L, Molinuevo JL, Brosseron F, Traschütz A, Heneka MT, Struyfs H, Engelborghs S, Sleegers K, Van Broeckhoven C, Zetterberg H, Nellgård B, Blennow K, Crispin A, Ewers M, Haass C (2016) sTREM2 cerebrospinal fluid levels are a potential biomarker for microglia activity in early-stage Alzheimer's disease and associate with neuronal injury markers. EMBO Mol Med 8, 466-476.
- [149] Ross J, Sharma S, Winston J, Nunez M, Bottini G, Franceschi M, Scarpini E, Frigerio E, Fiorentini F, Fernandez M, Sivilia S, Giardino L, Calza L, Norris D, Cicirello H, Casula D, Imbimbo BP (2013) CHF5074 reduces biomarkers of neuroinflammation in patients with mild cognitive impairment: A 12-week, double-blind, placebocontrolled study. Curr Alzheimer Res 10, 742-753.
- [150] Martin E, Boucher C, Fontaine B, Delarasse C (2017) Distinct inflammatory phenotypes of microglia and monocyte-derived macrophages in Alzheimer's disease models: Effects of aging and amyloid pathology. *Aging Cell* 16, 27-38.
- [151] Sudduth TL, Schmitt FA, Nelson PT, Wilcock DM (2013) Neuroinflammatory phenotype in early Alzheimer's disease. *Neurobiol Aging* 34, 1051-1059.
- [152] Wilcock DM (2014) Neuroinflammatory phenotypes and their roles in Alzheimer's disease. *Neurodegener Dis* 13, 183-185.
- [153] Latta CH, Brothers HM, Wilcock DM (2015) Neuroinflammation in Alzheimer's disease; A source of heterogeneity and target for personalized therapy. *Neu*roscience 302, 103-111.
- [154] Forloni G, Colombo L, Girola L, Tagliavini F, Salmona M (2001) Anti-amyloidogenic activity of tetracyclines: Studies in vitro. FEBS Lett 487, 404-409.

- [155] Stoilova T, Colombo L, Forloni G, Tagliavini F, Salmona M (2013) A new face for old antibiotics: Tetracyclines in treatment of amyloidoses. *J Med Chem* 56, 5987-6006.
- [156] Sultan S, Gebara E, Toni N (2013) Doxycycline increases neurogenesis and reduces microglia in the adult hippocampus. Front Neurosci 7, 131.
- [157] Mello BS, Monte AS, McIntyre RS, Soczynska JK, Custódio CS, Cordeiro RC, Chaves JH, Vasconcelos SM, Nobre HV Jr, Florenço de Sousa FC, Hyphantis TN, Carvalho AF, Macêdo DS (2013) Effects of doxycycline on depressive-like behavior in mice after lipopolysaccharide (LPS) administration. J Psychiatr Res 47, 1521-1529.
- [158] Mehla J (2017) Repurposing clinically used antibiotics for the treatment of Alzheimer's disease J Pharmacol Rep 2, e102.
- [159] Forloni G, Tettamanti M, Lucca U, Albanese Y, Quaglio E, Chiesa R, Villani F, Erbetta E, Redaelli V, Tagliavini F, Artuso V, Roiter I (2015) Preventive study in subjects at risk of FFI: Innovative approach to rare diseases. *Prion* 9, 75-79.
- [160] Forloni G, Artuso V, Roiter I, Morbin M, Tagliavini F (2013) Therapy in prion diseases. *Curr Top Med Chem* 13, 2465-2476.