

# Sleep Dysregulation Is Associated with $^{18}\text{F}$ -FDG PET and Cerebrospinal Fluid Biomarkers in Alzheimer's Disease

Mariana Fernandes<sup>a</sup>, Agostino Chiaravalloti<sup>b,c</sup>, Marzia Nuccetelli<sup>d</sup>, Fabio Placidi<sup>a,e</sup>, Francesca Izzi<sup>e</sup>, Riccardo Camedda<sup>b</sup>, Sergio Bernardini<sup>d</sup>, Giuseppe Sancesario<sup>e</sup>, Orazio Schillaci<sup>b</sup>, Nicola Biagio Mercuri<sup>a,e</sup> and Claudio Liguori<sup>a,e,\*</sup>

<sup>a</sup>Department of Systems Medicine, University of Rome "Tor Vergata", Rome, Italy

<sup>b</sup>Department of Biomedicine and Prevention, University of Rome "Tor Vergata", Rome, Italy

<sup>c</sup>IRCCS Neuromed, Pozzilli, Italy

<sup>d</sup>Department of Clinical Biochemistry and Molecular Biology, University of Rome "Tor Vergata", Rome, Italy

<sup>e</sup>Sleep Medicine Centre, Neurology Unit, University Hospital of Rome "Tor Vergata", Rome, Italy

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## Abstract.

**Background:** Sleep impairment has been commonly reported in Alzheimer's disease (AD) patients. The association between sleep dysregulation and AD biomarkers has been separately explored in mild cognitive impairment (MCI) and AD patients.

**Objective:** The present study investigated cerebrospinal-fluid (CSF) and  $^{18}\text{F}$ -fluoro-deoxy-glucose positron emission tomography ( $^{18}\text{F}$ -FDG-PET) biomarkers in MCI and AD patients in order to explore their association with sleep parameters measured with polysomnography (PSG).

**Methods:** MCI and AD patients underwent PSG,  $^{18}\text{F}$ -FDG-PET, and CSF analysis for detecting and correlating these biomarkers with sleep architecture.

**Results:** Thirty-five patients were included in the study (9 MCI and 26 AD patients).  $^{18}\text{F}$ -FDG uptake in left Brodmann area 31 (owing to the posterior cingulate cortex) correlated negatively with REM sleep latency ( $p = 0.013$ ) and positively with REM sleep ( $p = 0.033$ ).  $^{18}\text{F}$ -FDG uptake in the hippocampus was negatively associated with sleep onset latency ( $p = 0.041$ ). Higher CSF orexin levels were associated with higher sleep onset latency ( $p = 0.042$ ), Non-REM stage 1 of sleep ( $p = 0.031$ ), wake after sleep onset ( $p = 0.028$ ), and lower sleep efficiency ( $p = 0.045$ ). CSF levels of  $\text{A}\beta_{42}$  correlated negatively with the wake bouts index ( $p = 0.002$ ). CSF total-tau and phosphorylated tau levels correlated positively with total sleep time ( $p = 0.045$ ) and time in bed ( $p = 0.031$ ), respectively.

**Conclusion:** Sleep impairment, namely sleep fragmentation, REM sleep dysregulation, and difficulty in initiating sleep correlates with AD biomarkers, suggesting an effect of sleep on the pathological processes in different AD stages. Targeting sleep for counteracting the AD pathological processes represents a timely need for clinicians and researchers.

Keywords: Alzheimer's disease, biomarkers, brain, cerebrospinal fluid, dementia, orexin, positron emission tomography, sleep

## INTRODUCTION

Sleep is commonly impaired in patients with Alzheimer's disease (AD) [1–4]. Compelling evidence demonstrated that REM sleep is progressively fragmented and reduced, slow wave sleep becomes lower, the sleep-wake cycle becomes irregular, and

\*Correspondence to: Claudio Liguori, MD, PhD, Department of Systems Medicine, University of Rome "Tor Vergata", Via Montpellier 1, 00133, Rome, Italy. Tel.: +390620902107; Fax: +390620902116; E-mail: dott.claudioliguori@yahoo.it; ORCID: 0000-0003-2845-1332.

nocturnal sleep losses continuity and stability in the AD continuum [2, 5–9]. In particular, transversal studies including patients with mild cognitive impairment (MCI) and dementia due to AD demonstrated a progressive deterioration of sleep quality and continuity [10]. Recently, the bidirectional relation between sleep and AD has been postulated with several studies highlighting that sleep disturbances can increase the risk of developing AD and hasten amyloid- $\beta$  ( $A\beta$ ) pathology [1–4]. Increasing evidence suggests that sleep deteriorates in parallel with deposition of  $A\beta$  in AD [11–14]. Moreover, sleep dysregulation may occur earlier than cognitive decline, or can be coupled with the impairment of cognitive performance along the AD progression [12–14].

Consistently, previous studies demonstrated that sleep impairment, featured by increased sleep onset latency (SOL), decreased sleep efficiency (SE), or reduced total sleep time (TST), is associated with a greater  $A\beta$  burden measured by positron emission tomography (PET) and reduced cerebrospinal fluid (CSF)  $A\beta_{42}$  levels [5, 15–18].

Considering the significant role that sleep presents in the neurodegenerative AD trajectory, the impact of sleep dysregulation in MCI and AD patients and its modulating effect on the commonly used biomarkers for AD diagnosis is not completely known. Therefore, the present study aimed to evaluate the association among sleep impairment measured by polysomnography (PSG), CSF biomarkers ( $A\beta_{42}$  and tau proteins), and cerebral glucose metabolism evaluated using  $^{18}\text{F}$ -fluoro-deoxy-glucose PET ( $^{18}\text{F}$ -FDG-PET) in a cohort of patients affected by AD, either with MCI or dementia. Moreover, considering the role that orexin has in AD pathology [19], the correlation among CSF orexin levels, cerebral glucose metabolism and sleep in patients with MCI and dementia due to AD was also assessed.

## METHODS

### *Participants and study design*

This study included a retrospective cohort, which was already described in previous studies, of MCI due to AD patients and AD patients, selected on the basis of the additional  $^{18}\text{F}$ -FDG-PET performed at the time of the other analysis [20, 21]. All patients were recruited at the Neurology Unit of the University Hospital of Rome “Tor Vergata”, and underwent physical and neurological examination (including

Mini-Mental State Examination [MMSE] and Clinical Dementia Rating [CDR]), PSG, lumbar puncture for CSF AD biomarker and orexin analysis, and  $^{18}\text{F}$ -FDG-PET. All patients with AD included in this study received the diagnosis of MCI or AD according to the diagnostic guidelines and biomarker analysis [22, 23].

Patients were required to fulfil the following inclusion criteria: no additional neurological or psychiatric disease; and no intake of CNS active drugs. Exclusion criteria for patients were: systemic and/or neurologic infectious, inflammatory, or autoimmune diseases; diabetes; diagnosis of primary sleep disorders or other conditions interfering with sleep quality, such as symptomatic obstructive pulmonary disease and uncontrolled seizures; abnormal cell count ( $>4$  cells/mcL) at the CSF sample analysis; history of alcohol or other substance abuse; intake of caffeine or substances interfering with sleep quality at the time of analysis.

A group of age and sex-matched controls was also recruited at the Department of Biomedicine and Prevention at the University of Rome “Tor Vergata”. All the controls underwent physical and neurological examinations and  $^{18}\text{F}$ -FDG PET. Specifically, the control group included patients undergoing clinical neurologic examination and  $^{18}\text{F}$ -FDG PET for suspected malignancies, which were ruled out after diagnostic investigations.

The study protocol was considered observational and approved by the Ethical Committee of the University Hospital of Rome “Tor Vergata”. Written informed consent was obtained from all the participants in the study.

### *Polysomnographic recordings*

Patients with MCI and AD underwent the PSG study to evaluate their nocturnal sleep, as previously reported (SOMNOscreen; SOMNOmedics GmbH) [7, 21]. Briefly, the montage consisted of 2 oculographic channels, 3 electromyographic channels (mental and anterior tibialis muscles), and 8 electroencephalographic channels (F4, C4, O2, A2, F3, C3, O1, and A1). PSG recordings were evaluated by experts in sleep medicine (CL, FP, FI) and the following standard PSG variables were included in the analysis: time in bed (TIB, time spent in bed between lights off and lights on), SOL (the interval between lights off and the first sleep epoch), TST (the actual sleep time without sleep onset latency and awakenings), SE (the ratio of total sleep time to time in bed),

REM sleep latency (REML, the interval between sleep onset and the first epoch of REM), stage 1 of non-REM sleep (N1), stage 2 of non-REM sleep (N2), stage 3 of non-REM sleep (N3), REM sleep, and wakefulness after sleep onset (WASO). Sleep stage percentages were calculated over the TST. The wake bouts index, defined as continuous sequences of wake epochs occurring after the beginning of persistent sleep and then divided by the number of hours of sleep was identified and computed over TIB.

#### *CSF biomarker analysis*

All CSF samples were obtained the day after the PSG recording by lumbar puncture performed in the decubitus position between 8:00 and 9:30 AM, within 1 to 2 hours after morning awakening, using an atraumatic needle. For detailed methods please see previously published studies [20, 24]. The CSF levels of A $\beta$ <sub>42</sub>, total-tau (t-tau), phosphorylated tau (p-tau), and orexin were determined according to previously published standard procedures [25–27].

#### *PET/CT scanning protocol*

All subjects were intravenously injected with <sup>18</sup>F-FDG (dose range 185–295 megabecquerels) and hydrated with 500 mL of saline (0.9 % sodium chloride). PET/CT acquisition, using a General Electric VCT PET/CT scanner, started 30 ± 5 min after <sup>18</sup>F-FDG injection and lasted 10 min for all participants. <sup>18</sup>F-FDG-PET studies were performed within one week of the PSG recording. The reconstruction parameters were as follows: ordered subset expectation maximization, 4 subsets and 14 iterations; matrix 256 × 256; full width at half maximum (FWHM): 5 mm [28].

#### *Statistical analysis*

Data were analysed using the commercial statistical software SPSS (IBM Corporation, Armonk, NY, USA) [29]. Data are reported as mean and standard deviation or medians and interquartile ranges for continuous variables, whereas categorical data are reported as counts and percentages. The differences between MCI and AD patients groups for demographic, clinical, PSG, and CSF data were tested using Mann-Whitney U-test.

Statistical parametric mapping 12 (SPM12) implemented in MATLAB 2018a was used to analyse PET

scans in this study (<https://www.fil.ion.ucl.ac.uk/spm/software/spm12/>). PET data were converted from DICOM to NIfTI format using MRICro software available at <https://www.nitrc.org/projects/mricron> and then subjected to a normalisation process. A bias regularisation was applied (0.0001) to limit biases due to smooth, spatially varying artefacts that modulate the image's intensity and hinder automated images processing. To prevent the algorithm from trying to model the intensity variation due to different tissue types, the FWHM of the Gaussian smoothness of bias was set at a 60-mm cut-off. A tissue probability map implemented in SPM12 was used (TPM.nii). To achieve approximate alignment to the ICBM space template—European brains [30, 31], mutual information affine registration was used with the tissue probability maps [32]. Warping regularisation was set with the following 1 × 5 arrays (0, 0.001, 0.5, 0.05, 0.2). To cope with functional anatomical variability that is not compensated by spatial normalisation and improves the signal-to-noise ratio, smoothness was set at 5 mm, and the sampling distance, which encodes the approximate distance between sampled points when estimating the model parameters, was set at 3.

To blur the individual variations (especially gyral variations) and to increase the signal-to-noise ratio, an 8-mm isotropic Gaussian filter was employed. Before regression analysis, the following parameters and post-processing tools were used: global normalisation (which escalates images to a global value)=50 (using proportional scaling); masking threshold (to help identify voxels with an acceptable signal in them) was set to 0.8; transformation tool of statistical parametric maps into normal distribution; correction of SPM coordinates to match the Talairach coordinates, subroutine implemented by Matthew Brett (<https://www.mrc-cbu.cam.ac.uk/Imaging>). Brodmann areas (BA) were identified at a range from 0 to 3 mm from the corrected Talairach coordinates of the SPM output isocenter using a Talairach client available at <https://www.talairach.org/index.html>. As proposed by Bennett et al. [33], SPM t-maps were corrected for multiple comparisons using the false discovery rate ( $p \leq 0.05$ ) and corrected for multiple comparisons at the cluster level ( $p \leq 0.001$ ). The level of significance was set at 100 (5 × 5 × 5 voxels, i.e., 11 × 11 × 11 mm) contiguous voxels [33]. The following voxel-based comparisons were assessed: controls versus MCI and AD patients and vice versa. All the comparisons were performed using the 'two-sample *t*-test' design model

Table 1  
Patients' demographic and clinical data

	Total AD patients (n = 35)	Patients' subgroup	
		MCI patients (n = 9)	AD patients (n = 26)
		Mean±SD	Mean±SD
Mean age, y	70.66 ± 7.18	70.33 ± 7.70	70.77 ± 7.15
Sex, n. (%)			
Male	19 (54.3%)	5 (55.6 %)	14 (53.8 %)
Female	16 (45.7 %)	4 (44.4 %)	12 (46.2%)
Disease duration in years	2.89 ± 1.99	2.11 ± 1.05	3.16 ± 2.18
MMSE score	19.88 ± 5.09	25.84 ± 1.13	17.82 ± 4.20
CDR score	1.71 ± 1.07	0.50 ± 0.00	2.13 ± 0.91

AD, Alzheimer's disease; MCI, mild cognitive impairment; SD, standard deviation; MMSE, Mini-Mental State Examination; CDR, Clinical Dementia Rating.

available in SPM12 [28]. Sex and age were used as covariates in the analyses.

The cluster obtained in the regression analysis was then exported by means of the WFU Pick-atlas tool implemented in SPM 12. Specifically, the mean signal intensities calculated from each cluster within each subject were normalised to the average intensities of the Pons volume of interest, as previously suggested by Pagani and colleagues [34]. To determine whether the normalised  $^{18}\text{F}$ -FDG values for the studied cluster were Gaussian distributed, the D'Agostino K-squared normality test was applied (with the null hypothesis being normal distribution). Only the BA showing a significantly significance on  $^{18}\text{F}$ -FDG uptake (i.e., highest Z-scores; Supplementary Table 1) in the comparison between patients to controls were then selected for the correlations. Correlations between the CSF biomarker levels,  $^{18}\text{F}$ -FDG PET, and PSG data in all AD patients were performed using Kendalls' tau b correlation test. Statistical significance was set at  $p \leq 0.05$ .

## RESULTS

Thirty-five patients with biomarkers consistent with AD pathology (54.3% male, mean age of  $70.66 \pm 7.18$ ) were included in this study. Nine patients were diagnosed as affected by MCI due to AD (55.6% male, mean age of  $70.33 \pm 7.70$ ) and 26 patients with AD (53.8% male, mean age of  $70.77 \pm 7.15$ ). Patients' demographic and clinical characteristics are reported in Table 1.

With regard to CSF biomarkers, no significant differences were found between the MCI and AD groups (Table 2). From the group differences regarding PSG data, patients with AD showed a higher duration in TIB and WASO, as well as a lower SE when com-

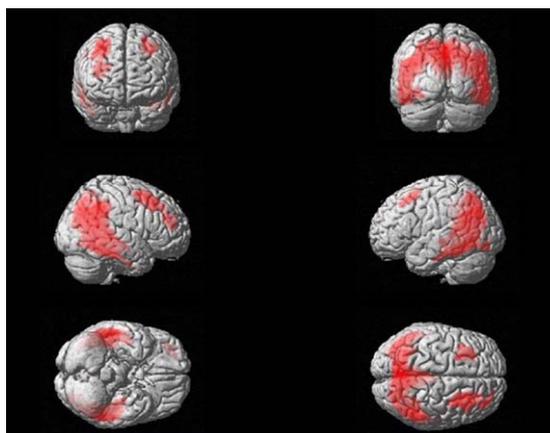


Fig. 1. Three-dimensional brain rendering displaying the results of SPM comparison between patients and controls, which shows a wide cluster that involves bilateral parietal and temporal lobes and frontal cortex. For the SPM analysis, see Supplementary Table 1.

pared to the patients with MCI due to AD (Table 2). No other significant differences were documented for PSG data between these two groups.

In the  $^{18}\text{F}$ -FDG PET analysis, the comparison between patients and controls lead to the detection of a wide cluster of reduced glucose metabolism in patients with MCI and AD. As reported in supplemental Table 1, this cluster reaches its highest significance in the following BA: 31, 7, and 19, which correspond to cingulate gyrus and praecuneus. Nevertheless, the wide cluster involves also parietal and temporal lobes, which are known to be substantially affected by AD (see Fig. 1). Considering these results, exclusively the BA showing the high significance (i.e., highest Z-scores) in the comparison between patients and controls were included in the correlation analysis.

Regarding the correlations between clinical, CSF, PSG, and  $^{18}\text{F}$ -FDG PET data, a significant associa-

Table 2  
Cerebrospinal fluid biomarkers and polysomnographic data of AD patients

	AD patients (n = 35)	Differences between groups				Mann-Whitney test	
		MCI due to AD (n = 9)		AD patients (n = 26)		U	p
		Median	25-75 <sup>th</sup> Percentile	Median	25-75 <sup>th</sup> Percentile		
CSF data							
A $\beta$ <sub>42</sub>	347.94 ± 242.26	312.00	139.00 – 460.50	315.00	207.50 – 399.00	131.0	0.616
t-tau.	660.20 ± 255.85	674.00	516.00 – 905.00	643.50	468.00 – 789.25	106.0	0.697
p-tau	94.26 ± 43.62	84.00	71.00 – 110.50	87.50	60.00 – 114.25	114.0	0.926
Orexin	147.73 ± 35.24	139.60	118.10 – 152.30	152.10	122.05 – 171.85	143.5	0.323
PSG data							
Total Time in Bed	518.02 ± 71.76	460.00	430.00 – 524.00	536.0	506.01 – 584.25	176.0	0.025
Total sleep time (min)	367.71 ± 67.45	382.70	334.00 – 422.50	363.0	335.84 – 404.44	101.0	0.565
Sleep efficiency (%)	71.52 ± 12.22	82.50	72.60 – 88.05	69.50	61.40 – 76.88	57.50	0.023
Sleep onset latency (min)	23.18 ± 26.64	17.00	8.00 – 21.50	18.50	2.00 – 31.25	126.0	0.753
REM sleep latency (min)	206.24 ± 145.34	140.50	41.00 – 251.00	208.50	122.75 – 298.00	151.0	0.210
REM (%)	6.26 ± 4.72	9.60	3.70 – 12.55	5.45	1.20 – 9.38	78.0	0.149
N1 (%)	34.16 ± 11.89	32.00	23.45 – 38.35	37.75	29.51 – 43.10	158.0	0.128
N2 (%)	49.03 ± 10.75	48.60	40.05 – 58.40	50.55	41.85 – 56.23	116.0	0.985
N3 (%)	10.19 ± 8.46	12.80	2.20 – 18.15	6.75	2.03 – 16.98	108.5	0.753
WASO (min)	150.70 ± 76.49	102.40	54.50 – 137.00	163.70	115.75 – 222.50	188.0	0.006
Wake bouts index	4.45 ± 1.99	4.30	2.65 – 5.80	4.00	2.75 – 6.10	124.0	0.810

AD, Alzheimer's disease; MCI, mild cognitive impairment; SD, standard deviation; CSF, cerebrospinal-fluid; A $\beta$ <sub>42</sub>, amyloid-beta 42; t-tau, total tau; p-tau, phosphorylated tau; PSG, polysomnographic, t-tau; WASO, wake after sleep onset; REM, Rapid Eye Movement, N1, Non-REM stage 1; N2, Non-REM stage 2; N3, Non-REM stage 3.

tion was found between the <sup>18</sup>F-FDG uptake in some BA and PSG parameters. <sup>18</sup>F-FDG uptake in left BA31 (owing to the posterior cingulate cortex) correlated negatively with LREM ( $r = -0.294$ ,  $p = 0.013$ ) and positively with REM sleep ( $r = 0.253$ ,  $p = 0.033$ ). <sup>18</sup>F-FDG uptake in the hippocampus correlated negatively with SOL ( $r = -0.245$ ,  $p = 0.041$ ). No significant associations were found between CSF data and brain <sup>18</sup>F-FDG uptake. MMSE scores correlated negatively with several PSG parameters, namely TIB ( $r = -0.357$ ,  $p = 0.003$ ), LREM ( $r = -0.244$ ,  $p = 0.041$ ), and WASO ( $r = -0.379$ ,  $p = 0.002$ ), whereas correlated positively with SE ( $r = 0.317$ ,  $p = 0.008$ ). Moreover, a positive correlation was found between MMSE and <sup>18</sup>F-FDG uptake in the left BA7 (owing to the inferior parietal lobe) ( $r = 0.241$ ,  $p = 0.043$ ).

Levels of CSF orexin correlated positively with SOL ( $r = 0.244$ ;  $p = 0.042$ ), N1 ( $r = 0.257$ ;  $p = 0.031$ ), and WASO ( $r = 0.262$ ;  $p = 0.028$ ). Furthermore, CSF orexin levels were negatively correlated with SE ( $r = -0.238$ ;  $p = 0.045$ ). CSF levels of A $\beta$ <sub>42</sub> only correlated negatively with the wake bouts index ( $p = 0.002$ ). CSF t-tau and p-tau levels were positive associated with TST ( $r = 0.237$ ;  $p = 0.045$ ) and TIB ( $r = 0.247$ ;  $p = 0.031$ ), respectively. Table 3 shows the significant correlations among clinical, CSF, PSG, and <sup>18</sup>F-FDG PET data in the whole group of patients.

## DISCUSSION

The progressive dysregulation of sleep and the sleep-wake cycle has been shown in the AD continuum [2, 10]. In the present study, as expected, sleep fragmentation was more evident in patients with dementia than in those with MCI due to AD. In particular, SE was lower in AD patients than in MCI subjects, which suggests a progressive sleep dysregulation occurring in parallel with the progression of AD pathology. However, the association between sleep impairment and the biomarkers of neurodegeneration in this continuum is not yet completely elucidated. Hence, this study aimed to shed light on this aspect in order to investigate sleep dysregulation in AD pathology and its association with CSF and <sup>18</sup>F-FDG PET biomarkers of neurodegeneration and neuronal injury. In the current study, the correlations between brain <sup>18</sup>F-FDG uptake and PSG data showed that sleep impairment was associated with the biomarkers of neurodegeneration in a sample of both MCI and AD patients. Consistently, sleep macrostructure is altered from the early stages of AD, with evidence of less continuous sleep, slow-wave sleep impairment, REM sleep reduction in AD patients. Moreover, these sleep alterations may also occur before the onset of cognitive impairment and increase the risk for dementia [2,

Table 3  
Correlations among clinical features, <sup>18</sup>F-FDG PET, CSF, and PSG data

	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	
1. MMSE	1																								
2. CSF t-tau	0.014	1																							
3. CSF p-tau	0.005	<b>0.602**</b>	1																						
4. CSF A $\beta$ <sub>42</sub>	-0.066	-0.024	0.073	1																					
5. CSF orexin	-0.039	0.030	0.019	-0.002	1																				
6. TIB	<b>-0.357**</b>	0.217	<b>0.257*</b>	-0.013	0.132	1																			
7. TST	0.031	<b>0.237*</b>	0.166	0.135	-0.034	<b>0.247*</b>	1																		
8. SE	<b>0.317**</b>	0.064	0.039	0.062	<b>-0.238*</b>	-0.229	<b>0.525**</b>	1																	
9. SOL	-0.108	0.099	0.012	-0.100	<b>0.244*</b>	<b>0.245*</b>	0.051	-0.148	1																
10. LREM	<b>-0.244*</b>	0.049	-0.054	0.007	0.132	<b>0.278*</b>	0.045	-0.175	0.119	1															
11. N1 (%)	-0.122	0.146	0.179	-0.003	<b>0.257*</b>	0.210	0.008	-0.148	0.078	0.089	1														
12. N2 (%)	-0.094	-0.135	-0.178	-0.019	-0.019	0.037	0.013	0.019	<b>-0.237*</b>	<b>0.270*</b>	<b>-0.246*</b>	1													
13. N3 (%)	0.088	-0.210	-0.081	-0.061	-0.220	-0.133	-0.106	0.067	0.082	<b>-0.180</b>	<b>-0.520**</b>	-0.091	1												
14. REM (%)	0.175	0.071	0.019	0.157	-0.198	-0.206	0.067	0.221	0.053	<b>-0.519**</b>	-0.219	<b>-0.370**</b>	0.192	1											
15. WASO (%)	<b>-0.379**</b>	-0.024	0.042	-0.045	<b>0.262*</b>	0.401**	<b>-0.353**</b>	<b>-0.823**</b>	0.202	0.205	<b>0.232*</b>	-0.008	-0.131	<b>-0.255*</b>	1										
16. Wake bout index	-0.032	-0.162	-0.144	<b>-0.363**</b>	-0.046	-0.007	<b>-0.432**</b>	<b>-0.319**</b>	0.002	-0.003	-0.017	-0.005	0.088	-0.107	<b>0.258*</b>	1									
17. Hippocampus	0.047	-0.052	0.105	-0.111	-0.054	-0.086	-0.170	0.034	<b>-0.245*</b>	-0.049	0.015	0.128	0.015	-0.111	-0.040	0.165	1								
18. Left BA31	0.207	0.012	0.196	0.168	-0.017	0.019	0.123	0.155	-0.194	<b>-0.294*</b>	0.012	-0.067	-0.035	<b>0.253*</b>	-0.165	-0.078	0.136	1							
19. Left BA7	<b>0.241*</b>	-0.055	0.115	0.108	-0.010	0.025	-0.032	-0.027	-0.092	-0.227	0.126	-0.229	-0.123	0.162	0.024	-0.037	-0.012	<b>0.516**</b>	1						
20. Right BA19	0.180	-0.015	0.051	0.118	0.125	-0.002	-0.099	-0.030	-0.065	-0.133	-0.049	-0.081	-0.035	0.182	0.024	0.091	0.123	<b>0.355**</b>	<b>0.449**</b>	1					
21. Right BA6	-0.014	0.022	0.054	0.077	0.020	0.170	0.079	-0.044	0.000	-0.062	0.156	-0.104	-0.153	0.064	0.104	0.030	-0.129	<b>0.277*</b>	<b>0.439**</b>	<b>0.264*</b>	1				
22. Right BA8	0.102	-0.123	-0.088	0.135	-0.030	0.042	-0.012	0.000	-0.112	-0.079	-0.035	0.040	-0.062	0.115	0.000	0.030	-0.079	<b>0.368**</b>	<b>0.455**</b>	<b>0.375**</b>	<b>0.620**</b>	1			
23. Left BA6	-0.007	0.018	0.068	0.141	0.047	0.184	0.096	-0.054	0.044	-0.139	0.180	-0.162	-0.130	0.145	0.108	-0.047	-0.200	0.287*	<b>0.462**</b>	0.220	<b>0.782**</b>	<b>0.576**</b>	1		
24. Left BA8	0.146	-0.129	-0.054	0.081	-0.003	0.045	0.042	0.034	0.003	-0.227	-0.035	-0.074	-0.049	0.202	-0.020	-0.047	-0.173	0.308**	<b>0.503**</b>	<b>0.274*</b>	<b>0.580**</b>	<b>0.644**</b>	<b>0.664**</b>	1	

\* $p < 0.05$ ; \*\*  $p < 0.01$ . MMSE, Mini-Mental State Examination; t-tau, total tau; p-tau, phosphorylated tau; A $\beta$ <sub>42</sub>, amyloid-beta 42; TIB, time in bed; TST, total sleep time; SE, sleep efficiency; SOL, sleep onset latency; LREM, REM sleep latency; N1, Non-REM stage 1; N2, Non-REM stage 2; N3, Non-REM stage 3; REM, Rapid Eye Movement; WASO, wake after sleep onset; BA, Brain Broadman areas.

10, 35, 36]. It is conceivable that sleep dysregulation is linked to the neuropathology of AD from the early stage of the disease, and this supposition is in line with the recent literature documenting the interplay between sleep impairment and the biomarkers of AD pathology. Moreover, a possible direct effect of sleep fragmentation and dysregulation on AD biomarkers has been hypothesised, even before the MCI stage [2]. In agreement with this evidence, the present study confirmed the association between sleep impairment, namely REM sleep dysregulation and symptoms of insomnia (prolonged SOL), and AD neurodegeneration. Moreover, cognitive decline was associated with both sleep fragmentation and cerebral glucose metabolism impairment. Specifically,  $^{18}\text{F}$ -FDG uptake in inferior parietal lobe correlated with cognitive impairment, while  $^{18}\text{F}$ -FDG uptake in the hippocampus and the CSF levels of  $\text{A}\beta_{42}$  were both associated with longer SOL and higher wake bouts index, respectively. These symptoms of sleep impairment and fragmentation are frequently documented in patients with MCI and AD [2, 10]. Moreover, REM sleep dysregulation, featured by longer REML and reduced amount of REM sleep, also correlated to the impaired  $^{18}\text{F}$ -FDG uptake in posterior cingulate cortex. This data, therefore, extends the findings of previous studies by highlighting the significant REM sleep alteration and its correlation with brain functioning in MCI and AD patients, as well as reinforcing the clinical potential of targeting REM sleep as part of a therapeutic strategy in AD for enhancing cerebral function [10, 37]. Furthermore, CSF AD biomarkers and orexin correlated with sleep dysregulation, confirming the previously shown evidence achieved separately in MCI and AD subjects [7, 10, 21, 38].

Overall, the findings of the present study allow to substantiate the hypothesis of sleep modulation on the AD biomarkers. In agreement with our results, animal model studies documented that sleep may exert a modulating effect on CSF dynamics, thus influencing CSF biomarkers, and also modifying CSF orexin levels and orexin system functioning [39–41]. In humans, sleep impairment and sleep deprivation, even when partial, can increase the CSF orexin levels. The results of the present study merged with these previous reports increase the knowledge about the interplay between CSF orexin levels and sleep in AD [7, 38, 42, 43]. Furthermore, in the present study, and as hypothesised, patients with AD showed a worse sleep quality than patients with MCI due to AD, showing higher sleep fragmentation and poorer

sleep hygiene. These findings are in line with the few previous studies that highlighted how sleep deteriorates in AD staging, suggesting a more disrupted nocturnal sleep as the disease progresses [6, 7]. Nonetheless, the assessment of sleep impairment measured by PSG in relation to the disease staging is still scarce and needs to be better studied, specially taking into consideration the PSG markers of sleep fragmentation and loss of sleep stability and continuity. This study, besides supporting previous findings on sleep impairment in AD, broadens previous knowledge by highlighting the recognition of sleep impairment, and in particular, REM sleep dysregulation, in patients with AD pathology, either in the MCI or dementia stage of the disease, since it significantly correlates with biomarkers of the ongoing neurodegeneration. These findings reinforce the importance of targeting sleep as a potential marker, and not only as a symptom since sleep impairment can accelerate the progression of AD pathology, burden AD patients and caregivers, and influence the neurodegenerative mechanisms [44–46]. Accordingly, sleep can impair cognitive and behavioral symptoms from the preclinical and early stages of AD, and therefore, sleep assessments should be considered in the diagnostic work-up of patients with MCI or AD. It is important to note that sleep impairment can also aggravate the prognosis of the AD neurodegenerative process, as well as impair the patient and caregiver's well-being since represents a disabling neuropsychiatric symptom with multiple consequences. Moreover, considering the association of sleep impairment with AD biomarkers, it is conceivable that sleep improvement, as a therapeutic target in the management of MCI and AD patients, may positively interfere and, thus, counteract the cascade of events fueling the AD progressive neurodegenerative process.

The present study presents some limitations that need to be acknowledged. The sample of patients is small since it was achieved in a single Neurology Unit. Despite the high-specialised investigations performed in our study, a multicentre study to increase the number of patients should be planned in the future. Although hypothesised, the cross-sectional nature of this study and the small sample size of MCI patients did not allow a demonstration of the impact of sleep dysregulation on the AD continuum and how AD may determine changes in sleep patterns. Longitudinal studies with a larger cohort of patients are needed and future correlation analysis should consider separately MCI and AD patients.

Even though high-resolution MRI images can provide better anatomical information than functional PET images, in the present study it was not possible to use MRI-based partial-volume correction due to the retrospective design of our study. Nonetheless, in order to partially mitigate the issues related to the partial volume effect, a smoothing procedure, coupled with the spatial normalisation process, was applied [20, 47]. Specifically, the smoothing procedure allows the suppression and noise due to residual differences in functional and gyral anatomy during inter-subject averaging [48]. To the best of our knowledge, few tools are available in order to mitigate the partial volume effect in brain PET imaging. Indeed, one of the latest tools developed for SPM aimed to improve local regional detection of radiopharmaceutical uptake in corrected images as compared to non-corrected images [49]. Nevertheless, the toolbox was developed specifically for amyloid imaging and does not apply to our case studies thus, future analyses on this topic should include MRI-based partial-volume correction.

Although the control group helped in the identification of BA significantly affected in MCI and AD patients, only three BAs (31, 7, and 19) were considered significant for the correlation analysis. These regions, corresponding to cingulate gyrus and praecuneus, which represent limited structures of the brain that do not play a major role in the AD process. Nevertheless,  $^{18}\text{F}$ -FDG PET analysis detected a wide cluster of reduced glucose metabolism in MCI and AD patients compared to the controls, involving parietal and temporal lobes that are known to be substantially affected by AD. These findings may be partially explained by the inclusion of MCI patients in the analysis, who usually show a lower degree of neurodegeneration. Finally, the lack of a PSG evaluation and a CSF assessment in the control group is also one of the limitations of the present study.

In conclusion, the present study reinforces the critical role of sleep not only in the AD neuropathological processes but also at the different clinical stages of the disease by documenting an association between sleep dysregulation and different AD biomarkers. These findings invite further longitudinal studies to monitor sleep during the AD continuum in order to set trials targeting sleep improvement as a potential therapeutic approach to antagonise the neurodegenerative processes in MCI and AD patients, also considering the association between CSF orexin levels and sleep dysregulation in AD.

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## CONFLICT OF INTEREST

The authors report no conflict of interest.

## DATA AVAILABILITY

The data that support the findings of this study are available from the corresponding author upon reasonable request.

## SUPPLEMENTARY MATERIAL

The supplementary material is available in the electronic version of this article: <https://dx.doi.org/10.3233/ADR-220111>.

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