Liquid biopsy in clinical outcomes and detection of T790M mutation in metastatic non-small cell lung cancer after progression to EGFR-TKI

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Abstract.

BACKGROUND: Liquid biopsy (LB) is used to detect epidermal growth factor receptor (EGFR) mutations in non-small cell lung cancer (NSCLC) and has been demonstrated to have prognostic and predictive value.

OBJECTIVE: To associate the rates of *EGFR* and T790M mutations detected by LB during disease progression after first- or second-generation EGFR-TKIs with clinical characteristics and survival outcomes.

METHODS: From January 2018 to December 2021, 295 patients with advanced EGFR mutant (EGFRm) NSCLC treated with first- or second-generation EGFR-TKIs were retrospectively analyzed. LB was collected at the time of progression. The frequency of EGFR^{T790M} mutations, overall survival (OS), and the clinical characteristics associated with LB positivity were determined.

RESULTS: The prevalence of EGFR^{T790M} mutation detected using LB was 44%. In patients with negative vs. positive LB, the median OS was 45.0 months vs. 25.0 months (p = 0.0001), respectively. Patients with a T790M mutation receiving osimertinib had a median OS of 44 months (95% CI [33.05–54.99]). Clinical characteristics associated with positive LB at progression extra-thoracic involvement, > 3 metastatic sites, and bone metastases.

CONCLUSIONS: Our findings showed that LB positivity was associated with worse survival outcomes and specific clinical characteristics. This study also confirmed the feasibility and detection rate of T790M mutation in a Latin American population.

Keywords: EGFRm NSCLC, liquid biopsy, T790M mutation, osimertinib, ctDNA

1. Introduction

*Corresponding author: Oscar Arrieta, Head of Thoracic Oncology Department, Instituto Nacional de Cancerología. Av. San Fernando #22, Sección XVI, Tlalpan. México City, CDMX, 14080, México. Tel.: +52 55 5628 0400 (ext. 71101); E-mails: ogar@unam.mx and oscar.arrieta.r@gmail.com. ORCID: 0000-0002-1164-3779. Non-small cell lung cancer (NSCLC) is the leading cause of cancer death worldwide [1,2]. The majority of NSCLC patients are diagnosed with advanced-stage disease [3,4]. Patients with NSCLC are treated based on a personalized approach, according to genomic profile, 6

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to ensure better oncological outcomes [5]. The detec-7 tion of epidermal growth factor receptor (EGFR) mu-8 tations produces a paradigm shift in the treatment of 9 NSCLC, which varies according to the geographic re-10 gion in Latin America, and. oscillates between 32–54%. osimertinib. 11 In Mexico, EGFR mutations represent around 36% of 12 druggable alterations, which is higher compared with 13 2. Materials and methods North America and Europe, ranking 10–15% [6,7]. 14 NSCLC patients harboring EGFR mutations have 15 improved oncological outcomes with EGFR-tyrosine 2.1. Ethics approval 16 kinase inhibitor (TKI) therapy [8–11]. According to the 17 results of the FLAURA trial, osimertinib is currently the 18 preferred upfront treatment [8]. However, in many other 19 countries, the upfront therapy for patients with EGFRm under the number Rev/032/20. 20 NSCLC still depends on first- and second-generation 21 TKIs followed by osimertinib in patients who devel-2.2. Experimental subjects 22 oped a positive T790M mutation at disease progression. 23 Among the common mechanism of acquired resistance 24 after this treatment is the acquisition of EGFR T790M 25 mutation in approximately 50–60% of cases [12–14]. 26 EGFR T790M resistance mutation can be detected 27 in tumor tissue (surgical biopsy or cytology specimens) 28 or cell-free circulating tumor DNA (ctDNA) extracted 29 from peripheral blood through liquid biopsy (LB) [15]. 30 New tumor tissue biopsy is the most reliable procedure 31 for detecting the T790M mutation at progression. How-32 ever, this is not always feasible and might be associated 33 with complications due to the invasiveness of tumor 34 biopsy. Other factors, such as tumor heterogeneity and 35 unwillingness, make this method challenging for many to monitor adverse events. 36 patients [16–18]. 37 LB refers to any tumor-derived material circulating 38 through the blood or other body fluid. In lung can-39 cer, circulating tumor cells and ctDNA are the most 40 widely studied substrates [19]. LB has been shown to 41 represent an innovative alternative for patients unable 42 to undergo re-biopsy as a non-invasive method that al-43 lows the detection of EGFR T790M [20]. Other advan-44 tages over tissue samples include the reduced cost, rapid 45 of the EGFR mutation. turnaround time, and potential for longitudinal monitor-46 ing by serial biopsies, showing a high concordance with 47 standard tissue genotyping [20,21]. In addition, some 48 studies have demonstrated that detection of EGFRm 49 ctDNA is associated with worse outcomes and disease 50 burden, which could be predictive of response, increas-51 ing ctDNA levels which may anticipate progression to 52 standard imaging progression by RECIST [22,23]. 2.3. Statistical analysis 53 Scarce information is available on the utility of LB to 54 detect T790M in the LATAM population in a real-world 55 context and its association with clinical outcomes. Thus, 56

this study aimed to determine the rate of T790M mu-57

tation as a mechanism of resistance to first or secondgeneration EGFR-TKI according to plasma ctDNA detected on LB, associate positiveness to clinical characteristics, and evaluate outcomes in those who received

This study was approved by the ethics committee of participating medical institution and was authorized

In this retrospective cohort, we evaluated 295 patients with Stage IV NSCLC treated in one institution between January 2018-December 2021. Patients harboring a sensitive EGFR mutation (delexon19 or L858R) and disease progression in the first-line setting with one first or second-generation EGFR-TKI were included. Patients included had 1) confirmed diagnosis of NSCLC, 2) EGFR mutation, 3) disease progression according to RECIST 1.1 criteria, 4) Treatment with first or second-generation EGFR-TKI. 5) All patients required standard laboratory workups before starting therapy and during treatment according to local policies

All patients performed an LB at radiological progression with the Biocept Target Selector TM ctDNA platform [24]. Each blood samples consist of 20 ml of peripheral blood collected in the CEE-Sure TM tubes (Biocept, San Diego, California, USA). EGFR mutation assay is a quantitative Real-Time PCR-based mutant enrichment assay with selective blocking of EGFR wildtype amplification. The amplification products are purified and subject to sequencing to confirm the presence

Two expert medical oncologists extracted all clinical and pathological data from electronic medical records. Collected data included the patient's clinical characteristics at baseline and disease progression, and date, results, and number of LBs before a positive result.

Categorical variables were summarized as frequencies and percentages, while continuous variables were reported as mean or median with their corresponding 100

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Table 1Baseline characteristicsCharacteristics n^a %Sex, female19666.4Age, years66.4	
Characteristics n^a %Sex, female19666.4	
Sex, female 196 66.4	
A de vears	
Mean \pm Std. Dev. 62.5 \pm 12.31	
Smoking status	
Ever smokers 61 20.6	
Nonsmokers 243 79.4	
Tobacco index, pack/yrs.	
Mean \pm Std. Dev. 10.42 ± 11.12	
Body Mass Index (BMI)	
Mean \pm Std. Dev 24.24 ± 4.34	
Comorbidities	
Hypertension 85 28.0	
Diabetes Mellitus Type 2 44 15.0	
Obesity 19 15.5	
Chronic Obstructive pulmonary disease 6 2.0	
Heart failure 4 1.3	
ECOG PS scale	
0–1 271 92.1	
≥ 2 24 7.9	
Adenocarcinoma 285 96.1	
Squamous or mixed histology 10 3.9	
LADC subtype	
Lepidic 25 8.5	
Acinar 84 28.5	
Papillary 24 8.1	
Solid 76 25.8	
Micropapillary 6 2.0	
Not otherwise specified 80 27.1	
Histological grade	
Well-differentiated 16 5.4	
Moderately differentiated 116 39.3	
Poor differentiated 113 38.3	
Not otherwise specified 50 16.9	
Clinical stage IV 295 100	
First line treatment	
1st generation TKI (Erlotinib, Gefitinib) 164 55.6	
2d generation TKI (Afatinib) 131 44.4	
EGFR mutations at diagnosis	
Exon 19 Del 197 66.0	
L858R Exon 21 89 30.1	
Other mutations 9 3.9	

Notes. ECOG: Eastern Cooperative Oncology Group performance status scale. LADC: lung adenocarcinoma. EGFR: epidermal growth factor receptor. ^aTotal number of subjects = 295.

dispersion measures. Progression-free survival (PFS), 101 intracranial progression free-survival (icPFS), and 102 Overall survival (OS) were estimated by the Kaplan-103 Meier method, and statistical differences among groups 104 of interest were calculated with the Log-Rank test 105 method. PFS was defined from the beginning of the sec-106 ond line treatment until disease progression according 107 to RECIST Criteria or death; icPFS was defined from 108 diagnosis to the date of appearance of new brain metas-109 tasis. OS was defined as the period from diagnosis to 110 death or loss of follow-up. A Cox proportional hazard 111 model was performed for the multivariate analysis. A 112

Table 2 Clinical characteristics at progression Characteristics n^{a} % Metastatic sites at progression 30 47.6 CNS Bone 28 38.1 Pleura 32 50.8 Adrenal glands 12 19 Liver 13 20.6 Type of progression Intrathoracic 89 30.2Extra-thoracic 206 69.8 Second-line therapy Carboplatin Pemetrexed 139 47.1 Other systemic treatment 30 10.2 51 17.3 Other TKI Osimertinib 22 7.5 TKI + local control 19 6.4 None 26 8.8

Notes. CNS: central nervous system. TKI: tyrosine kinase inhibitor. ^aTotal number of subjects = 295.

significant p-value was < 0.05 in a two-sided test. For these statistical analyses, SPSS version 23 was used.

3. Results

3.1. Clinical characteristics

Two hundred ninety-five patients were analyzed. Of 117 the total, 66.4% were women, 79.4% had never smoked, 118 92.1% had an ECOG performance status (PS) of 0 to 1, 119 and 96.61% were adenocarcinomas. Regarding muta-120 tions at the time of diagnosis, 66% had exon 19 dele-121 tions (exon19del), and 30.1% had a point mutation of 122 exon 21 L858R. The most frequent metastatic sites at 123 diagnosis were pleura (50%), followed by the central 124 nervous system (47.6%) and bones (38.1%). All patient 125 demographics and clinical characteristics are summa-126 rized in Table 1.

3.2. Clinical characteristics at disease progression

At the time of progression, 69.8% had extra-thoracic progression, whereas 30.2% had exclusively an intrathoracic affection (Table 2). All patients had an initial LB at the first disease progression, 23 underwent a second LB at the next progression, and one underwent a third LB after a third progression.

3.3. Detection of T790M in liquid biopsy

Of the 295 analyzed patients, 122 (41.4%) had positive T790M at the first disease progression. In most

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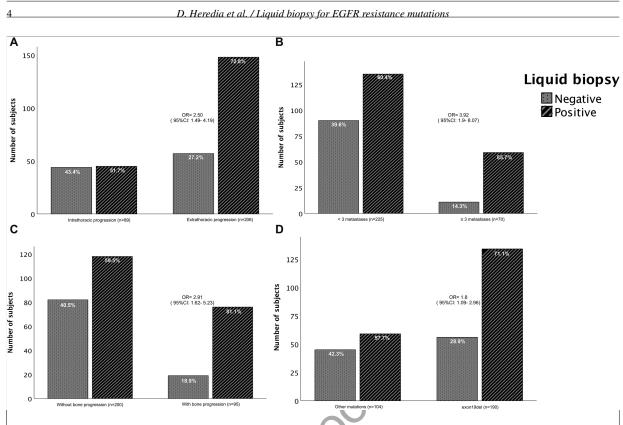
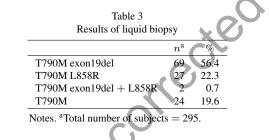


Fig. 1. Detection of ctDNA positivity or negativity by liquid biopsy. Based on clinicopathological characteristics: a) type of thoracic progression $(X_a^{1df} = 12.445, p \le 0.001)$, b) number of metastases $(X_a^{1df} = 15.290, p \le 0.001)$, c) bone progression $(z_a = 13.418, df = 1, p \le 0.001)$, and d) type of EGFR mutation ($X_a^{1df} = 5.372, p \le 0.020$). Pearson's chi-square test was applied to nominal variables with an expected count of < 5.372Odds ratio (OR) with CI: confidence interval at 95 % (95 % CI). Statistical significance was set at *p < 0.05.



of the cases, T790M was not the only detected al-138 teration; 69 (56.4%) combined with exon19del, 27 139 (22.3%) with L858R, and 2 (0.7%) with exon19del and 140 L858R. T790M alone was present in 24 (19.6%) pa-141 tients. In additional nine patients, subsequent LBs were 142 performed, and the T790M was detected for a total of 143 131 patients (44%) (Table 3). 144

3.4. Characteristics associated with positivity in liquid 145 biopsy 146

The patients with exon19del at extra-thoracic pro-147 gression, more than three sites of metastases involved 148 at baseline, and the presence of bone metastases at pro-149

gression had a higher LB detection rate. In patients 150 with initial exon19del, LB positivity was 71%. Mean-151 while, the positive rate in patients with other muta-152 tions was 57.7% (Fig. 1). Similarly, patients with extra-153 thoracic progression had a higher LB detection rate (72.8%) compared with patients with intrathoracic progression (51.7%) (p = 0.001). Patients with three or more metastatic sites at disease progression had a higher positivity of LB (85.7%) in comparison with patients with two or less than two metastatic sites (60.4%) (p =0.0001). Also, patients with bone involvement at disease progression had an LB-positive rate of 81.1%, compared with 59.5% in patients without bone affection. No differences in LB positivity were found in patients with or without brain metastases.

3.5. Survival outcomes

The median OS for all patients was 30 months (95%) 166 CI: 26.06–33.93). After TKI progression, the median 167 PFS was eight months (95% CI: 6.49–9.50). The me-168 dian OS was 45.0 months (95% CI: 37.03-52.96) vs. 169

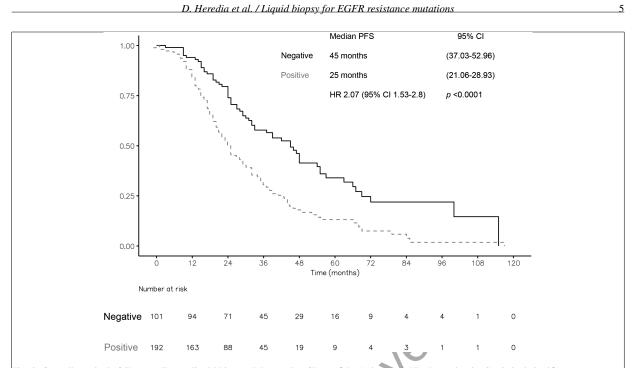


Fig. 2. Overall survival (OS) according to liquid biopsy (LB) results. CI: confidence interval. HR: hazard ratio. Statistical significance was set at p-value < 0.05.

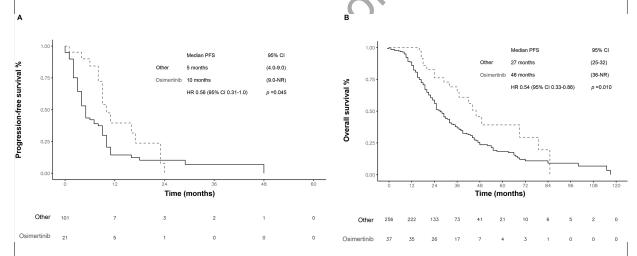


Fig. 3. Survival of patients with T790M mutation at disease progression. A) PFS of patients treated with osimertinib versus those treated with other drugs. B) Overall survival (OS) in patients treated with osimertinib vs. other treatments. PFS: progression-free survival. CI: confidence interval. NR: not rated. HR: hazard ratio. Statistical significance was set at a p-value < 0.05.

25.0 months [95% CI: 21.06–28.93; HR 2.00 (1.48, 170

2.71); p = 0.0001 in patients with negative vs positive 171

LB respectively. Of note, no differences in OS were 172

- observed, between EGFR sensitive mutations (exon-173
- del19 and L858R) and T790M mutation, 23.0 months 174
- (95% CI: 19.27–26.72) vs 27.0 months (95% CI: 21.51– 175
- 32.49) (Fig. 2). Of note, no differences in OS were ob-176
- served, between EGFR sensitive mutations (exondel19 177
- and L858R) and T790M mutation, 24.0 months (95%) 178

CI: 20.0–32.0) vs 25.0 months (95% CI: 21.0–32.0) (Fig. S1)

The median PFS in patients with a T790M muta-181 tion detected at first progression (n = 122) by LB and 182 who were treated with second-line osimertinib was 10.0 months (95% CI 9.0-14.24), whereas, in patients treated with other therapy, the median PFS was 5.0 months [95% CI 4.0-9.0; HR 0.56 (0.31.1.00); p = 0.045](Fig. 3A). 187

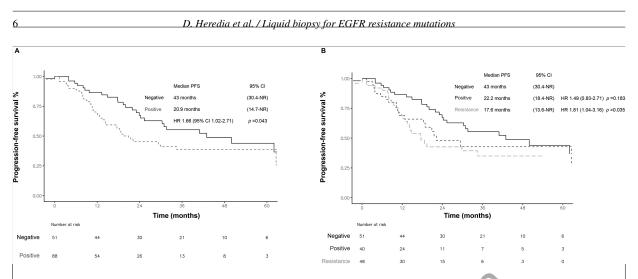


Fig. 4. icPFS according to liquid biopsy results. A) positive and negative. B) Positivity, sensitizing, or resistance mutations. icPFS: intracranial progression-free survival. CI: confidence interval. NR: not rated. HR: hazard ratio. Statistical significance was set at a *p*-value < 0.05.

In patients treated with osimertinib at any time (n =37), irrespective of the line of treatment, the median OS was 46 months (95% CI 36.0–NR), and in those treated with other therapy was 27 months [95% CI 25.0–32; HR 0.54 (0.33–0.86); p = 0.010] (Fig. 3B).

In the case of intracranial disease progression in patients without baseline brain metastases (n = 139), the median icPFS was 20.9 months with positive LB compared with a median of 43.0 months in patients with a negative liquid biopsy (HR 1.66; 95% CI 1.02–2.71; p = 0.043).

According to the type of mutation detected at progression in the LB, patients with a positive T790M mutation had significantly lower icPFS (17.6 months; HR 1.81 95% CI 1.04–3.16; p = 0.035) compared with those with only EGFR sensitive mutations (22.2 months; HR 1.49; 95% CI 0.83–2.71) (Fig. 4).

205 **4. Discussion**

In our study, we demonstrated that the positivity of 206 liquid biopsy at disease progression to a first or second-207 generation TKI was associated with worse survival out-208 comes including a shorter icPFS, and the detection 200 rate of T790M mutation as the main mechanism of 210 resistance in metastatic EGFRm NSCLC was 44% in 211 Hispanic population. Moreover, we could identify that 212 those patients with more than three metastatic extra-213 thoracic diseases, and bone metastasis were associated 214 with a greater probability of a positive result on LB. 215

A T790M as main resistance mechanism after first line EGFR-TKI treatment has been reported to be
 around 47.1% in Hispanic population in tumor speci-

mens [14]. In largest series that assessed mechanisms of
acquired resistance to EGFR-TKI therapy, 63% of the
patients developed T790M mutations. Of note, biop-
sies were obtained by the least invasive procedure and
typically consisted of either a fine-needle aspiration or
image-guided core biopsy, but not employed LB [13].
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More complex scenarios have been identified in
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pretreated patients, in which the presence of T790M 226 and persistence of common EGFR-activating muta-227 tions (exon 19 del and L858R) has been associated 228 with poor prognosis in patients with advanced EGFRm 229 NSCLC [25]. Our study found that in patients with a 230 positive LB, the detection of T790M mutation alone 231 was observed in 19.6%. In contrast, most patients have 232 persistence of EGFR sensitive mutations, T790M plus 233 exon 19 del in 56.4%, T790M plus L858R in 22.3%, 234 and more complex combinations (T790M plus exon 235 19 del plus L858R) were extremely rare (< 1%). In 236 other studies, Liang and colleagues reported similar 237 results. In a first-line post-TKI scenario, 53% of pa-238 tients had coexistence of T790M mutation and exon19 239 del, and T790M mutation plus L858R was observed in 240 36% [26]. 241

One meta-analysis confirmed that the T790M mu-242 tation used to coexist more frequently with an L858R 243 than with the exon19del [odds ratio 1.65; 95% CI (1.17 244 to 2.32)], which potentially might explain the more ag-245 gressive biology of L858R tumors [25]. However, this 246 information has been contradictory with further evi-247 dence which suggested a stronger association between 248 the emergence of T790M and the persistence of the pri-249 mary activating alteration, exon19del, among patients 250 with acquired resistance to TKI (53% vs. 36%; OR 251 1.87; p < 0.001 [26]. In our cohort, we found that 252

exon19del was associated with a higher LB positive rate 253 which is in line with Liang et al. study. Different results 254 in the survival outcomes can be related to subsequent 255 treatments. Patients who develop T790M mutation are 256 candidates to osimertinib at progression to a first-line 257 TKI, which can impact survival outcomes based on the 258 positivity rate. Even so, the type of EGFR mutation 259 has been demonstrated to be a factor associated with 260 response to first-line EGFR-TKI treatment. In clinical 261 trials, L858R mutation has been related to a shorter me-262 dian PFS, even for patients receiving osimertinib [27]. 263 Although the feasibility and reliability of LB have 264 been confirmed in clinical practice [28–30], the test's 265 sensitivity is related to the detection limits of the tech-266 nique and the characteristics of ctDNA. Based on the 267 different platforms, in some assays (BEAMing,) the 268 detection rate of LB is around 70% [31], along with 269 this, a relatively low sensitivity (60%) in comparison 270 with high specificity (80-90%) has been reported in an 271 independent meta-analysis [20,32]. Due to this possi-272 ble low sensitivity, the current recommendation is that 273 patients with negative LB should undergo tissue test-274 ing for T790M [33,34]. In our study, the LB detection 275 rate was 41.4%, then the detection rate went up to 44%276 with subsequent testing with the same technique. This 277 reinforces that even the LB could be a suitable option. 278 for some patients to detect additional resistance mech-279 anisms if tissue is unavailable or the risk of undergo-280 ing an invasive procedure is too high. In line with our 281 work, previous evidence has confirmed an additional 282 12.5% of detection rate in patients with an initial nega-283 tive LB and then tested positive for T790M at a second 284 LB [35]. In another study that used a median of 3 LB, 285 the prevalence of T790M was 34.5% [36]. 286

In addition to the procedure itself and technique em-287 ployed in the ctDNA analysis, some clinical charac-288 teristics have gained relevance in the performance and 289 sensitivity of the LB. In previous studies, the number 290 of metastatic sites which reflects disease burden was 291 associated with a higher positive rate [37]. We were 292 able to identify the most common metastatic sites of 293 metastases in the present cohort; pleura (47%), central 294 nervous system (47%), and bone (38.1%). Remarkably, 295 most patients had a moderate disease burden between 296 2 or 3 involved sites. We found that in patients with 297 three or more sites of metastasis, especially when the 298 affection was extra-thoracic and the bone was involved, 299 the detection rate of the LB was significantly higher. 300

Patterns of progression and sites have gained rele-301 vance in clinical practice to consider subsequent strate-302 gies and overcome resistance in EGFR NSCLC extra-303

thoracic disease, as the main site of progression has 304 been associated with a positive LB [38]. However, one-305 half of patients progressed within the lung as the initial 306 progression after TKI treatment [38,39]. Al Halabil et 307 al., in a retrospective study of patients with NSCLC 308 treated with first and second-generation TKI, found that 309 the main sites of progression were those initially in-310 volved in 50% of cases, being the intra-thoracic the 311 most common site [38]. This was confirmed by Pa-312 tel and colleagues, in which 17.5% had extra-thoracic 313 failure, and 22.2% had intra- and extra-thoracic pro-314 gression [38]. In contrast with these previous reports, 315 our study found that extra-thoracic progression was the 316 main site of progression (69%), and this finding was 317 associated with a higher detection rate by LB and bone 318 lesions, in line with previous reports [28,35]. 319

ctDNA shedding is related to the disease burden [22, 320 40] and can help distinguish between residual disease 321 or anticipated imaging changes [41]. This might ex-322 plain why LB has been associated with worse survival, 323 especially in patients with a persistent EGFR-sensitive 324 mutation. This was replicated in our study in which 325 patients with a positive LB had worse survival indepen-326 dent of the type of mutation [42]. Despite these find-327 ings, starting osimertinib at the moment of the T790M 328 mutation detection is not justified, which precedes the 329 radiological progression [43]. In this regard, shedding 330 ctDNA with persistent EGFR-sensitive mutation could 331 be the most essential factor associated with survival. 332

We found that CNS disease was not associated with 333 LB positivity. However, in patients without baseline brain metastases, the icPFS was significantly shorter in patients with a positive LB at progression. This lower icPFS was statistically significant in patients with a positive T790M mutation. However, a trend to a worse icPFS was also observed for patients with a persistent EGFR sensitive mutation (delexon19 and L858R) regardless of the T790M status. The prognostic utility of 341 liquid biopsy for intracranial PFS is particularly inter-342 esting, considering the increased risk of brain affection 343 in patients with EGFR alterations. Very limited data 344 have assessed this key point; based on our findings, it 345 will be very attractive to assess prospectively the prog-346 nostic and predictive value of liquid biopsy regarding 347 central nervous system efficacy in this subpopulation. 348 Also, the potential utility of liquid biopsy is to guide 349 clinicians to develop strategies with high brain pen-350 etration in the subgroup of patients with the highest 351 risk. 352

Acquired T790M has been associated with indolent growth and a favorable prognosis compared to other ac-

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quired resistance mechanisms. In this sense, the absence 5. Conclusions 355 of T790M after progression probably indicates the pres-356 This study supports a comparable rate of T790M deence of alternative resistance mechanisms, which are 357 associated with a higher disease burden and poorer pertection by liquid biopsy in the Latin American popu-358 lation, and a higher detection rate was associated with formance status, contributing to the shorter survival of 359 clinical characteristics. Moreover, it emphasizes that the these patients [42]. As demonstrated in our study, pa-360 detection of ctDNA by LB is feasible after progression tients with better outcomes were those with a positive 361 to first-line EGFR therapy and has a prognostic and T790M mutation who received osimertinib treatment, 362 potentially predictive value in EGFRm NSCLC. Pahighlighting the importance of detecting T790M and 363 tients with positive LB results also had shorter intracrareceiving the appropriate treatment. 364 nial progression-free intervals, a finding that might be In clinical trials, such as the AURA3 [44] trial, a low confirmed in a further prospective analysis. proportion of patients (51.2%) had T790M mutation, 366 as assessed using plasma ctDNA. However, the T790M 367 mutation detected by ctDNA has been demonstrated to 368 Acknowledgments be a surrogate marker for T790M in tumor tissues and 369 is predictive of treatment response [27]. In our study, 370 The authors would like to recognize the work of the median PFS of patients with the T790M mutation 371 the team of the Thoracic Oncology Department at our treated with osimertinib was similar to that reported in 372 institution for the support and care of patients. the AURA-3 trial [44,45]. In another trial [31,46], no 373 difference in outcomes was found with osimertinib us-374 ing plasma or tumor tissue to detect T790M, supporting 375 **Author contributions** the preferred use of LB and leaving tissue biopsy only 376 for patients with negative LB [31,46]. 377 Conception: D.H., L.L.-M., O.A. Some limitations of the present study were the retro-378 Interpretation and analysis of data: D.H., A.V.-V., E.V.spective single-institution design that could not reflect 379 S., D.C.-F., G.C.-R., M.O. daily practice in other Latin American institutions, and 380 Preparation of manuscript: D.H., A.V.-V., L.L.-M. the presence of some bias in our results due to the na-381 Revision for important intellectual content: O.A., D.H., ture of the research. Even though, our institution is a 382 L.B-G. high-reference center with a complete representation of 383 Supervision: O.A., D.H., L.L.-M. the entire country, reflecting important real-world evi-384 dence. In addition, due to the retrospective design, we 385 could not evaluate the concordance between the tissue 386 Supplementary data and LB because most patients did not have available 387 tissue samples after progression. We recognize that the 388 The supplementary files are available to download standard procedure for patients with negative LB results 389 from http://dx.doi.org/10.3233/CBM-230124. is to perform tissue re-biopsy; however, in real-life, 390 tissue re-biopsy is performed in 26.1% of cases [47]. 391 Tissue confirmation after the first negative LB test was 392 References extremely low in our cohort because of the high propor-393 tion of patients who had more than one LB, and refused [1] The Global Cancer Observatory, Globocan 2020, International 394 Agency for Research on Cancer 419 (2020), 1-2. invasive procedures. We consider comparing real-world 395 A. Jemal, F. Bray, M.M. Center, J. Ferlay, E. Ward and D. [2] scenarios paramount in optimizing clinical practice, 396 Forman, Global cancer statistics, CA Cancer J Clin 61 (2011), considering that awareness and knowledge of liquid 69-90. 397 M.D.P. S S Birring, Symptoms and the early diagnosis of lung biopsy use are still limited. This study is of high value [3] 398 cancer, Thorax 60 (2005), 268-269. because in real life, most patients do not have access 399 T.S. Mok, Y.-L. Wu, S. Thongprasert, C.-H. Yang, D.-T. Chu, [4] to a biopsy at progression as demonstrated previously. 400 N. Saijo, P. Sunpaweravong, B. Han, B. Margono, Y. Ichinose, With the short turnaround of liquid biopsy, most pa-Y. Nishiwaki, Y. Ohe, J.-J. Yang, B. Chewaskulyong, H. Jiang, 401 E.L. Duffield, C.L. Watkins, A.A. Armour and M. Fukuoka, tients receive adequate treatment with good outcomes, 402 Gefitinib or carboplatin-paclitaxel in pulmonary adenocarciand the rest of the patients can be considered candidates 403 noma, N Engl J Med 361 (2009), 947-957. for clinical trials. [5] F. Passiglia, S. Rizzo, M. Di Maio, A. Galvano, G. Badala-404

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of Medicine 378 (2018), 113-125.

(2019), ix158.

[6]

[8]

[9]

[10]

[11]

[12]

13]

1984.

(2005), 786-792.

446

menti, A. Listì, L. Gulotta, M. Castiglia, V. Bazan, A. Russo

and F. Fulfaro, The diagnostic accuracy of circulating tumor

DNA for the detection of EGFR-T790M mutation in NSCLC:

A systematic review and meta-analysis, Sci Rep 8 (2018), 1-

O. Arrieta, L.A. Ramírez-Tirado, R. Báez-Saldaña, O. Peña-

Curiel, G. Soca-Chafre and E.O. Macedo-Perez, Different mu-

tation profiles and clinical characteristics among Hispanic pa-

tients with non-small cell lung cancer could explain the "His-

panic paradox", Lung Cancer 90(2) (2015 Nov), 161-6. doi:

10.1016/j.lungcan.2015.08.010. Epub 2015 Aug 22. PMID:

rodríguez, G. Bramuglia, O. Castillo-fernandez, M. Meyerson,

E. Amieva-rivera, A.D. Campos-parra, H. Carranza, J. Carlos,

G. De, Y. Powazniak, F. Aldaco-sarvide, C. Vargas, M. Trigo,

M. Magallanes-maciel, J. Otero, R. Sánchez-reyes and M.

Cuello, Updated Frequency of EGFR and KRAS mutations

in NonSmall-Cell Lung Cancer in Latin America The Latin-

American Consortium for the Investigation of Lung Cancer, J

J.-C. Soria, Y. Ohe, J. Vansteenkiste, T. Reungwetwattana, B.

Chewaskulyong, K.H. Lee, A. Dechaphunkul, F. Imamura,

N. Nogami, T. Kurata, I. Okamoto, C. Zhou, B.C. Cho, Y.

Cheng, E.K. Cho, P.J. Voon, D. Planchard, W.-C. Su, J.E.

Gray, S.-M. Lee, R. Hodge, M. Marotti, Y. Rukazenkov and

S.S. Ramalingam, Osimertinib in Untreated EGFR-Mutated

Advanced Non-Small-Cell Lung Cancer, New England Journal

Y.-L. Wu, T.S.K. Mok, J.-Y. Han, M.-J. Ahn, A. Delmonte,

S.S. Ramalingam, S.-W. Kim, F.A. Shepherd, J. Laskin, Y.

He, H. Akamatsu, W.S.M.E. Theelen, W.-C. Su, T. John, M.

Sebastian, H. Mann, M. Miranda, G. Laus, Y. Rukazenkov

and V. Papadimitrakopoulou, Overall survival (OS) from the

AURA3 phase III study: Osimertinib vs platinum-pemetrexed

(plt-pem) in patients (pts) with EGFR T790M advanced non-

small cell lung cancer (NSCLC) and progression on a prior

EGFR-tyrosine kinase inhibitor (TKI), Annals of Oncology 30

T.S. Mok, Y. Cheng, X. Zhou, K.H. Lee, K. Nakagawa, S.

Niho, M. Lee, R. Linke, R. Rosell, J. Corral, M.R. Migliorino,

A. Pluzanski, E.I. Sbar, T. Wang, J.L. White and Y.L. Wu,

Improvement in overall survival in a randomized study that

compared dacomitinib with gefitinib in patients with advanced

non-small-cell lung cancer and EGFR-activating mutations,

H. Yoshioka, M. Shimokawa, T. Seto, S. Morita, Y. Yatabe, I.

Okamoto, J. Tsurutani, M. Satouchi, T. Hirashima, S. Atagi, K.

Shibata, H. Saito, S. Toyooka, N. Yamamoto, K. Nakagawa and

T. Mitsudomi, Final overall survival results of WJTOG3405, a

randomized phase III trial comparing gefitinib versus cisplatin

with docetaxel as the first-line treatment for patients with stage

IIIB/IV or postoperative recurrent EGFR mutation-positive

non-small-cell lung, Annals of Oncology 30 (2019), 1978-

S. Kobayashi, T.J. Boggon, T. Dayaram, P.A. Jänne, O. Kocher,

M. Meyerson, B.E. Johnson, M.J. Eck, D.G. Tenen and B.

Halmos, EGFR mutation and resistance of non-small-cell lung

cancer to gefitinib, New England Journal of Medicine 352

H.A. Yu, M.E. Arcila, N. Rekhtman, C.S. Sima, M.F. Za-

kowski, W. Pao, M.G. Kris, V.A. Miller, M. Ladanyi and G.J.

Riely, Analysis of Tumor Specimens at the Time of Acquired

Resistance to EGFR-TKI Therapy in 155 Patients with EGFR-

Journal of Clinical Oncology 36 (2018), 2244–2250.

[7] O. Arrieta, F. Andrés, C. Martín, L. Más-lópez, L. Corrales-

Mutant Lung Cancers, Clinical Cancer Research 19 (2013),

Reguart, N. Karachaliou, H. Carranza, C. Vargas, J. Otero, P.

Archila, C. Martín, L. Corrales, M. Cuello, C. Ortiz, L.E. Pino.

R. Rosell and Z.L. Zatarain-Barrón, Acquired Resistance to

Erlotinib in EGFR Mutation-Positive Lung Adenocarcinoma

among Hispanics (CLICaP), Target Oncol 12 (2017), 513-523.

T. Spence, S. Perera, J. Weiss, S. Grenier, L. Ranich, F. Shep-

herd and T.L. Stockley, Clinical implementation of circulating

tumour DNA testing for EGFR T790M for detection of treat-

ment resistance in non-small cell lung cancer, J Clin Pathol 74

K. Koyama, S. Miura, S. Watanabe, S. Shoji, J. Koshio, Y.

Havashi, D. Ishikawa, K. Sato, T. Miyabayashi, M. Okajima.

T. Ota, T. Tanaka, N. Matsumoto, H. Kuriyama, T. Abe, K.

Nozaki, K. Ichikawa, R. Kondo, H. Tanaka and T. Kikuchi,

Observatiore-biopsyy of re-biopsy in EGFR-TKI-resistant pa-

tients with EGFR mutation-positive advanced NSCLC, Sci Rep

S.O. Takahisa Kawamura, Hirotsugu Kenmotsu*, Clinical Fac-

tors Predicting Detection of T790MRe-biopsyn in Re-biopsy

for EGFR-Mutant Non-Small Cell Lung Cancer, Clin Lung

Q.S.C. Chu, A. Agha, N. Devost, R.N. Walton, S. Ghosh and

C. Ho, Biopsy on progression in patients with egfr mutation-

feld, Are liquid biopsies a surrogate for tissue EGFR testing,

J. Luo, L. Shen and D. Zheng, Diagnostic value of circulating

free DNA for detecting EGFR mutation status in NSCLC: A

R. Zhang, B. Chen, X. Tong, Y. Wang, C. Wang, J. Jin, P.

Tian and W. Li, Diagnostic accuracy of droplet digital PCR

for detection of EGFR T790M mutation in circulating tumor

K.S. Thress, A. Markovets, J.C. Barrett, J. Chmielecki, S.B.

Goldberg, F.A. Shepherd, S. Vowler and G.R. Oxnard, Com-

plete clearance of plasma EGFR mutations as a predictor of

outcome on osimertinib in the AURA trial. Https://DoiOrg/

O'Connell, N. Feeney, S.L. Mach, P.A. Jänne and O. Geoffrey,

Prospective Validation of Rapid Plasma Genotyping for the

Detection of EGFR and KRAS Mutations in Advanced Lung

L. Arnold, V. Alexiadis, T. Watanaskul, V. Zarrabi, J. Poole

and V. Singh, Clinical validation of qPCR Target SelectorTM

assays using highly specific switch-blockers for rare mutation

L.Y. Chen, M.A. Molina-Vila, S.Y. Ruan, K.Y. Su, W.Y. Liao,

K.L. Yu, C.C. Ho, J.Y. Shih, C.J. Yu, J.C.H. Yang, R. Rosell

and P.C. Yang, Coexistence of EGFR T790M mutation and

common activating mutations in pretreatment non-small cell

lung cancer: A systematic review and meta-analysis, Lung

H. Liang, Z. Pan, W. Wang, C. Guo, D. Chen, J. Zhang, Y.

Zhang, S. Tang, J. He and W. Liang, The alteration of T790M

between 19 del and L858R in NSCLC in the course of EGFR-

TKIs therapy: A literature-based pooled analysis, J Thorac Dis

J. Remon, C. Caramella, C. Jovelet, L. Lacroix, A. Lawson, S.

101200/JCO20173515_suppl9018 35 (2017), 9018-9018.

[23] A.G. Sacher, C. Paweletz, S.E. Dahlberg, R.S. Alden, A.

Cancer, JAMA Oncol 2 (2016), 1014–1022

detection, J Clin Pathol 73 (2020), 648-655.

Cancer 94 (2016), 46–53.

10 (2018), 2311-2320.

positive advanced non-small-cell lung cancer - a canadian

experience, Current Oncology 27 (2020), 27-33.

Annals of Oncology 29 (2018), i38-i46.

[19] J.W. Goldman, Z.S. Noor, J. Remon, B. Besse and N. Rosen-

systematic review and meta-analysis, Sci Rep 4 (2014).

DNA, Cancer Manag Res 10 (2018), 1209-1218.

[14] A.F. Cardona, O. Arrieta, M.I. Zapata, L. Rojas, B. Wills, N.

9

510

511

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514

515

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517

518

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D. Heredia et al. / Liquid biopsy for EGFR resistance mutations

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[17]

[18]

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[21]

[22]

[24]

[25]

[26]

[27]

2240-2247

(2021), 91-97.

12 (2022), 8–15.

Cancer 11 (2010), 211-215.

- 502 503

- 508

574

575

576

577

578

579

581

584

602

603

604

605

D. Heredia et al. / Liquid biopsy for EGFR resistance mutations

- Smalley, K. Howarth, D. Gale, E. Green, V. Plagnol, N. Rosenfeld, D. Planchard, M.V. Bluthgen, A. Gazzah, C. Pannet, C. Nicotra, E. Auclin, J.C. Soria and B. Besse, Osimertinib benefit in EGFR-mutant NSCLC patients with T790M-mutation detected by circulating tumour DNA, Ann Oncol 28 (2017), 784-790.
- S. Mondaca, M. Offin, L. Borsu, M. Myers, S. Josyula, A. [28] 580 Makhnin, R. Shen, G.J. Riely, C.M. Rudin, M. Ladanyi, H.A. Yu, B.T. Li and M.E. Arcila, Lessons learned from routine, 582 targeted assessment of liquid biopsies for EGFR T790M re-583 sistance mutation in patients with EGFR mutant lung cancers, Acta Oncol 58 (2019), 1634. 585
- [29] M. Reck, K. Hagiwara, B. Han, S. Tjulandin, C. Grohé, T. 586 Yokoi, A. Morabito, S. Novello, E. Arriola, O. Molinier, R. 587 McCormack, M. Ratcliffe and N. Normanno, ctDNA Determi-588 589 nation of EGFR Mutation Status in European and Japanese Patients with Advanced NSCLC: The ASSESS Study, J Thorac 590 Oncol 11 (2016), 1682-1689. 591
- A. Chiang, A. Fernandes, M. Pavilack, J. Wu, F. Laliberté, [30] 592 M.S. Duh, N. Chehab and J. Subramanian, MA15.11 Real 593 594 World Biomarker Testing and Treatment Patterns in Patients with Advanced NSCLC Receiving EGFR-TKIs, Journal of 595 Thoracic Oncology 13 (2018), S410–S411. 596
- [31] G.R. Oxnard, K.S. Thress, R.S. Alden, R. Lawrance, C.P. 597 Paweletz, M. Cantarini, J.C.H. Yang, J.C. Barrett and P.A. 598 Jänne, Association Between Plasma Genotyping and Outcomes 599 600 of Treatment With Osimertinib (AZD9291) in Advanced Non-Small-Cell Lung Cancer, J Clin Oncol 34 (2016), 3375-3382. 601
 - [32] M. Qiu, J. Wang, Y. Xu, X. Ding, M. Li, F. Jiang, L. Xu and R. Yin, Circulating tumor DNA is effective for the detection of EGFR mutation in non-small cell lung cancer: A meta-analysis, Cancer Epidemiol Biomarkers Prev 24 (2015), 206–212.
- [33] A. Passaro, N. Leighl, F. Blackhall, S. Popat, K. Kerr, M.J 606 Ahn, M.E. Arcila, O. Arrieta, D. Planchard, F. de Marinis 607 A.M. Dingemans, R. Dziadziuszko, C. Faivre-Finn, J. Feld-608 man, E. Felip, G. Curigliano, R. Herbst, P.A. Jänne, T. John, 609 T. Mitsudomi, T. Mok, N. Normanno, L. Paz-Ares, S. Rama-610 lingam, L. Sequist, J. Vansteenkiste, I.I. Wistuba, J. Wolf, Y.L. 611 Wu, S.R. Yang, J.C.H. Yang, Y. Yatabe, G. Pentheroudakis and 612 613 S. Peters, ESMO expert consensus statements on the management of EGFR mutant non-small-cell lung cancer, Ann Oncol 614 33 (2022) 466-487 615
- C. Rolfo, P. Mack, G.V. Scagliotti, C. Aggarwal, M.E. Arcila, [34] 616 F. Barlesi, T. Bivona, M. Diehn, C. Dive, R. Dziadziuszko, N. 617 Leighl, U. Malapelle, T. Mok, N. Peled, L.E. Raez, L. Sequist, L. Sholl, C. Swanton, C. Abbosh, D. Tan, H. Wakelee, I. Wis-618 619 tuba, R. Bunn, J. Freeman-Daily, M. Wynes, C. Belani, T. Mit-620 sudomi and D. Gandara, Liquid Biopsy for Advanced NSCLC: 621 A consensus statement from the international association for 622 the study of lung cancer. Journal of Thoracic Oncology 16 623 (2021), 1647-1662. 624
- [35] R. Minari, G. Mazzaschi, P. Bordi, L. Gnetti, G. Alberti, A. 625 Altimari, E. Gruppioni, F. Sperandi, C. Parisi, G. Guaitoli, 626 S. Bettelli, L. Longo, F. Bertolini, M. Pagano, C. Bonelli, E. 627 Tagliavini, D. Nicoli, A. Ubiali, A. Zangrandi, S. Trubini, M. 628 Proietto, M. Fiorentino and M. Tiseo, Detection of EGFR-629 Activating and T790M Mutations Using Liquid Biopsy in 630 Patients With EGFR-Mutated Non-Small-Cell Lung Cancer 631 Whose Disease Has Progressed During Treatment With First-632 and Second-Generation Tyrosine Kinase Inhibitors: A Multi-633 634 center Real-Life Retrospective Study, Clin Lung Cancer 21 (2020), e464-e473. 635
- A. Dal Maso, P. Del Bianco, F. Cortiula, G. Nardo, E. Zulato, [36] 636 L. Bonanno, A. Follador, G. De Maglio, G. Pasello and S. In-637

draccolo, EGFR T790M testing through repeated liquid biopsy over time: A real-world multicentric retrospective experience, J Thorac Dis 14 (2022), 3364-3375.

638

639

640

641

642

643

644

645

646

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648

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652

653

654

655

656

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676

677

678

679

680

681

682

683

684

685

686

687

688

689

690

691

692

693

694

695

696

- F. Passiglia, S. Rizzo, C. Rolfo, A. Galvano, E. Bronte, L [37] Incorvaia, A. Listi, N. Barraco, M. Castiglia, V. Calo, V. Bazan and A. Russo, Metastatic Site Location Influences the Diagnostic Accuracy of ctDNA EGFR-Mutation Testing in NSCLC Patients: A Pooled Analysis, Curr Cancer Drug Targets 18 (2018), 697-705.
- H. Al-Halabi, K. Sayegh, S.R. Digamurthy, A. Niemierko, Z. [38] Piotrowska, H. Willers and L. V. Sequist, Pattern of Failure Analysis in Metastatic EGFR-Mutant Lung Cancer Treated with Tyrosine Kinase Inhibitors to Identify Candidates for Consolidation Stereotactic Body Radiation Therapy, J Thorac Oncol 10 (2015), 1601-1607.
- [39] S.H. Patel, A. Rimner, A. Foster, Z. Zhang, K.M. Woo, H.A. Yu, G.J. Riely and A.J. Wu, Lung cancer, 108 (2017), 109-114.
- [40] P. Bordi, M. Del Re, R. Minari, E. Rofi, S. Buti, G. Restante, A. Squadrilli, S. Crucitta, C. Casartelli, L. Gnetti, C. Azzoni, L. Bottarelli, I. Petrini, A. Cosenza, L. Ferri, E. Rapacchi, R. Danesi and M. Tiseo, From the beginning to resistance: Study of plasma monitoring and resistance mechanisms in a cohort of patients treated with osimertinib for advanced T790M-positive NSCLC, Lung Cancer 131 (2019), 78-85.
- [41] A.M. Newman, S.V. Bratman, J. To, J.F. Wynne, N.C.W. Eclov, L.A. Modlin, C.L. Liu, J.W. Neal, H.A. Wakelee, R.E. Merritt, J.B. Shrager, B.W. Loo, A.A. Alizadeh and M. Diehn, An ultrasensitive method for quantitating circulating tumor DNA with broad patient coverage, Nat Med 20 (2014), 548-554.
- 42] G.R. Oxnard, M.E. Arcila, C.S. Sima, G.J. Riely, J. Chmielecki, M.G. Kris, W. Pao, M. Ladanyi and V.A. Miller, Acquired resistance to EGFR tyrosine kinase inhibitors in EGFRmutant lung cancer: distinct natural history of patients with tumors harboring the T790M mutation. Clin Cancer Res 17 (2011), 1616-1622.
- [43] J. Remon, B. Besse, S.P. Aix, A. Callejo, K. Al-Rabi, R. Bernabe, L. Greillier, M. Majem, N. Reguart, I. Monnet, S. Cousin, P. Garrido, G. Robinet, R. Garcia Campelo, A. Madroszyk, J. Mazières, H. Curcio, B. Wasag, Y. Pretzenbacher, B. Fournier, A.M.C. Dingemans and R. Dziadziuszko, Osiertinib treatment based on plasma T790M monitoring in patients with EGFRmutant non-small-cell lung cancer (NSCLC): EORTC Lung Cancer Group 1613 APPLE phase II randomized clinical trial, Ann Oncol 34 (2023), 468-476.
- [44] V.A. Papadimitrakopoulou, J.Y. Han, M.J. Ahn, S.S. Ramalingam, A. Delmonte, T.C. Hsia, J. Laskin, S.W. Kim, Y. He, C.M. Tsai, T. Hida, M. Maemondo, T. Kato, S. Jenkins, S. Patel, X. Huang, G. Laus, A. Markovets, K.S. Thress, Y.L. Wu and T. Mok, Epidermal growth factor receptor mutation analvsis in tissue and plasma from the AURA3 trial: Osimertinib versus platinum-pemetrexed for T790M mutation-positive advanced non-small cell lung cancer, Cancer 126 (2020), 373-380
- [45] T.S. Mok, Y.-L. Wu, M.-J. Ahn, M.C. Garassino, H.R. Kim, S.S. Ramalingam, F.A. Shepherd, Y. He, H. Akamatsu, W.S.M.E. Theelen, C.K. Lee, M. Sebastian, A. Templeton, H. Mann, M. Marotti, S. Ghiorghiu and V.A. Papadimitrakopoulou, Osimertinib or Platinum-Pemetrexed in EGFR T790M-Positive Lung Cancer, N Engl J Med 376 (2017), 629-640.
- [46] D. Planchard, S. Popat, K. Kerr, S. Novello, E.F. Smit, C. 698 Faivre-Finn, T.S. Mok, M. Reck, P.E. Van Schil, M.D. Hell-699 mann and S. Peters, Corrigendum: Metastatic non-small cell 700 lung cancer: ESMO Clinical Practice Guidelines for diagno-701

708

709

710

D. Heredia et al. / Liquid biopsy for EGFR resistance mutations 11 sis, treatment and follow-up (Annals of Oncology (2018) 29 I. Chmielewska, R. Kieszko, M. ójcik-Superczyńska, M. 702 (iv192-iv237) DOI: 10.1093/annonc/mdy275), Annals of On-Szczyek, T. Jankowski and J. Milanowski, The efficacy of 703 T790M mutation testing in liquid biopsy - Real clinic data, cology 30 (2019), 863-870. 704 [47] P. Krawczyk, L. Grzycka-Kowalczyk, J. Błach, K. Reszka, PLoS One 17 (2022). 705 706 corrected proof version